



Laboratori Nazionali di Legnaro – INFN

Work Package 1: activities 2025

V. Di Marco, D. Maniglio

June 18th, 2025

Brief review of WP1 activities in 2025:

I. (Di Marco)

II. (Maniglio)

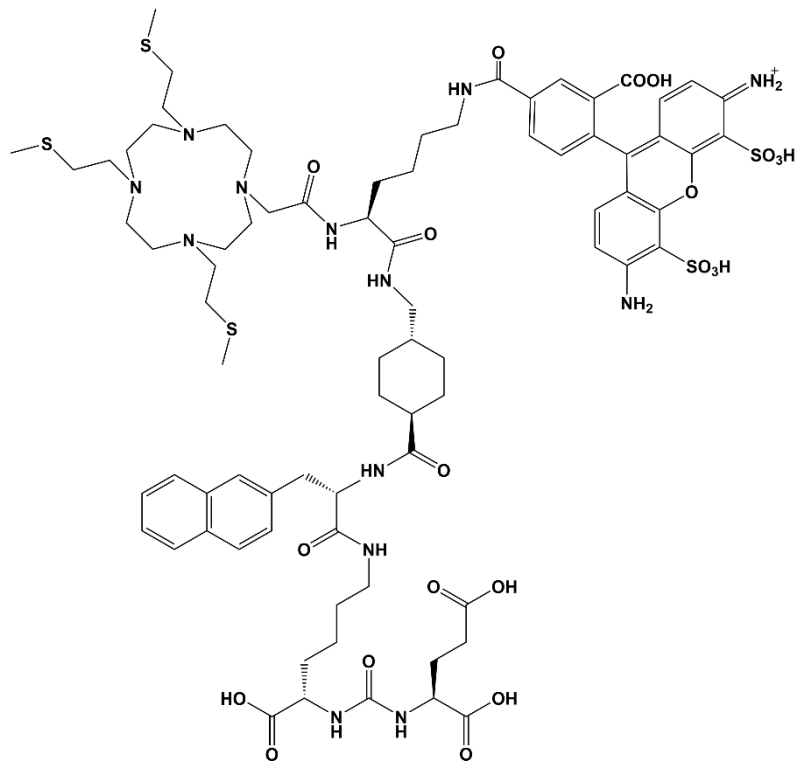
		Done
MS1.0	Optimization of Ag-111 production and Ag/Pa separation	
MS1.1	Routine production of Ag-111 at the LENA facilities, purification and quality control	
MS1.2	Synthesis of improved stable Ag chelators and radiochemical characterization	
MS1.3	Development of the macromolecule for active targeting	
MS1.4	Bioengineering of 3D scaffolds for in vitro tissue mimicking	

This talk (Maniglio)

This talk (Di Marco)

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Cooperation agreement in due course with
A.C. Eder, University of Freiburg, Department of Nuclear
Medicine, Germany



Chemical Formula: $C_{79}H_{108}N_{13}O_{21}S_5$

Exact Mass: 1734,64

Molecular Weight: 1736,10

Expected shipping date
from Germany: July 2025

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Just arrived (17/06/2025): results of the ICP-MS analysis of $^{111}\text{Ag}/^{110}\text{Pd}$ separations

Resin/sample	Pd (ppb) $^{111}\text{Ag}/^{110}\text{Pd}$
LN1-1	4765
LN1-2	2230
LN1-3	3300
LN1-4	43
LN2-5	481
LN2-6	292
LN2-7	385
LN2-8	23
TK200-9	3
TK200-10	1.5
TK200-11	11
TK200-12	0.26

Data are in good agreement with previous analyses (March, 2025) performed on $^{\text{nat}}\text{Ag}/^{\text{nat}}\text{Pd}$ separations

Pd (ppb) $^{\text{nat}}\text{Ag}/^{\text{nat}}\text{Pd}$
2.6
2.4
0.5
2.1

Month	Starting date	#	Activity [MBq]	#	Activity [mCi]	Experiment
January	27/01/2025	55,5		1,5		SPECT imaging with phantoms
February	10/02/2025	20		0,5		Vitro (UMR-106 foci)
March	31/03/2025	60		1,6		Vitro (MDA-MB-231 uptake and clonogenic assay)
April						
May	12/05/2025	18		0,5		Vitro (LNCaP clonogenic assay) + γ imaging
June	16/06/2025	60		1,6		Vitro (MDA-MB-231 foci assay)
July	30/06/2025	18		0,5		Vitro (LNCaP foci) + γ and β imaging
August						
September		18		0,5		Vitro 3D (LNCaP) + γ and β imaging
October						Imaging test with phantoms (Ag-111 vs Lu-177)
November		18		0,5		Vitro + γ and β imaging (extra tests)
December		60		1,6		Vitro (extra tests)

