



**BEYOND VISION:** *Physics meets AI*



ALMA MATER STUDIORUM  
UNIVERSITÀ DI BOLOGNA



Istituto Nazionale di Fisica Nucleare

# Optimizing Deep Learning Models for Cell Recognition in Fluorescence Microscopy: **the Impact of Loss Functions on Performance and Generalization**

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# Outline



Why is this relevant?



Background & Challenges



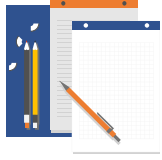
Loss functions



Performance metrics

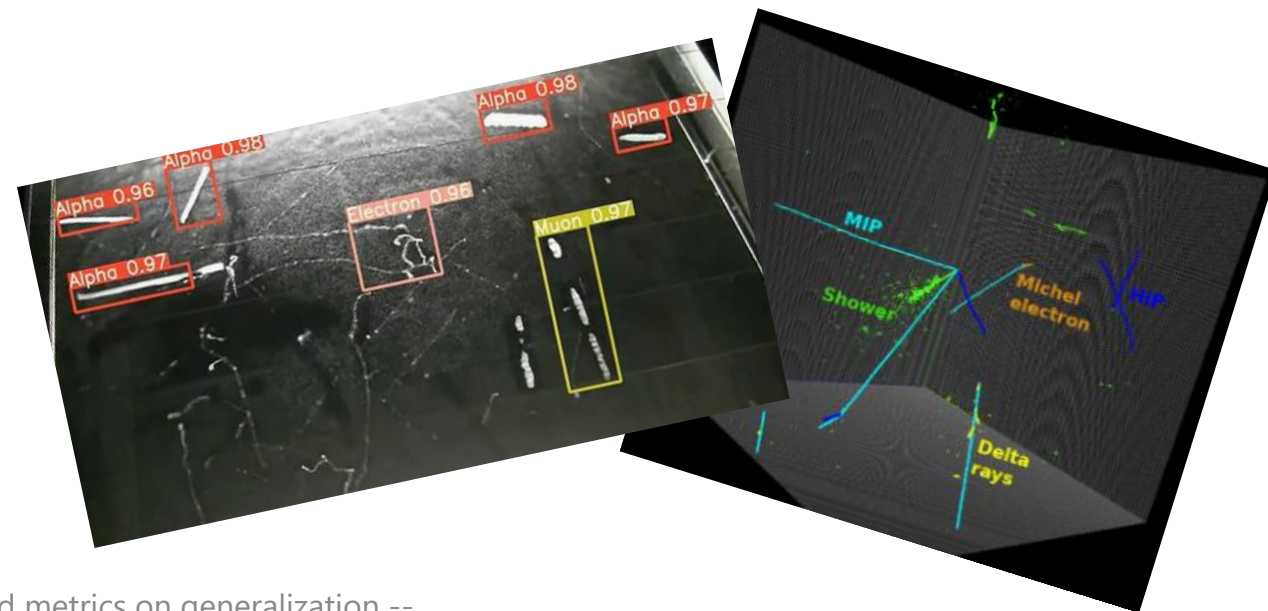


Results & Conclusions



# Why is this relevant?

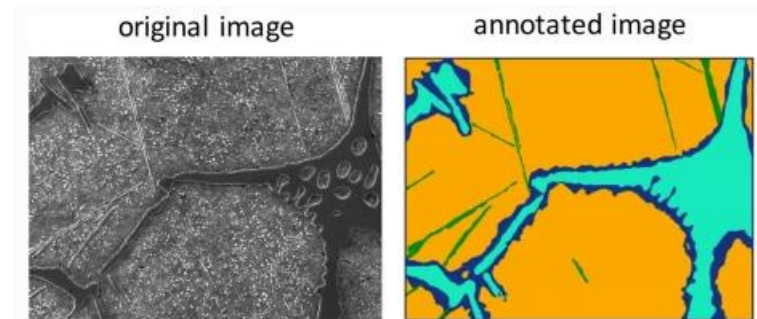
- Object «recognition» is a popular learning task
  - Segmentation, detection and counting objectives
- Applications also in physics
  - 3D semantic segmentation task on simulated LArTPC samples [1]
  - Particle identification in cloud chamber

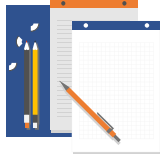




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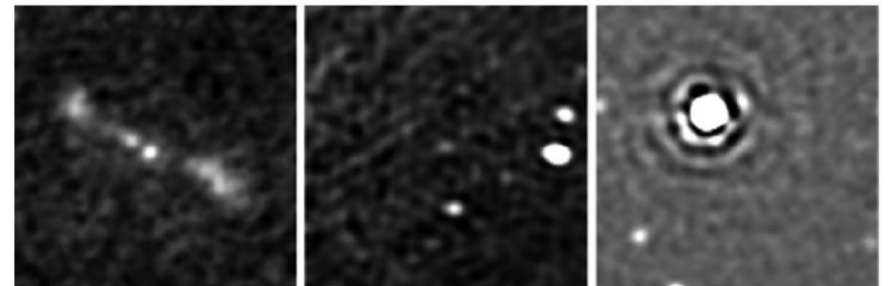
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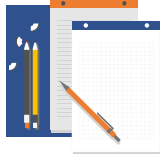


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  - Semantic Segmentation of Radio-Astronomical Images [3]

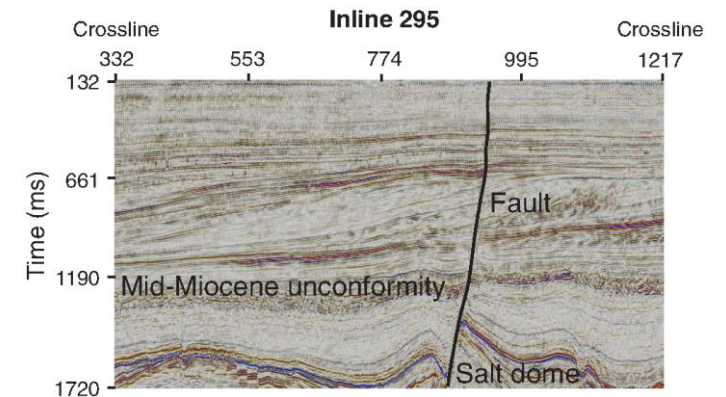


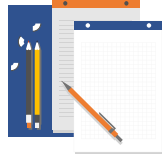
Examples of (left) galaxies, (center) sources and (right) sidelobes.



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  - Seismic facies interpretation [4]





# Why is this relevant?

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- Object «recognition» is a popular learning task
  - Segmentation, detection and counting objectives
- Applications also in physics
- Different applications typically share **similar challenges**
- **Aim**
  - Investigate loss effectiveness
  - Explore and compare several evaluation strategies



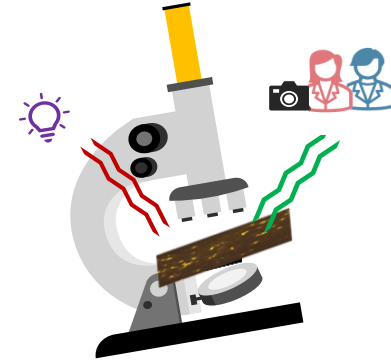
# Background

[6] Morelli, R., et al.: *Automating cell counting in fluorescent microscopy through deep learning with c-ResUnet*. Scientific Reports 11(1), 22920 (2021).

[7] Clissa, L., et al.: *Fluorescent neuronal cells v2: Multi-task, multi-format annotations for deep learning in microscopy*. arXiv preprint (in review at Scientific Data) (2023)

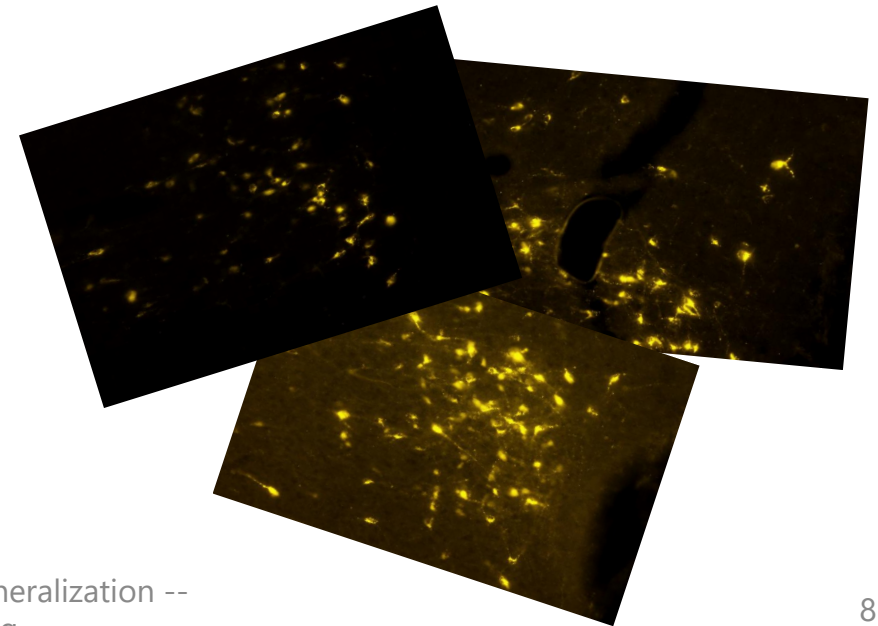
## Fluorescent Microscopy

- Physics-based imaging technique
- Exploits light absorption/emission properties
- Used to mark/tag/stain biological compounds