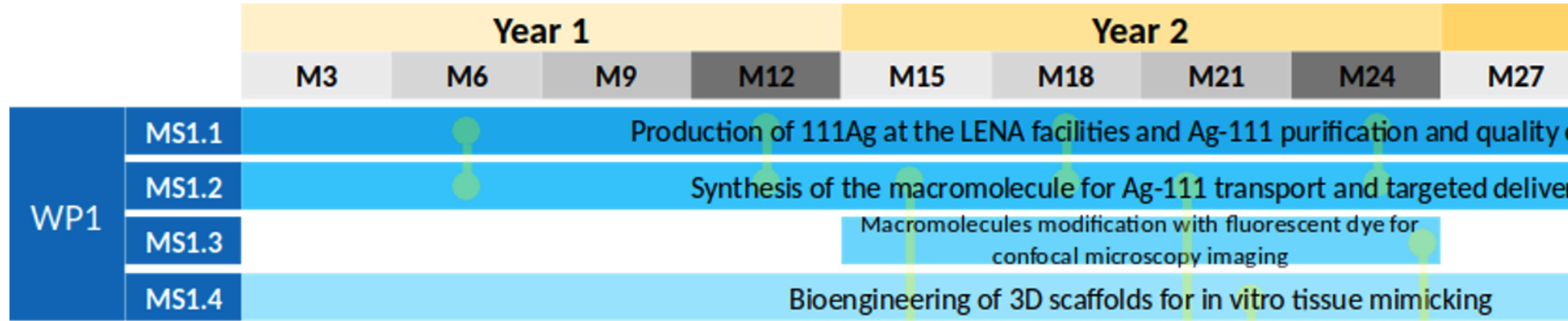


Advancement in the creation of 2.5D constructs

Annual meeting of WP1

September 13rd, 2024

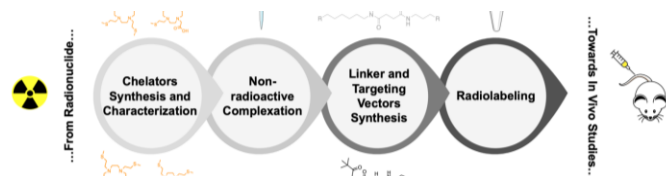
Planned Activities





1-2 - Radiopharmaceutical

Produced radionuclides must be inserted in a **macromolecular context** able to bind and transport them into the tumor tissue where they should **selectively** recognize and associate with cancer cells. The resulting research involves **studying chelators, spacers and targeting agents** to develop a **stable radiopharmaceutical** for ^{111}Ag complexation. In addition, **radiochemistry experiments** focused on the dissolution of the irradiated target where ^{111}Ag is obtained are required to optimize the radioisotopic purification.

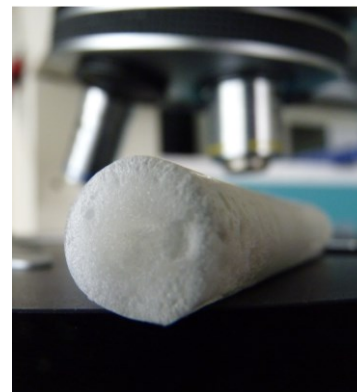


Radiopharmaceutical design & development framework

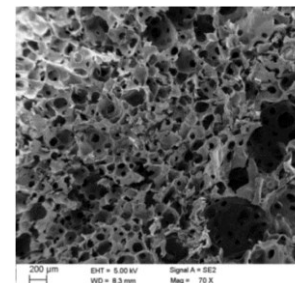


1-3 - Bioengineering

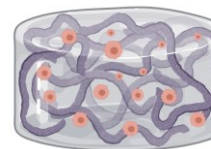
One of the goals of the new ISOLPHARM project is to extend this kind of studies to targeted radionuclide therapy and to more realistic **dynamic cell cultures** in 3D tissue-mimicking scaffolds, employing the bioengineering expertise of the collaboration.



3D printed scaffold.

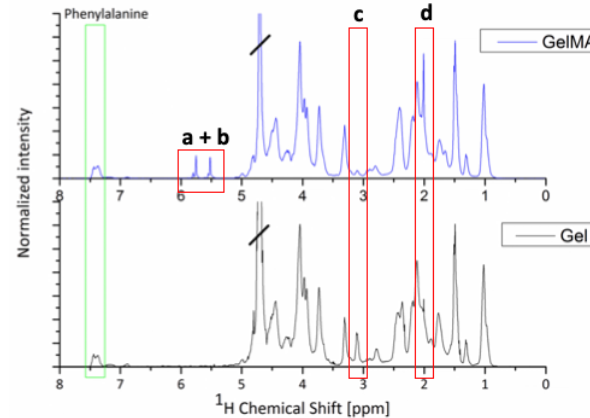
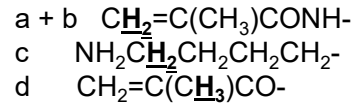
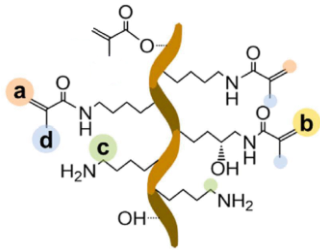
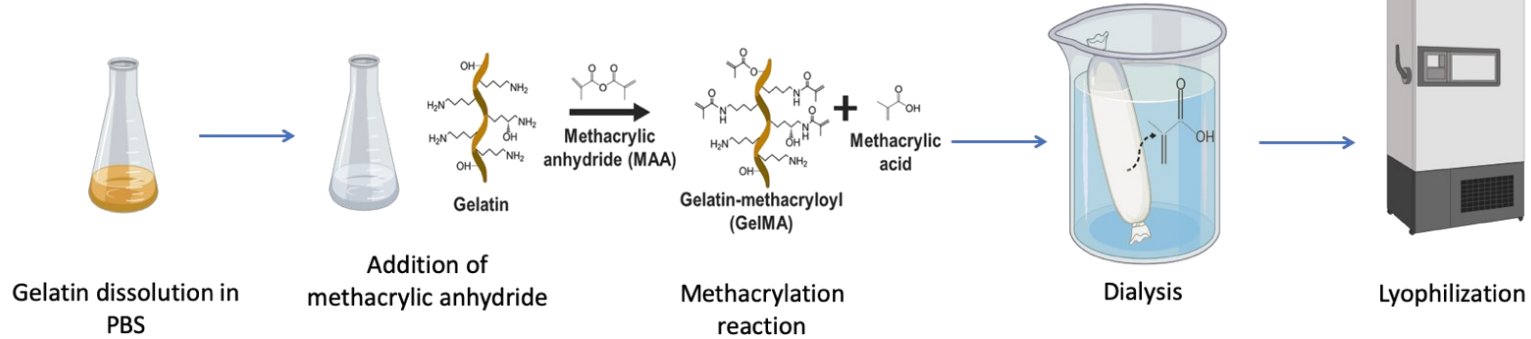


Scaffold porosity as detected by SEM.



Natural polymer hydrogel embedding cells and supporting them viable and functional

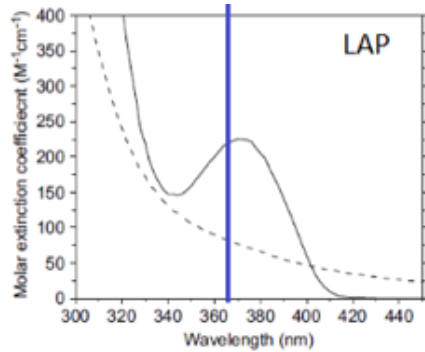
The materials: base bioink preparation



New protocols
experimented by Daniel
10 days \rightarrow 3 days

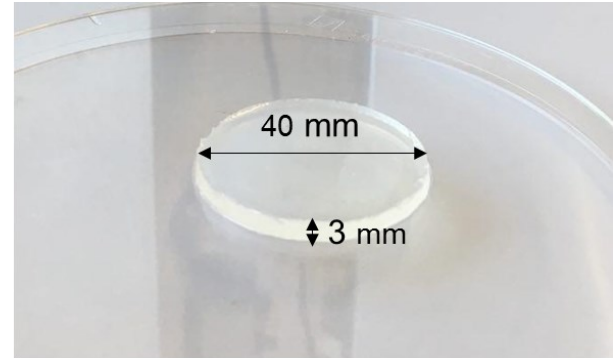
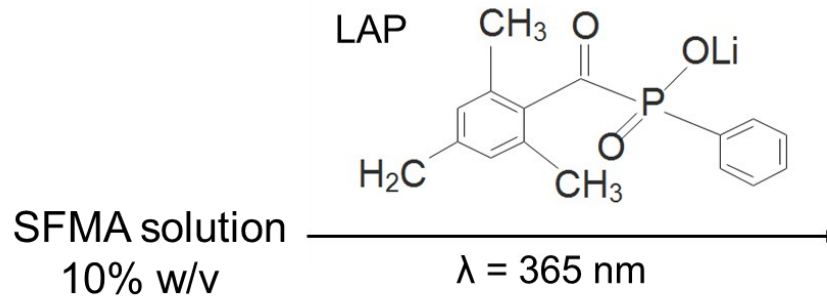
$$\text{DoF}_{\text{lysine}} \% = \left(1 - \frac{\int \text{lysine GelMA}}{\int \text{lysine unmodified gelatin}} \right) * 100$$

DoF* = 70%



Lithium Phenyl-2,4,6-trimethylbenzoylphosphinate (LAP)

- Low cytotoxicity
- High solubility in water
- High absorption at 365 nm ($\epsilon = 218 M^{-1}cm^{-1}$)



How to turn bioink to a 3D scaffold?

Matches Photolithography and SLA advantages:

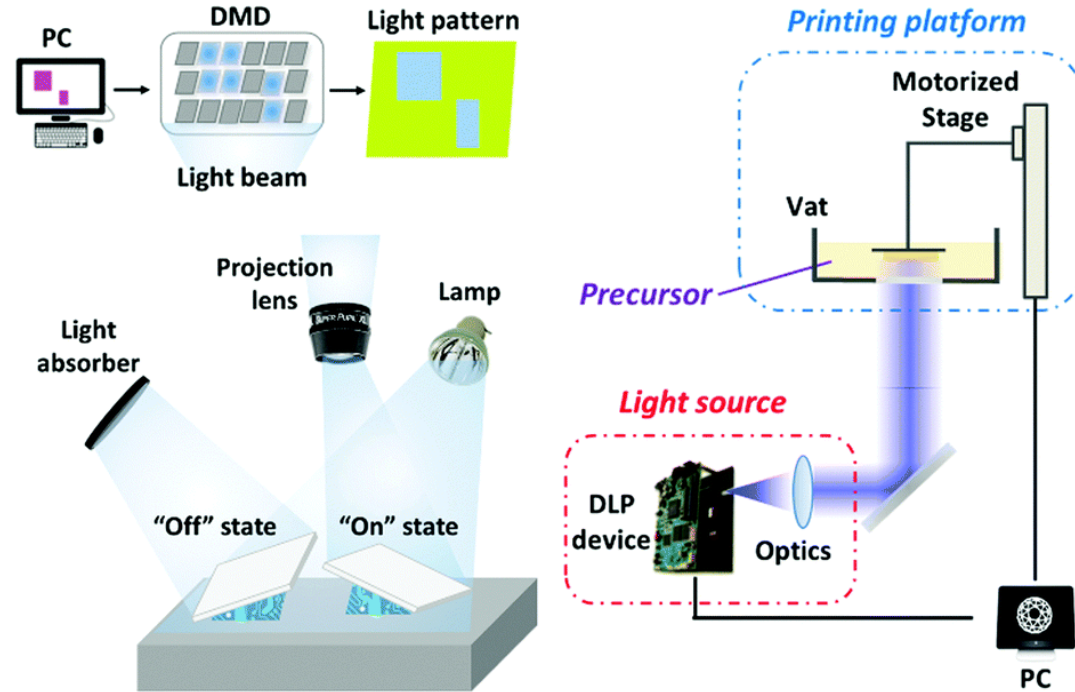
- **Maskless**
- **Each layer is printed at once**

Working principle:

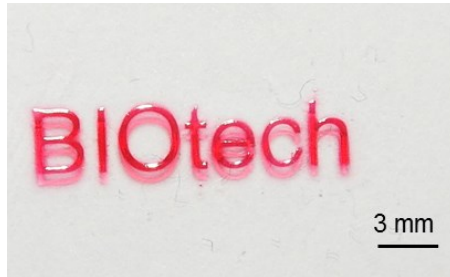
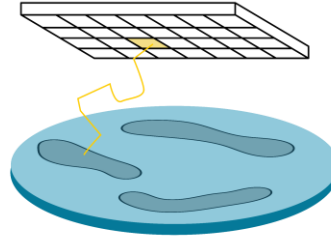
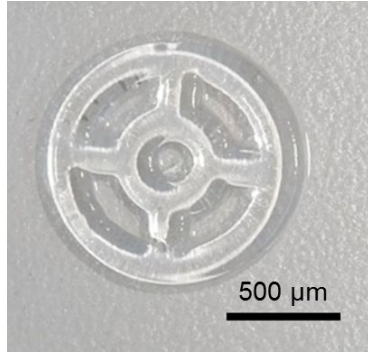
- Projector with LED source for illumination
- DMD (Digital Micromirror Device): digital programmable photomask to control the crosslinkable area
- No CAD modeling but any raster image

Biomaterial requirements:

- Photopolymerizable
- Photoinitiator with low toxicity

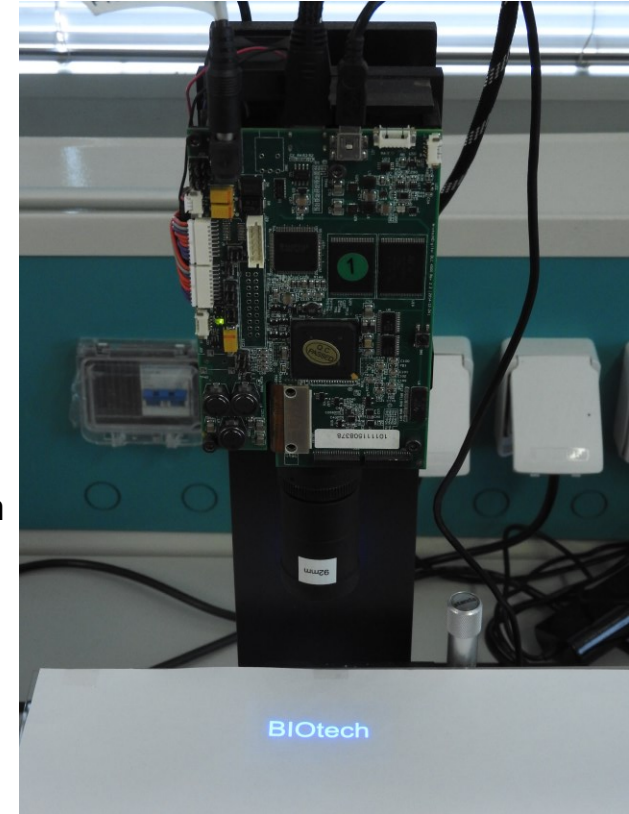


Complex patterns with DLP



It is optimal for 2D 1/2 scaffolds design for Beta imaging

Staining used: Betanin

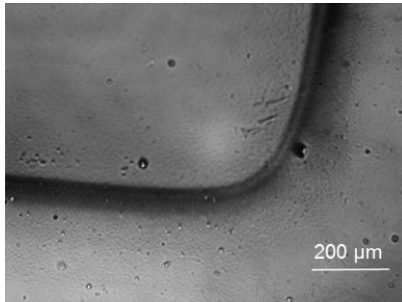
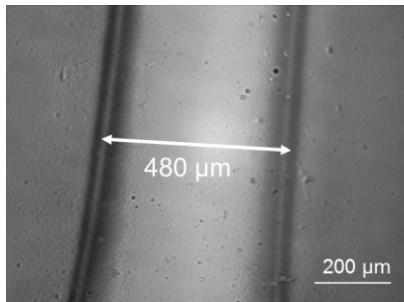
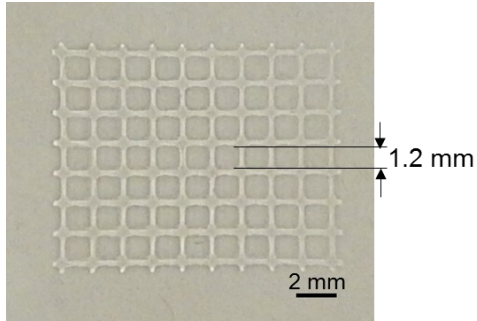


Printed meshes with DLP

Expected dimensions:

Lines = 400 μm

Holes = 1200 μm

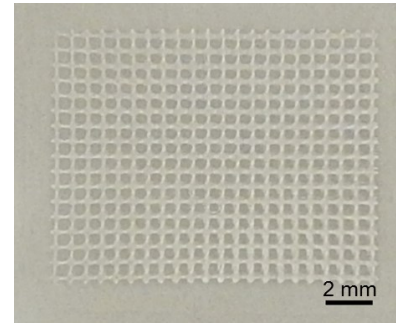
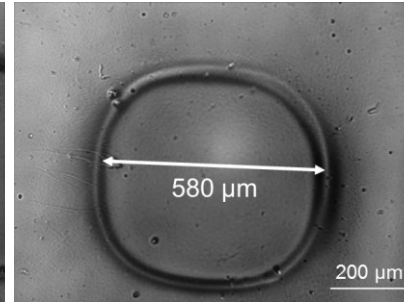
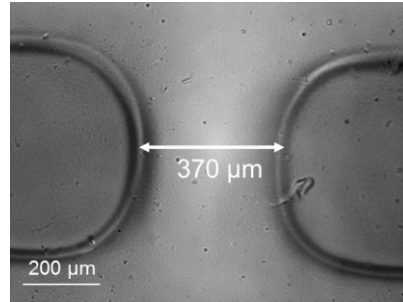


Lower limit of printability

Expected dimensions:

Lines = 240 μm

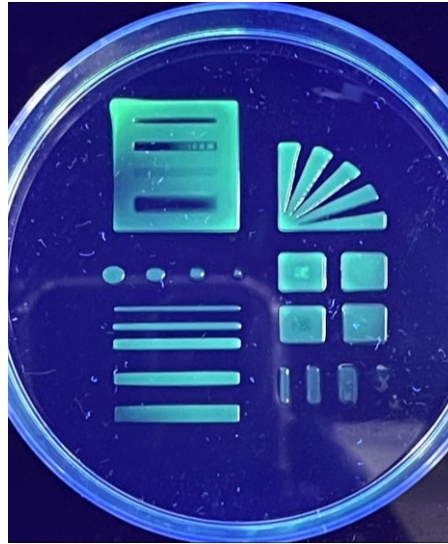
Holes = 480 μm



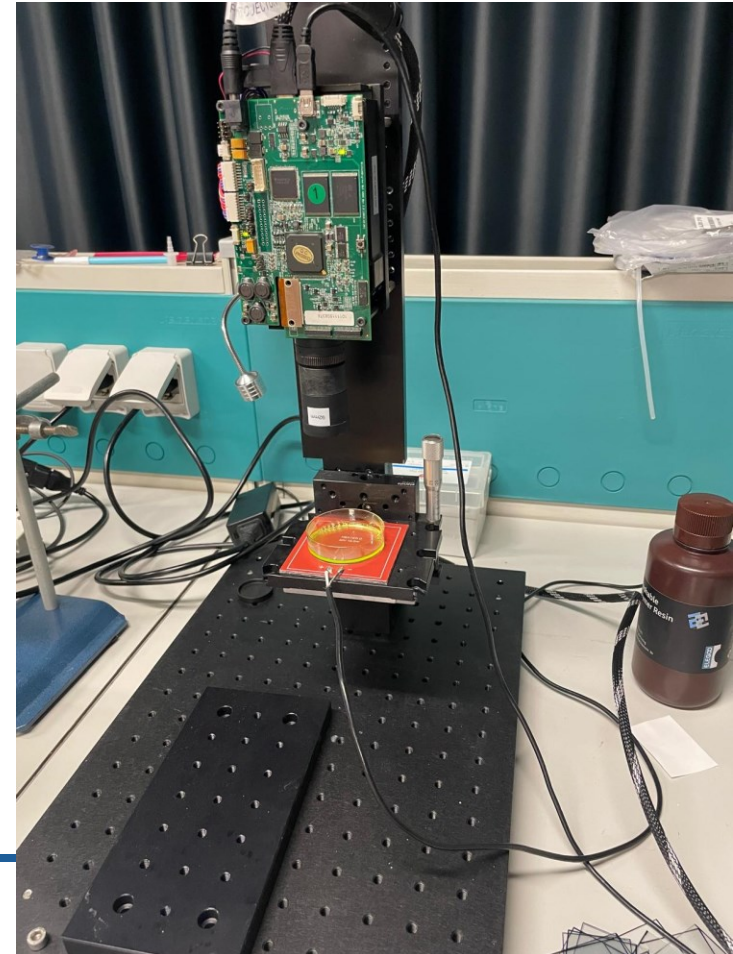
Geometries
suitable to 2D β -
imaging detector