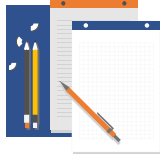


## Applications

- Very popular in life science
- Torpor onset [5, 6, 7]
- Cytoplasmatic neuronal structures
- Variability in shape, size and color hue
- **Goal:** count stained structures







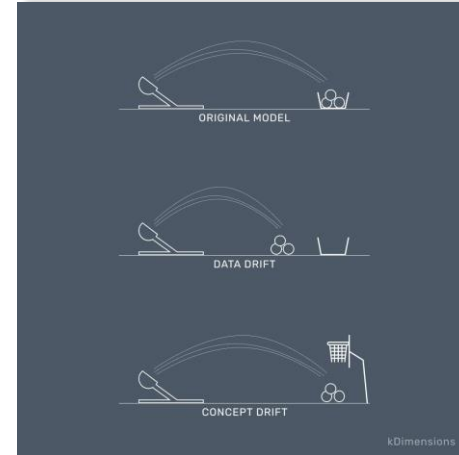
# Problem



## Manual processing

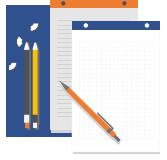
- Time-consuming
- Error-prone
- Subjectivity of borderline cases

Visual by [kDimensions](#)

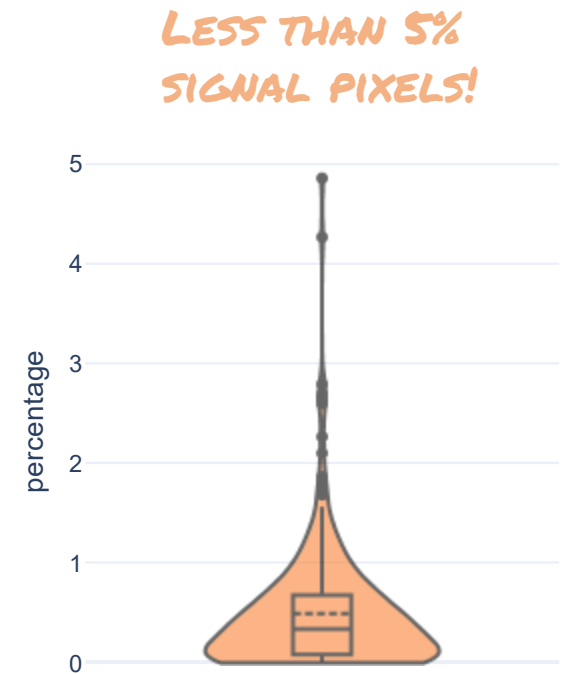
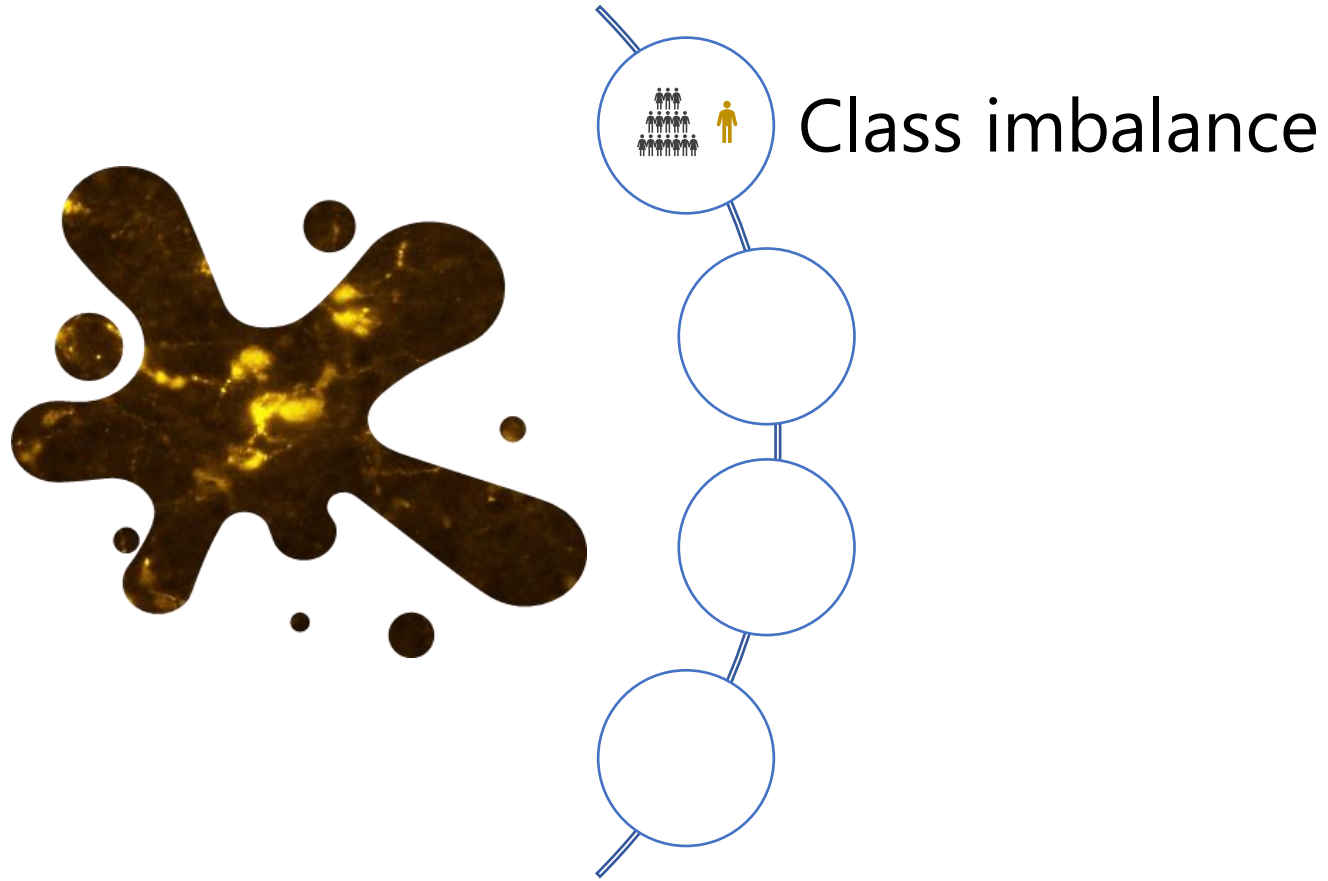


## Hard to adapt Deep Learning solutions

- Domain shift
- Few in-domain annotated datasets
- How to train? How to evaluate?

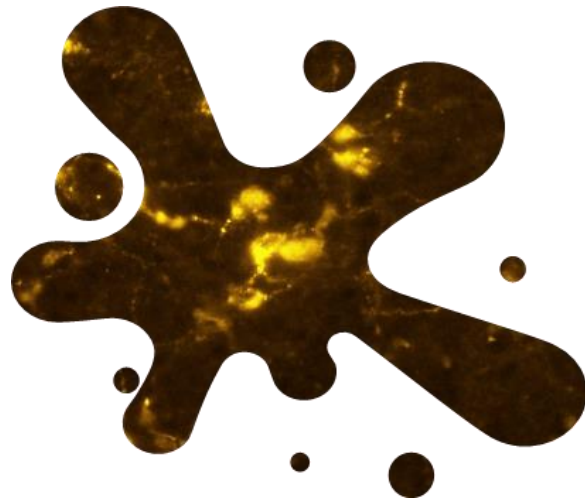


# Challenges

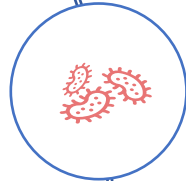




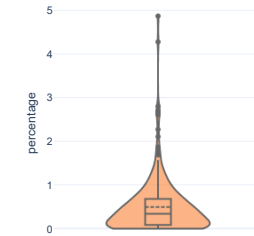
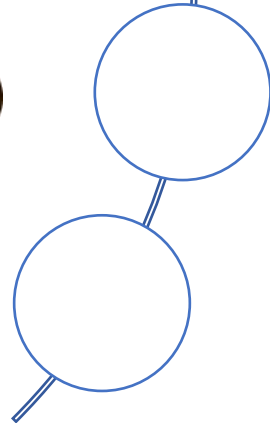
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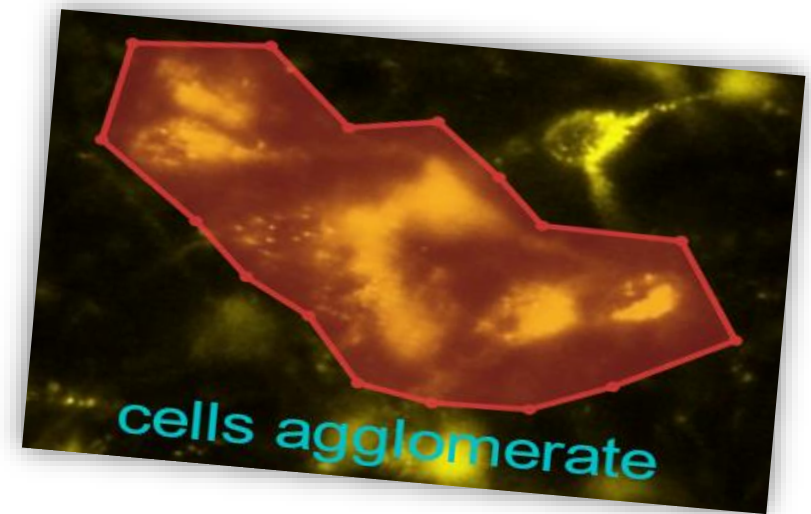
Class imbalance

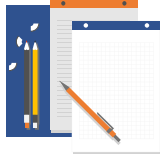


Overcrowding

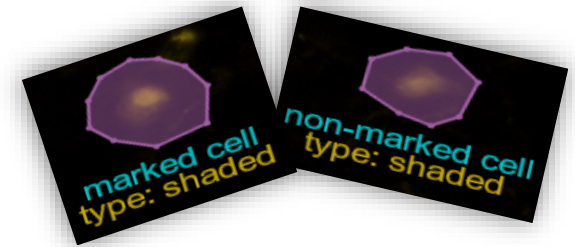
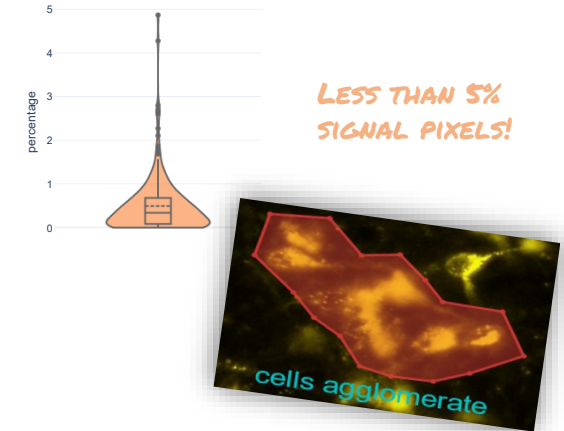
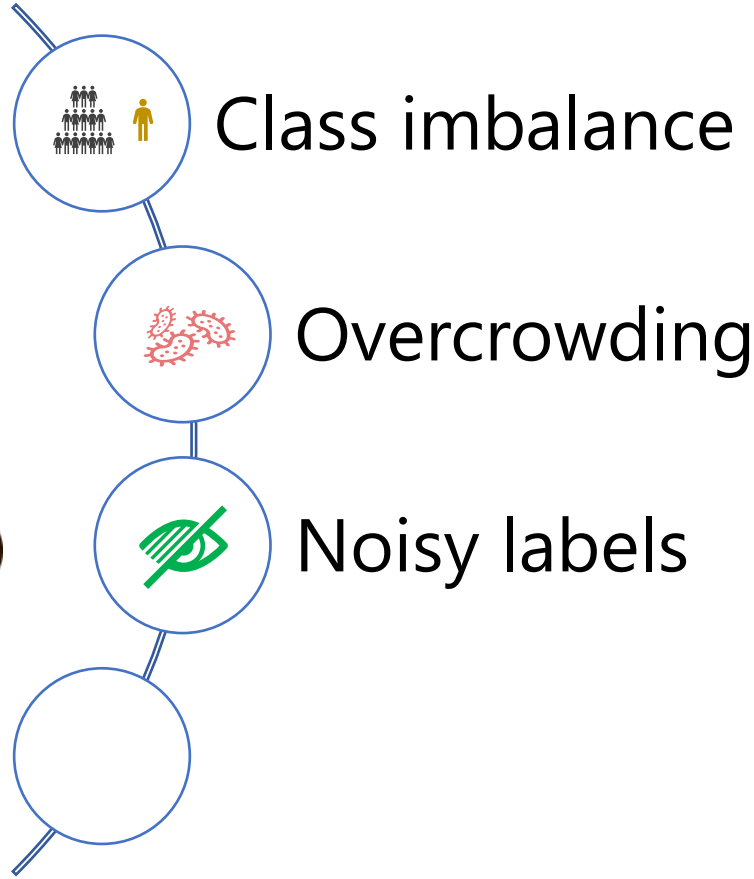
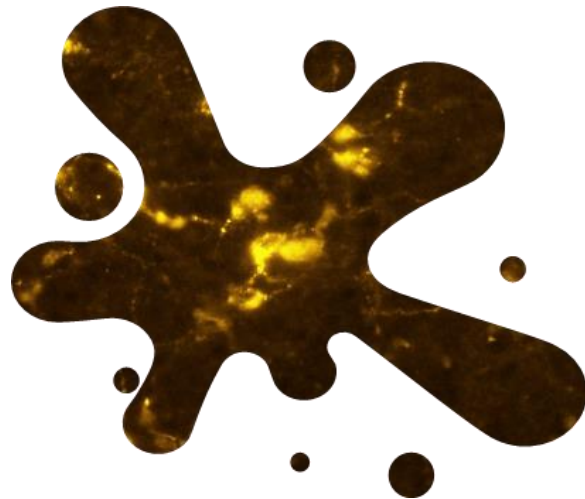


LESS THAN 5%  
SIGNAL PIXELS!



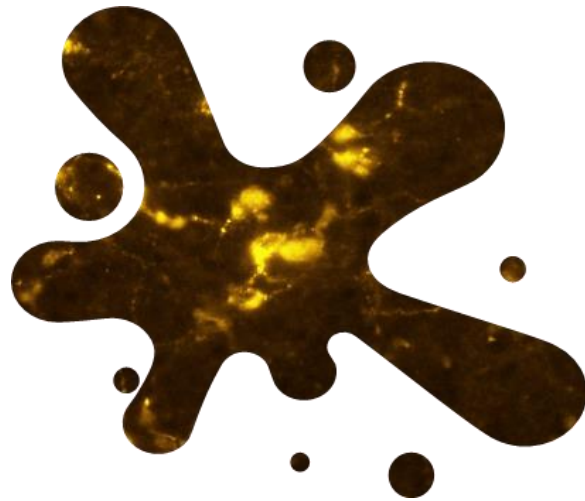


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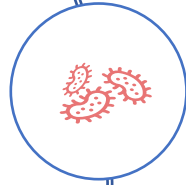




# Challenges



Class imbalance



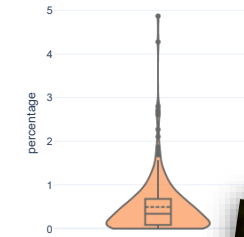
Overcrowding



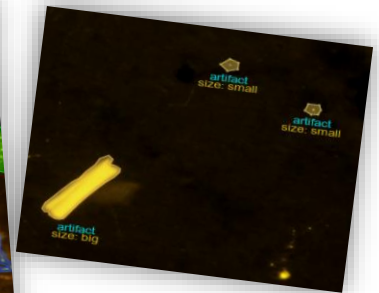
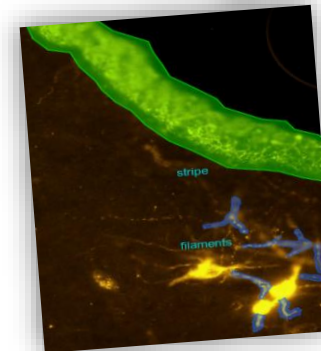
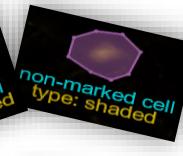
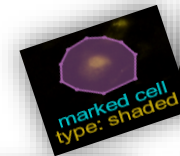
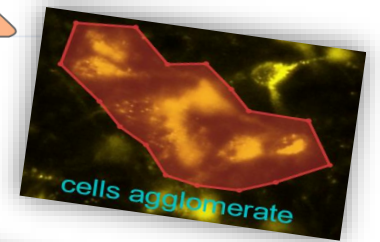
Noisy labels



Artefacts



LESS THAN 5%  
SIGNAL PIXELS!





# Loss functions

Which loss should we choose to address or mitigate these challenges?



# Weighted Binary Cross Entropy [8]

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$$L_{W-BCE}(y, \hat{y}) = -(\beta * y \log(\hat{y}) + (1 - y) \log(1 - \hat{y}))$$

- 👍 de facto standard for classification
  - suitable for binary segmentation
- 👍 weighted version mitigates **class imbalance**
  - assign higher weights to underrepresented class
- 👎 No explicit **segmentation focus**
- 👎 No explicit **object-level error focus**
- 👎 No explicit **noise and systematics focus**



# Focal Loss [9]

$$FL(p_t) = -\alpha_t(1 - p_t)^\gamma \log(p_t) \quad p_t = \begin{cases} p, & \text{if } y = 1 \\ 1 - p, & \text{otherwise} \end{cases}$$

- 👍 Oversample wrong predictions, focus on hard examples  
→ indirectly helpful for any challenge
- 👎 No explicit **segmentation focus**
- 👎 No explicit **object-level error focus**

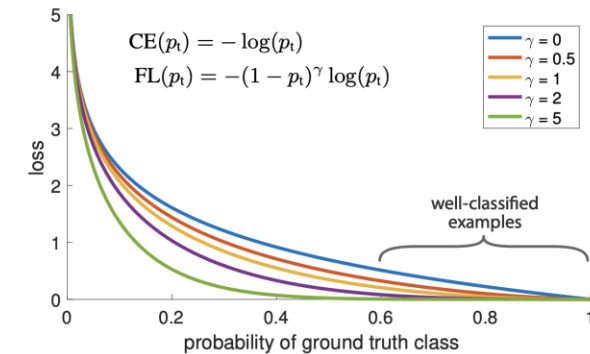
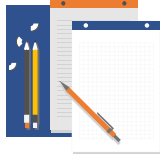


Figure 1. We propose a novel loss we term the *Focal Loss* that adds a factor  $(1 - p_t)^\gamma$  to the standard cross entropy criterion. Setting  $\gamma > 0$  reduces the relative loss for well-classified examples ( $p_t > .5$ ), putting more focus on hard, misclassified examples. As our experiments will demonstrate, the proposed focal loss enables training highly accurate dense object detectors in the presence of vast numbers of easy background examples.