Assessing large image printing

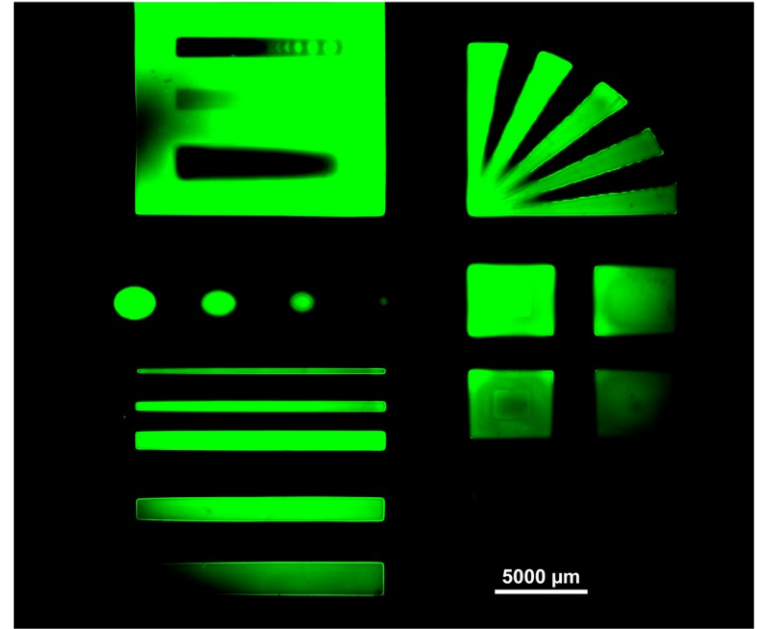


GelMA 20% + fluorescein dye + 1% LAP

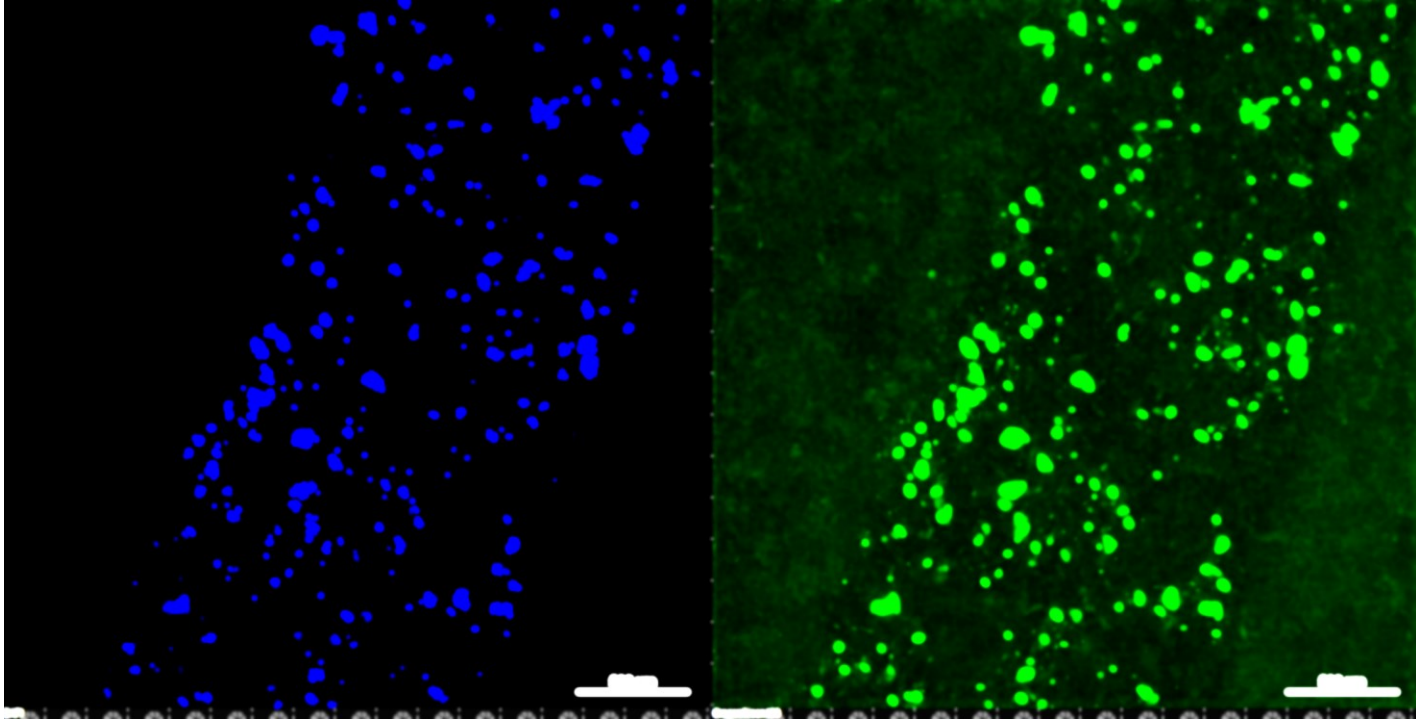




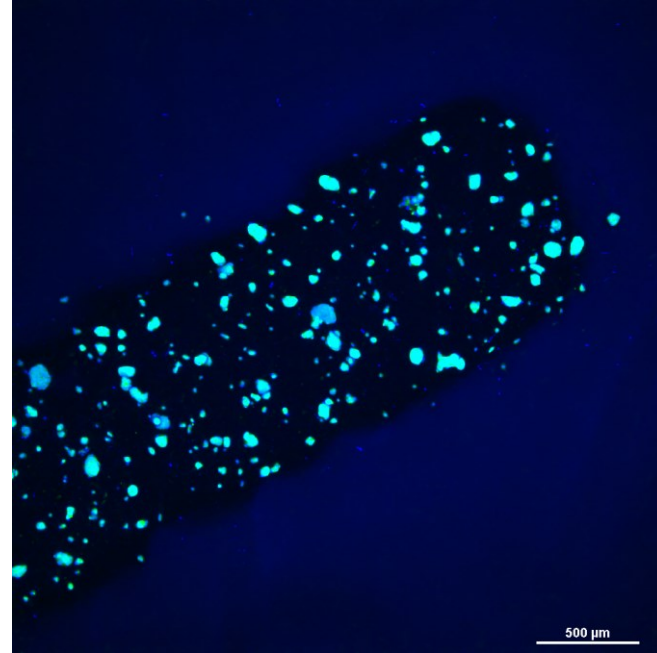
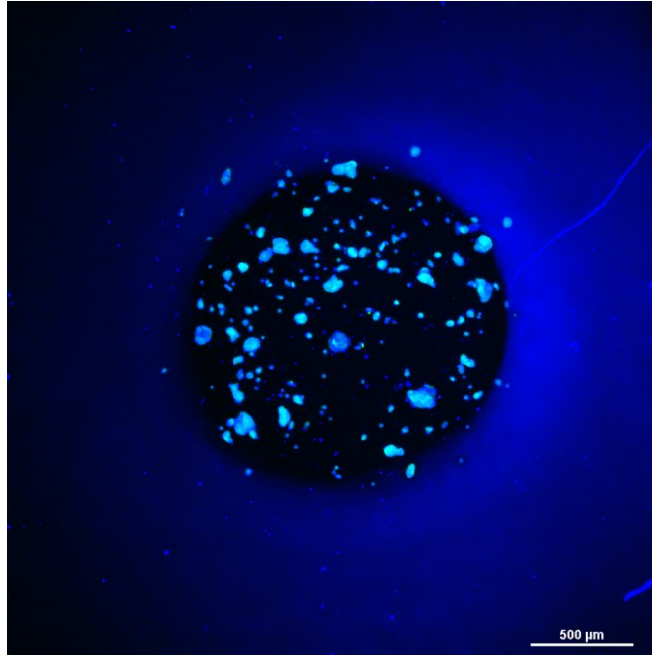
UV Transilluminator



Confocal microscopy



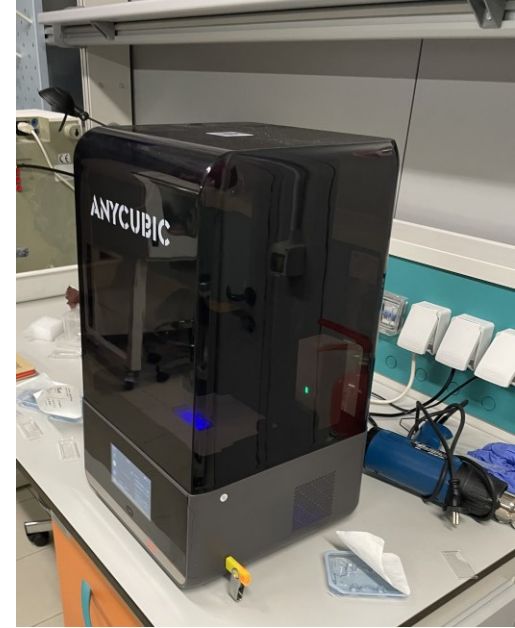
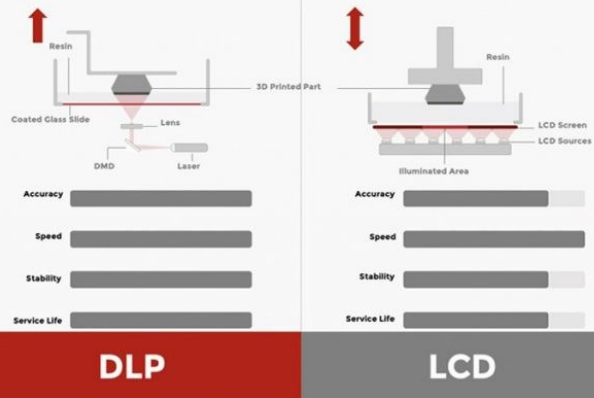
Stripes and cells day 1



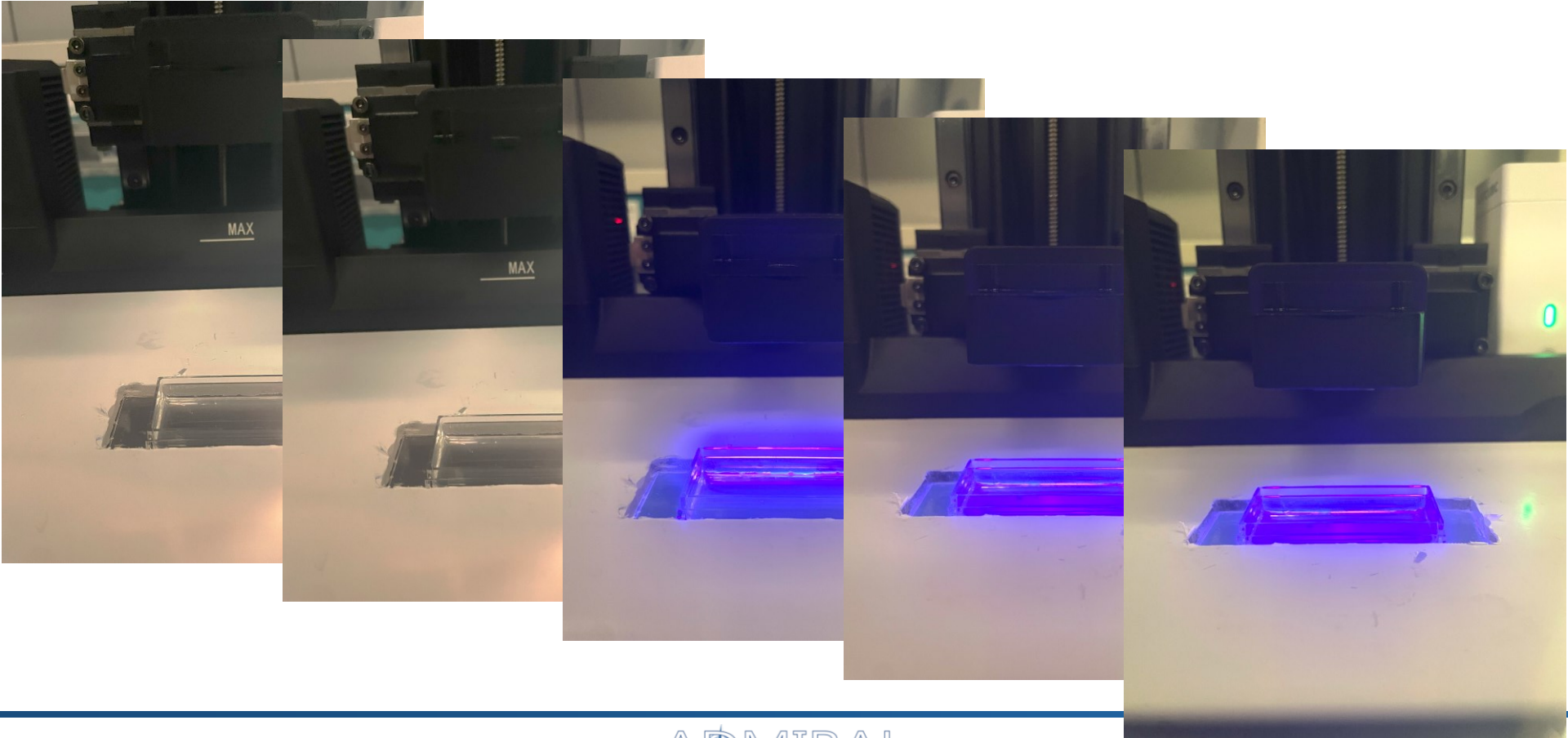
- Finding optimal projection support
(sterile – thin interlayer material – transportable – size compatible with gamma camera)
- Adapting twin commercial 3D printers for 2D^{1/2} bioprinting
to feed WP2 activities