# Dice Loss [10]

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$$DL(y, \hat{p}) = 1 - \frac{2y\hat{p} + 1}{y + \hat{p} + 1}$$

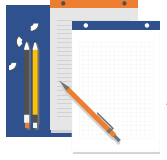
- ✔ targets **segmentation performance** directly  
→ optimization aligned with learning goal
- ✘ No explicit **object-level error focus**
- ✘ No explicit **noise and systematics focus**
- ✘ Low impact of **small objects**  
→ poor generalization WRT object size



# Focal-Tversky Loss [11]

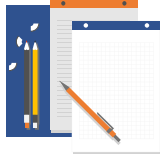
$$FTL = \sum_c (1 - TI_c)^\gamma \quad TI(p, \hat{p}) = \frac{p\hat{p}}{p\hat{p} + \beta(1-p)\hat{p} + (1-\beta)p(1-\hat{p})}$$

- 👍 Brings together advantages of Dice and Focal losses
  - direct focus on segmentation
  - indirectly helpful for any challenge
- 👎 No explicit **object-level error focus**



# Ablation studies

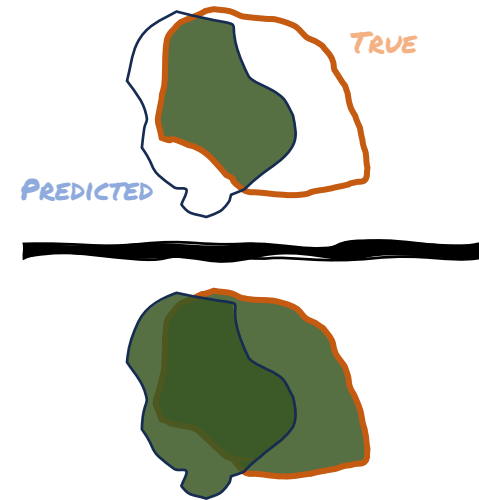
- 4 alternative losses
  - Weighted Binary Cross Entropy (BCE):  $w_{cell} = 50, 100, 200$ ;  $w_{bkgd} = 1$
  - Dice Loss
  - Focal Loss
  - Focal Tversky Loss
- 2 combined losses
  - CombinedLoss =  $\lambda_1 BCE + \lambda_2 Dice + \lambda_3 Focal$
  - CombinedFTLoss =  $\lambda_1 BCE + \lambda_2 Dice + \lambda_3 Focal Tversky$ 
    - Balanced:  $\lambda_1 = 0.3, \lambda_2 = 0.3, \lambda_3 = 0.4$
    - Overcrowd:  $\lambda_1 = 0.2, \lambda_2 = 0.5, \lambda_3 = 0.3$
    - CellViT:  $\lambda_1 = 0.5, \lambda_2 = 0.3, \lambda_3 = 0.5$

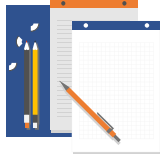


# Evaluation Metrics

- Segmentation

- Mean Intersection over Union (mIoU) =  $\frac{\text{Intersection}}{\text{Union}}$
- threshold: 0.4

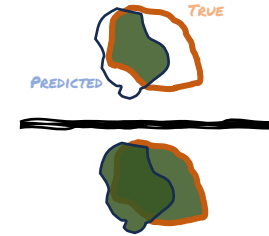


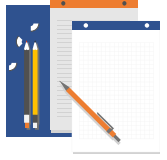


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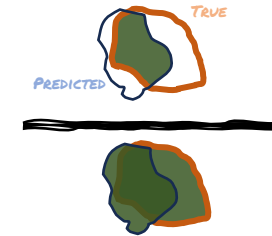




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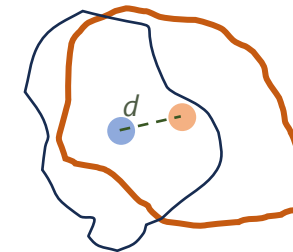
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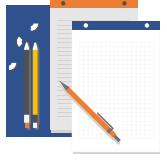
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- Detection

- Centers distance
- threshold: 40 pixels (mean cell diameter)

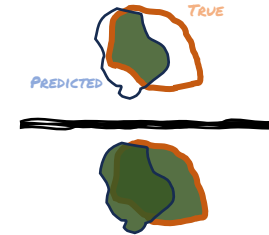




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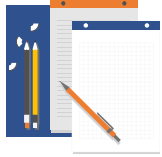
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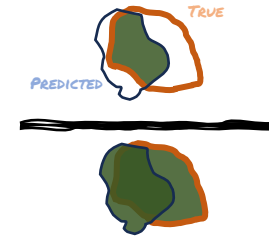




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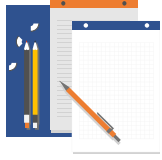
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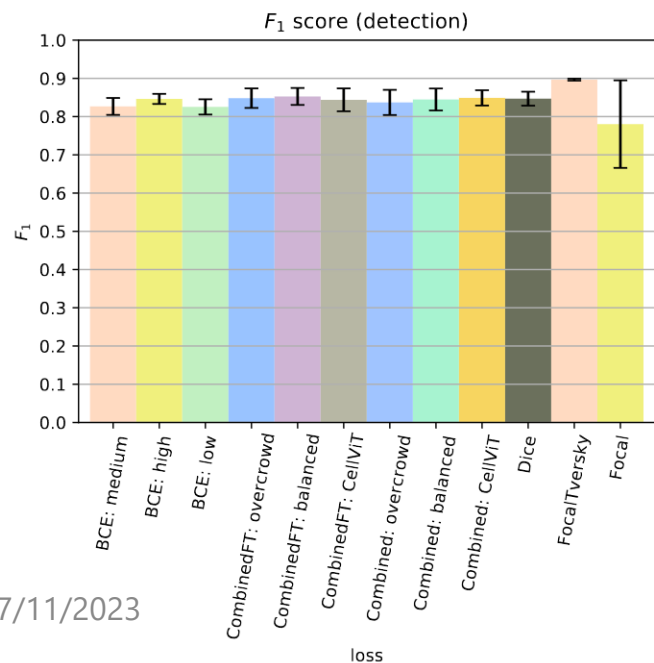
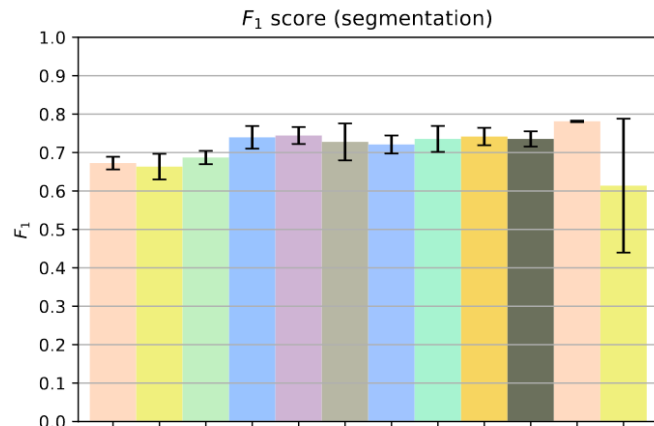
- Counting

- Mean Absolute Error
- Median Absolute Error
- Mean Percentage Error:  $\frac{(n_t - n_p)}{\max(n_t, 1)} * 100$





# Segmentation & Detection

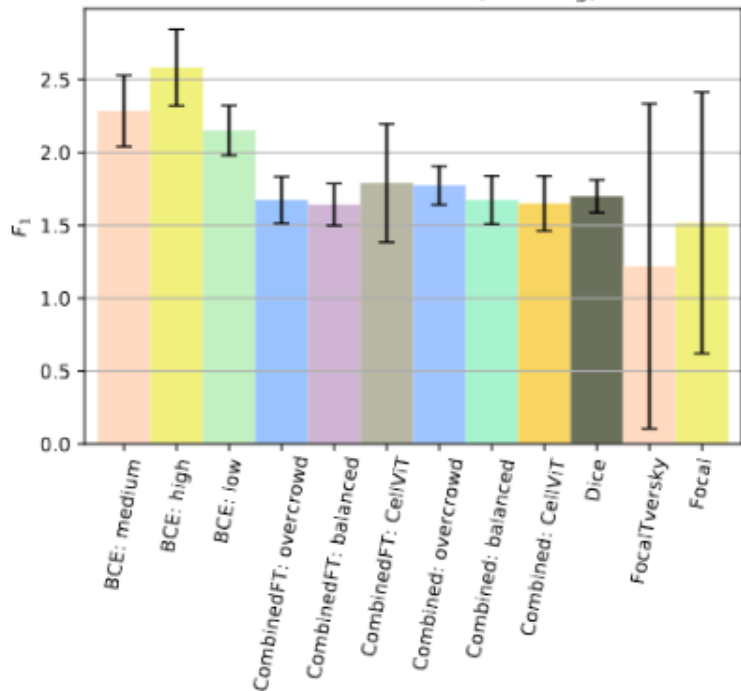


Loss	$F_1$ score (IoU)	$F_1$ score (distance)
BCE: medium	$0.673 \pm 0.017$	$0.827 \pm 0.022$
BCE: high	$0.663 \pm 0.033$	$0.846 \pm 0.013$
BCE: low	$0.687 \pm 0.017$	$0.825 \pm 0.020$
CombinedFT: overcrowd	$0.740 \pm 0.029$	$0.848 \pm 0.026$
CombinedFT: balanced	$0.744 \pm 0.022$	$0.853 \pm 0.022$
CombinedFT: CellViT	$0.728 \pm 0.048$	$0.844 \pm 0.030$
Combined: overcrowd	$0.721 \pm 0.023$	$0.837 \pm 0.033$
Combined: balanced	$0.735 \pm 0.034$	$0.845 \pm 0.029$
Combined: CellViT	$0.742 \pm 0.023$	$0.849 \pm 0.020$
Dice	$0.735 \pm 0.020$	$0.847 \pm 0.018$
<b>Focal Tversky</b>	<b><math>0.781 \pm 0.002</math></b>	<b><math>0.897 \pm 0.003</math></b>
Focal	$0.614 \pm 0.027$	$0.780 \pm 0.034$

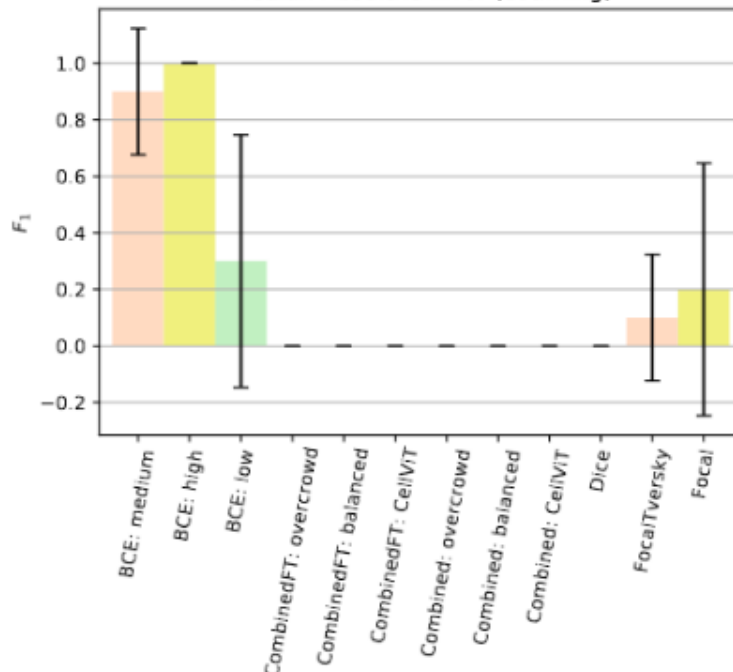


# Counting

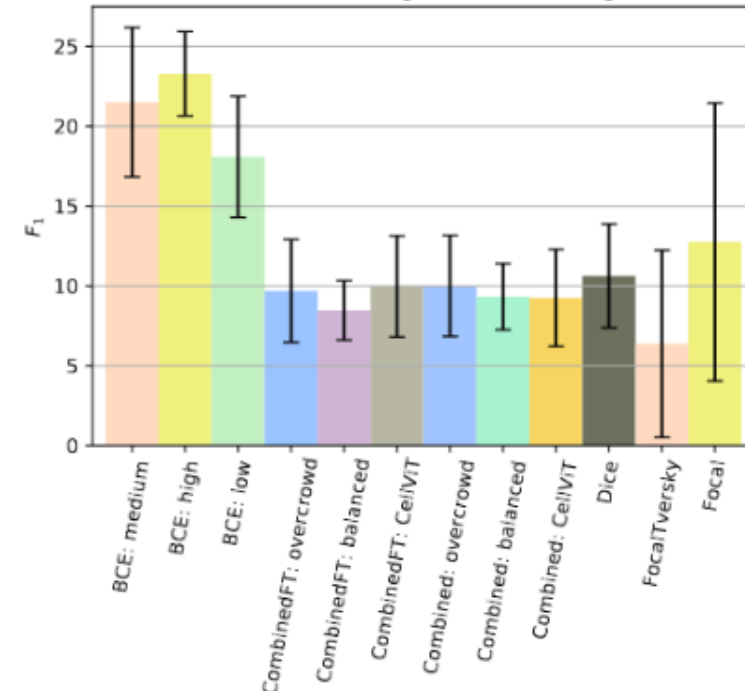
Mean Absolute Error (counting)



Median Absolute Error (counting)



Mean Percentage Error (counting)



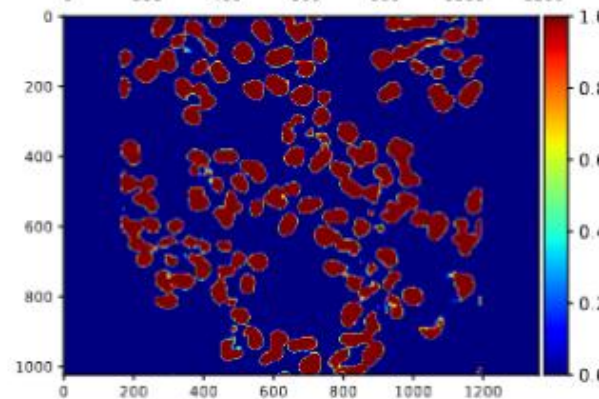
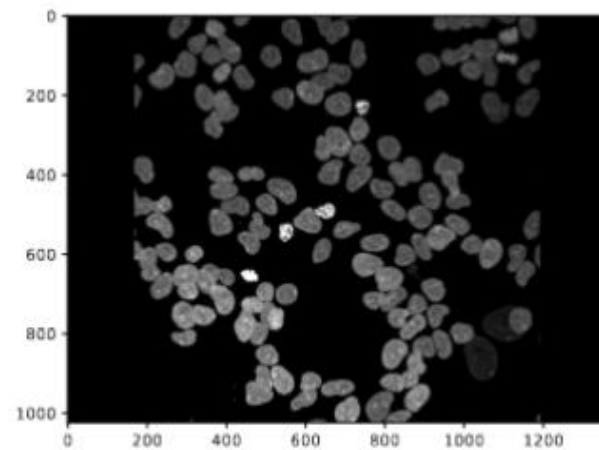
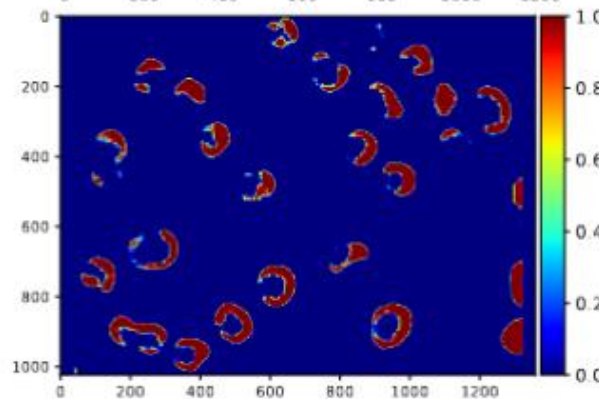
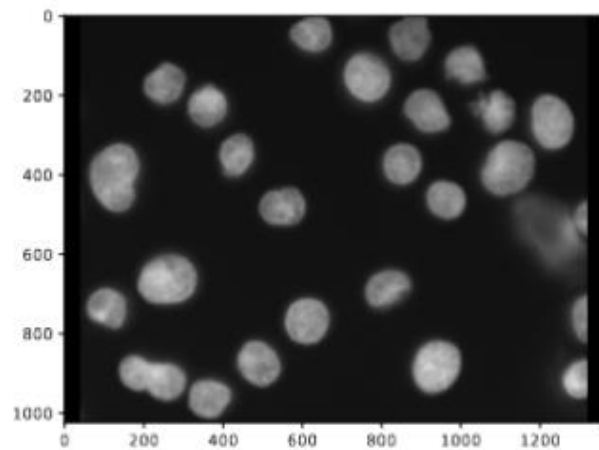
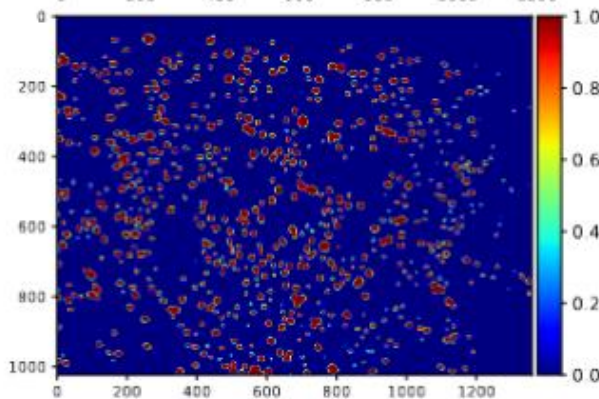
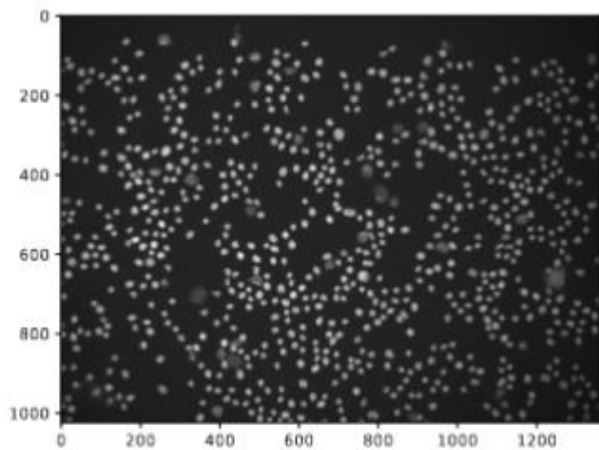
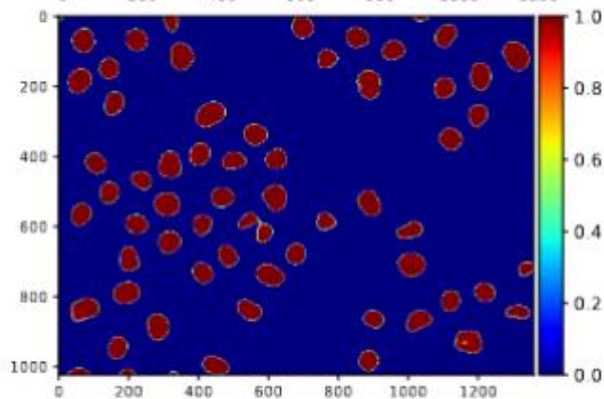
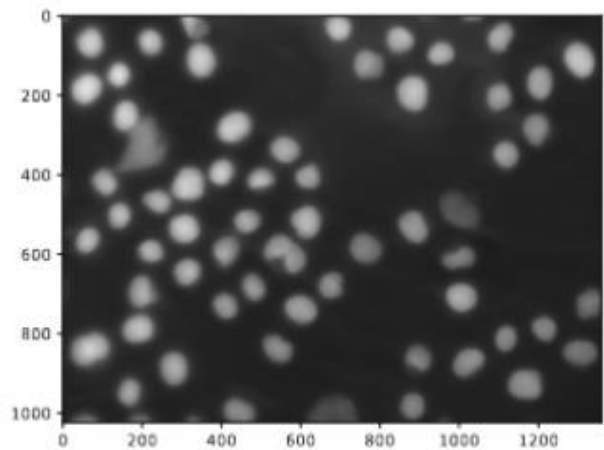
*BCE: medium*    *BCE: high*    *BCE: low*    *CombinedFT: overcrowd*    *CombinedFT: balanced*    *CombinedFT: CellViT*    *Combined: overcrowd*    *Combined: balanced*    *Combined: CellViT*    *Dice*    *Focal Tversky*    *Focal*