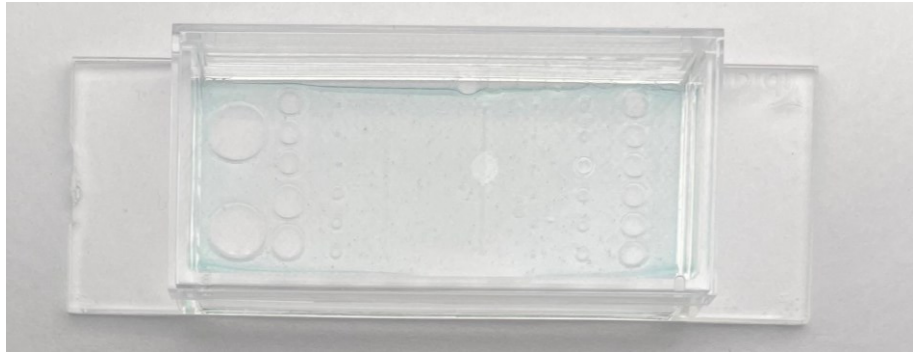
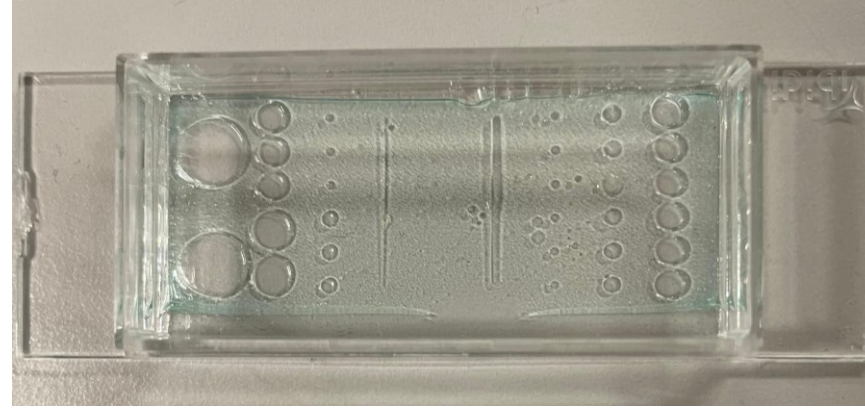
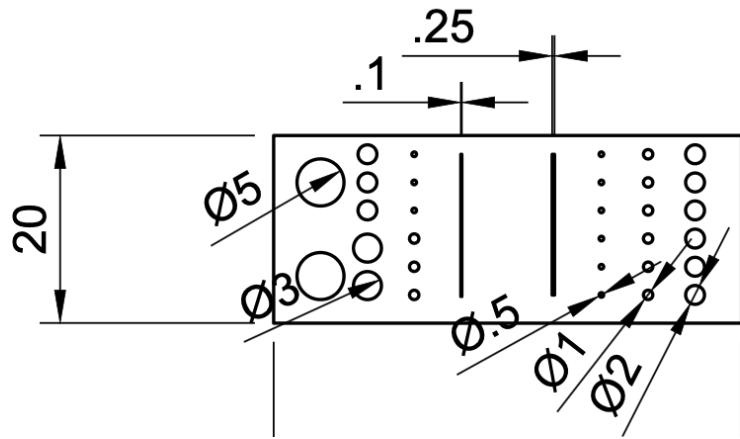
- LCD projector technology (not DLP)
- 405 nm wavelength, compatible with LAP photoinitiator
- Higher resolution for 400 €
- Needs modification



The printing process



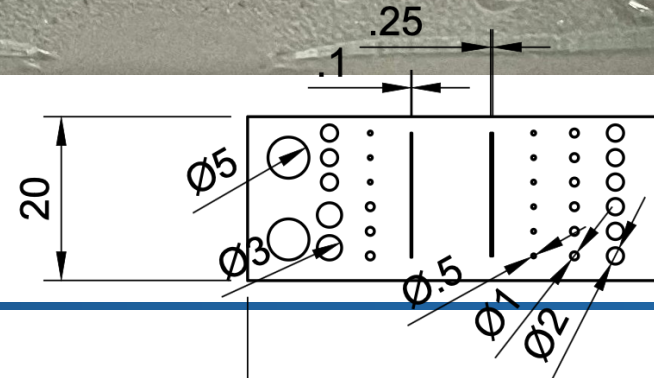
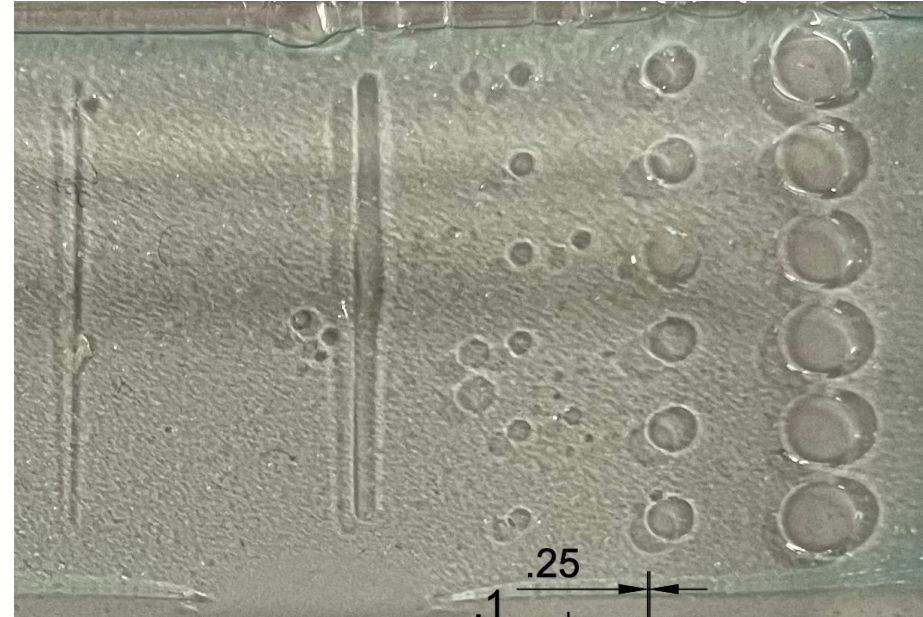
Resulting printed scaffolds



High aspect ratio structures are difficult to be printed
(0.2 mm limit)

The correct insertion of cells is reasonably made by pipette

There are also some considerations on the volume to be complemented



Thank you