	<i>BCE: medium</i>	<i>BCE: high</i>	<i>BCE: low</i>	<i>CombinedFT: overcrowd</i>	<i>CombinedFT: balanced</i>	<i>CombinedFT: CellViT</i>	<i>Combined: overcrowd</i>	<i>Combined: balanced</i>	<i>Combined: CellViT</i>	<i>Dice</i>	<i>Focal Tversky</i>	<i>Focal</i>
<b>MAE</b>	2.286±0.245	2.583±0.263	2.151±0.171	1.674±0.161	1.643±0.144	1.791±0.406	1.774±0.132	1.674±0.166	1.651±0.188	1.700±0.113	1.220±1.115	1.517±0.365
<b>MedAE</b>	0.9±0.224	1±0	0.3±0.447	0±0	0±0	0±0	0±0	0±0	0±0	0±0	0.1±0.224	0.2±0.4
<b>MPE</b>	21.498±4.679	23.281±2.637	18.079±3.791	9.682±3.221	8.465±1.859	9.968±3.163	10±3.160	9.322±2.074	9.249±3.039	10.621±3.247	<b>6.373±1.607</b>	12.747±3.525



# Out-of-sample generalization

[12] S-BSST265 dataset

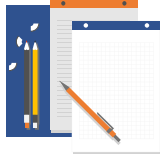


(a) Medium-sized, sharp objects

(b) Small objects

(c) Uneven texture and filling

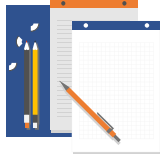
(d) Overcrowding



# Conclusions

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- **Focal Tversky** loss overperforms other losses
- Still some trouble separating **crowded objects**  
→ careful post-processing needed (hole filling, small objects, watershed)
- Combined losses are competitive
  - Better tuning of lambda weights
- Generalization
  - High variability
  - Dedicated augmentation may help
  - **Panoptic loss** → **single object errors**
- Multiple metrics enable comprehensive assessment



# References

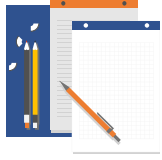
- [1] [Dominé, L. and Terao, K: Scalable deep convolutional neural networks for sparse, locally dense liquid argon time projection chamber data. \*Phys. Rev. D\* 102 \(2020\).](#)
- [2] [Holm, E.A., et al. Overview: Computer Vision and Machine Learning for Microstructural Characterization and Analysis. \*Metall Mater Trans A\* 51 \(2020\).](#)
- [3] [Pino, C., et al.: Semantic Segmentation of Radio-Astronomical Images. In Progress in Artificial Intelligence and Pattern Recognition. IWAIPR 2021. Lecture Notes in Computer Science \(2021\).](#)
- [4] [Zhang, H. et al.: Automatic seismic facies interpretation using supervised deep learning. \*Geophysics\* \(2021\)](#)
- [5] [Hitrec, T., et al.: Neural control of fasting-induced torpor in mice. \*Scientific Reports\* 9\(1\) \(oct 2019\).](#)
- [6] [Morelli, R., et al.: Automating cell counting in fluorescent microscopy through deep learning with c-ResUnet. \*Scientific Reports\* 11\(1\), 22920 \(2021\).](#)
- [7] [Clissa, L., et al.: Fluorescent neuronal cells v2: Multi-task, multi-format annotations for deep learning in microscopy. arXiv preprint \(submitted to Scientific Data\) \(2023\)](#)
- [8] [Pihur, V., Datta, S., Datta, S.: Weighted rank aggregation of cluster validation measures: a monte carlo cross-entropy approach. \*Bioinformatics\* 23\(13\), \(2007\).](#)
- [9] [Lin, T. Y., et al.: Focal loss for dense object detection. \*ICCV Proceedings\* \(2017\).](#)
- [10] [Sudre, C.H., et al.: Generalised dice overlap as a deep learning loss function for highly unbalanced segmentations. \*Deep Learning in Medical Image Analysis and Multimodal Learning for Clinical Decision Support.\* \(2017\).](#)
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- [12] [Kromp, F., et al.: An annotated fluorescence image dataset for training nuclear segmentation methods. \*Scientific Data\* 7\(1\), 262 \(2020\)](#)
- [13] [Jadon, S.: A survey of loss functions for semantic segmentation. In: 2020 IEEE conference on computational intelligence in bioinformatics and computational biology \(CIBCB\). pp. 1-7 \(2020\)](#)



**Thanks for  
your attention!**

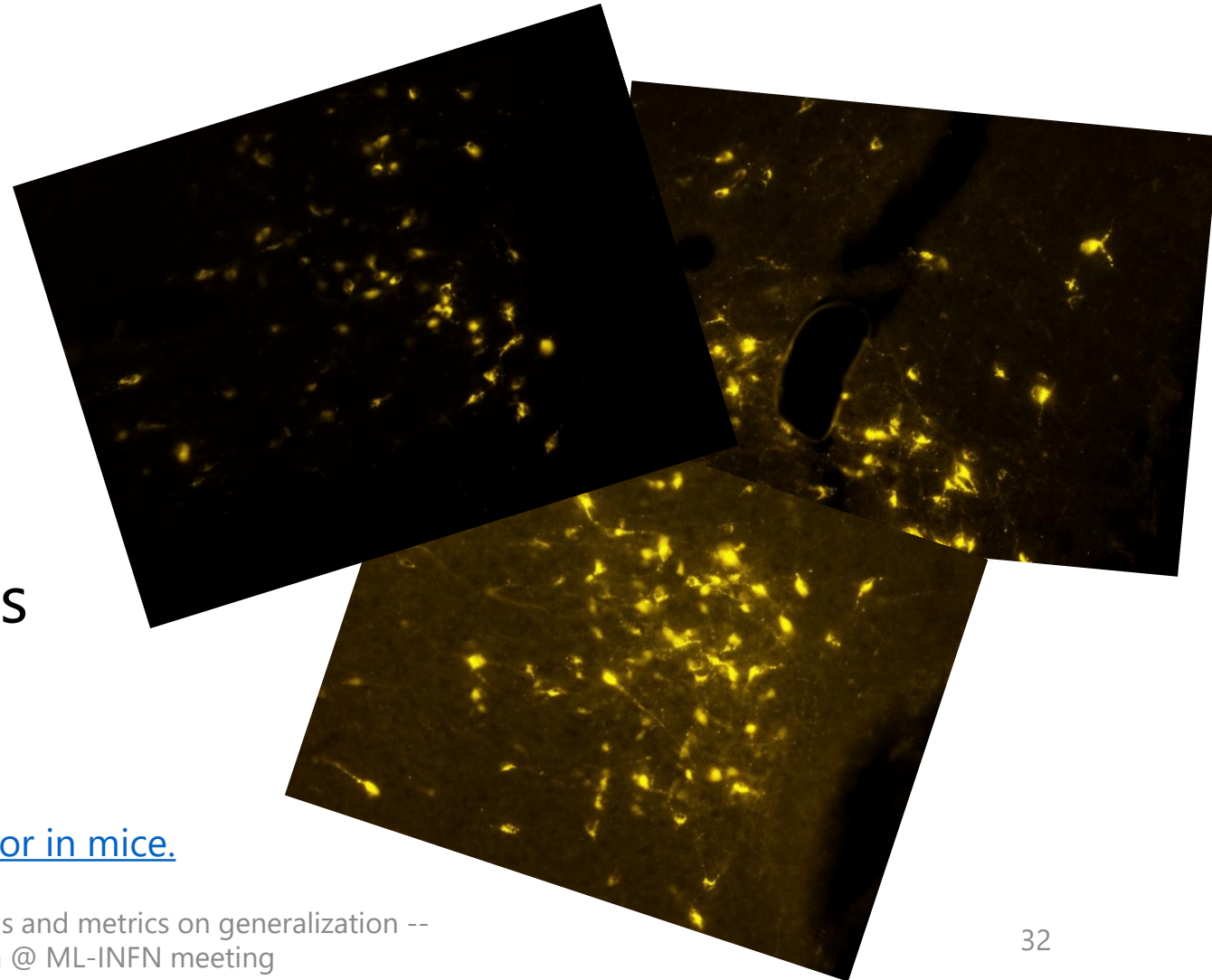
Questions?

# Backup



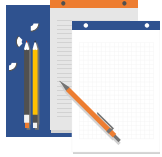
# Torpor onset

- Very popular in life science
  - Torpor onset [\[1\]](#)
- Cytoplasmatic neuronal structures
- Variability in shape, size and color hue
- **Goal:** count stained structures



[\[1\] Hitrec, T., et al.: Neural control of fasting-induced torpor in mice. Scientific Reports 9\(1\) \(oct 2019\).](#)





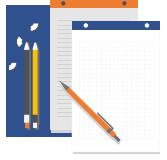
# Contributions

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- Semantic segmentation using c-ResUnet [\[2\]](#) and Fluorescent Neuronal Cells v2 dataset (FNC v2) [\[3\]](#)
- Show the impact of loss functions on model performance
  - 60 ablation studies
  - 6 loss functions
- Inspect pros and cons of several evaluation metrics
- Discuss characteristics affecting out-of-sample generalization

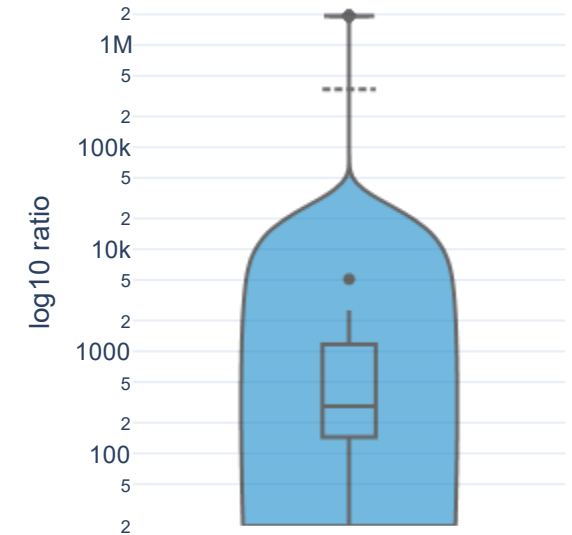
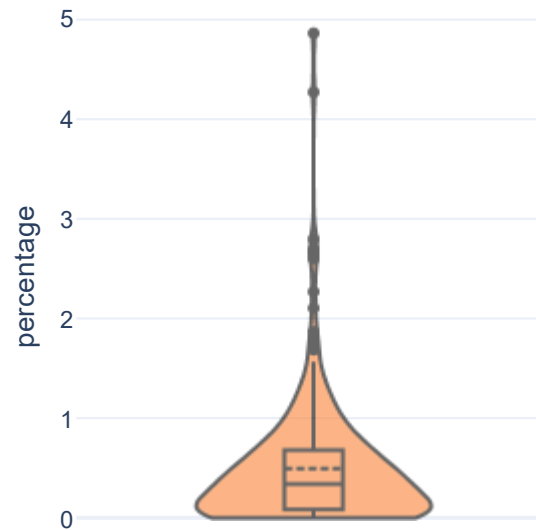
[\[2\] Morelli, R., et al.: Automating cell counting in fluorescent microscopy through deep learning with c-ResUnet. Scientific Reports 11\(1\), 22920 \(2021\).](#)

[\[3\] Clissa, L., et al.: Fluorescent neuronal cells v2: Multi-task, multi-format annotations for deep learning in microscopy. arXiv preprint \(submitted to Scientific Data\) \(2023\)](#)



# Class imbalance

quantile	signal (%)	signal ratio
mean	0.50	367k
s.d.	0.61	756k
min	0	19.57
10%	0	92.39
25%	0.09	145.35
50%	0.34	291.10
75%	0.68	1k
90%	1.07	1.9M
max	4.86	1.9M



10  $\mu$ m

cells agglomerate

cells agglomerate

non-marked cell  
type: shaded

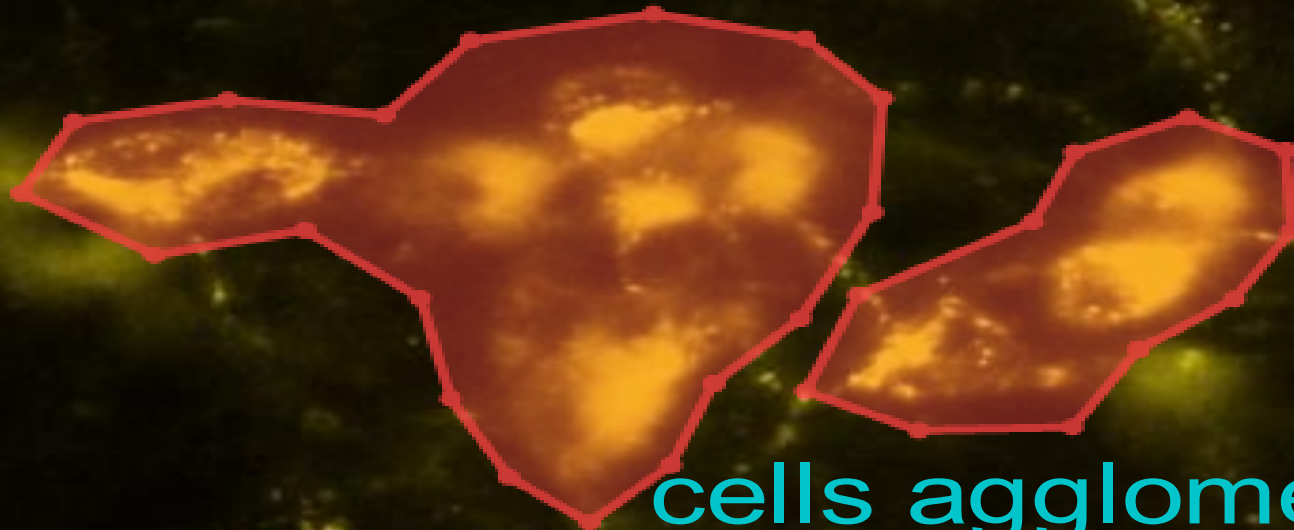
marked cell  
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cells agglomerate  
cells agglomerate

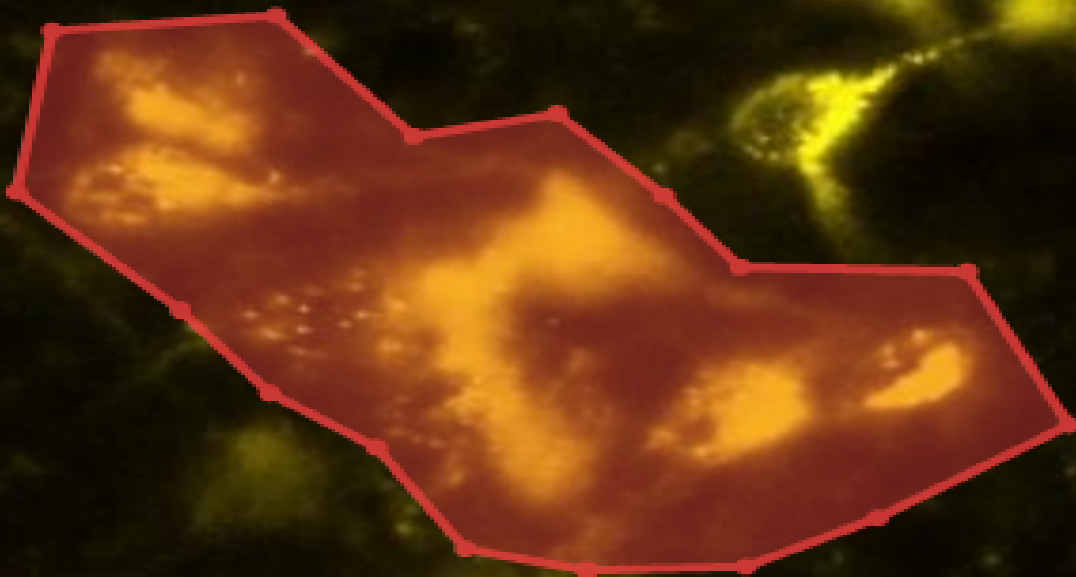
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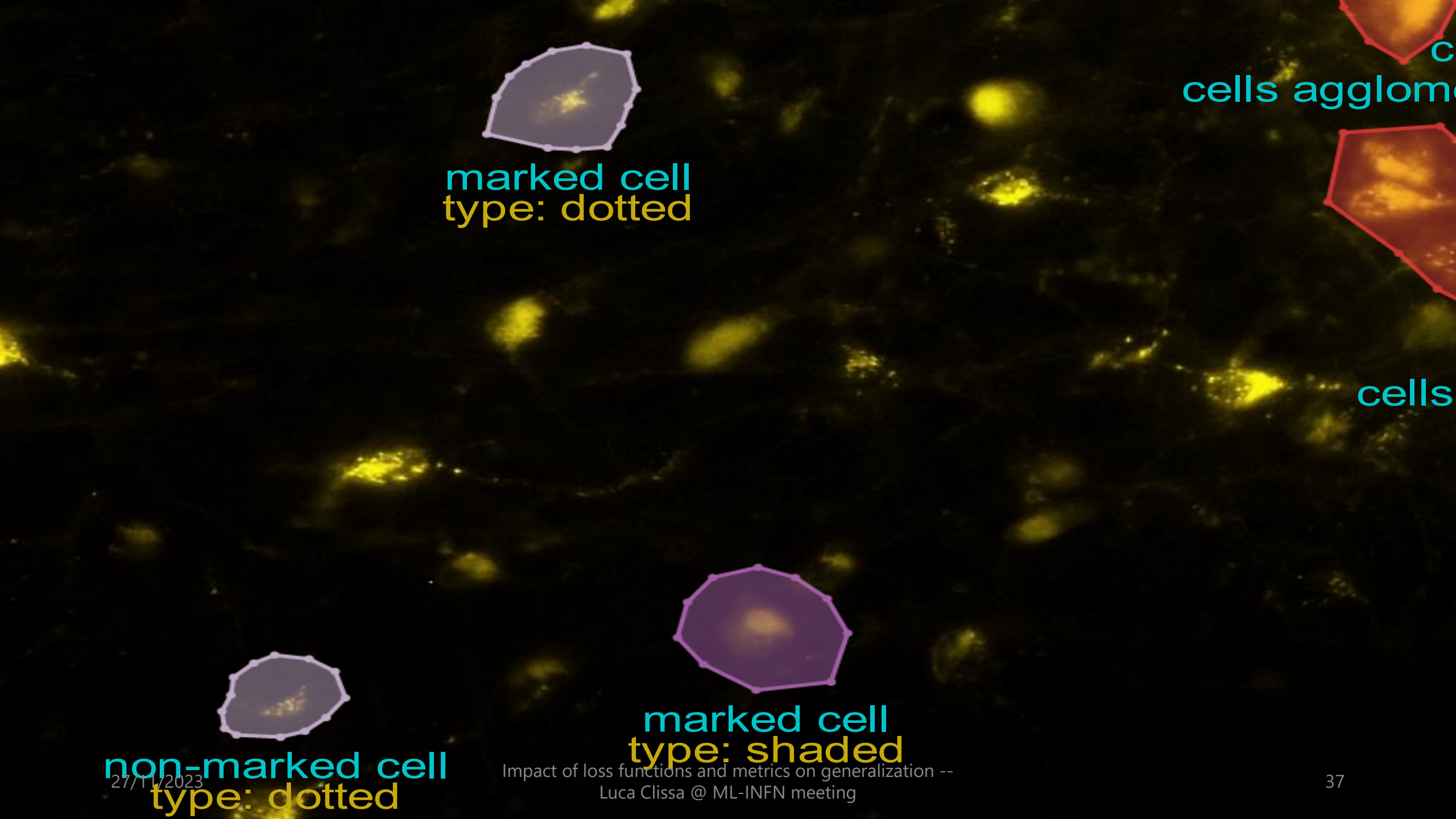
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cells agglomerate  
cells agglomerate



cells agglomerate



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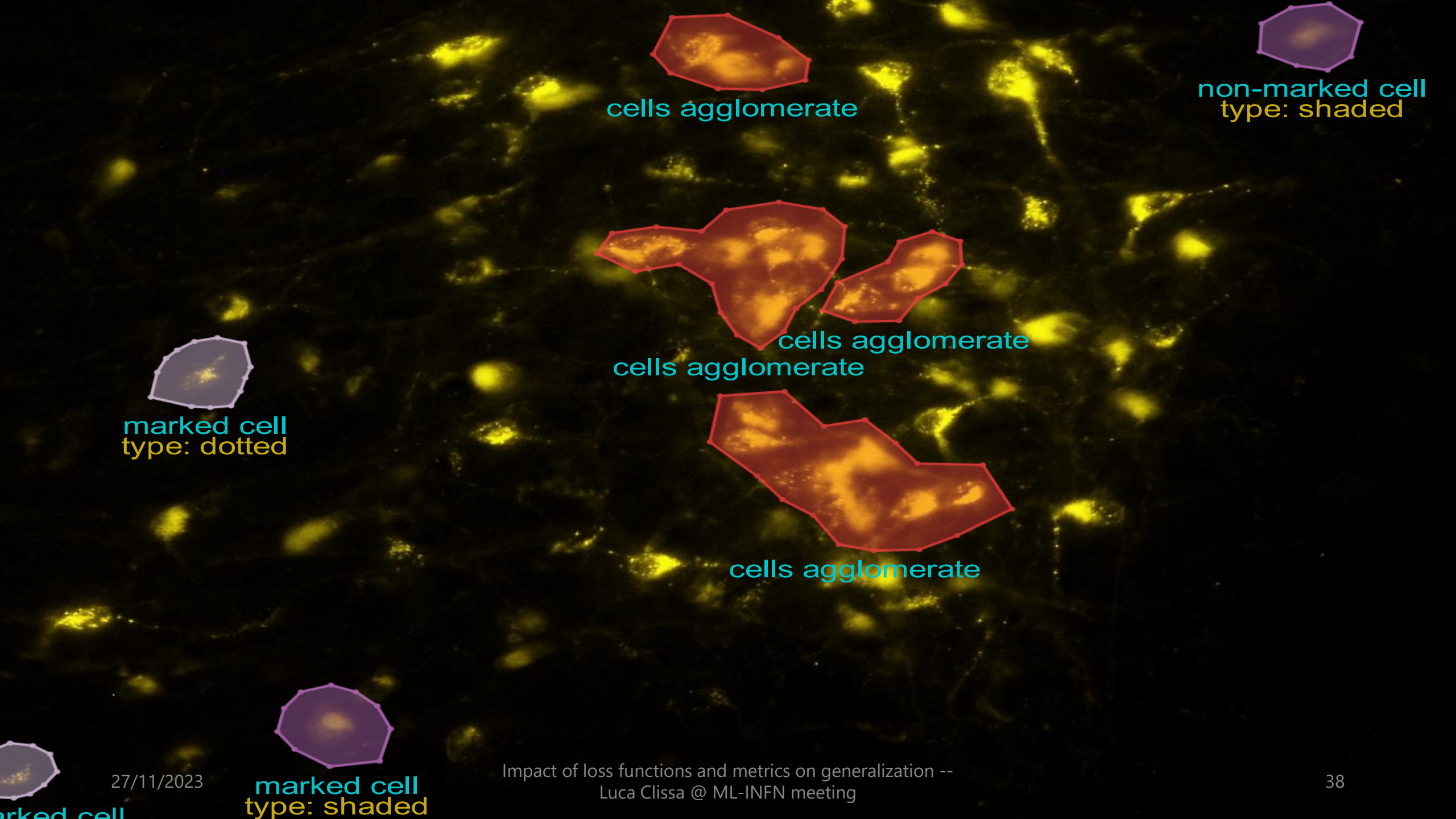
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cells

non-marked cell  
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cells agglomerate

non-marked cell  
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cells agglomerate  
cells agglomerate

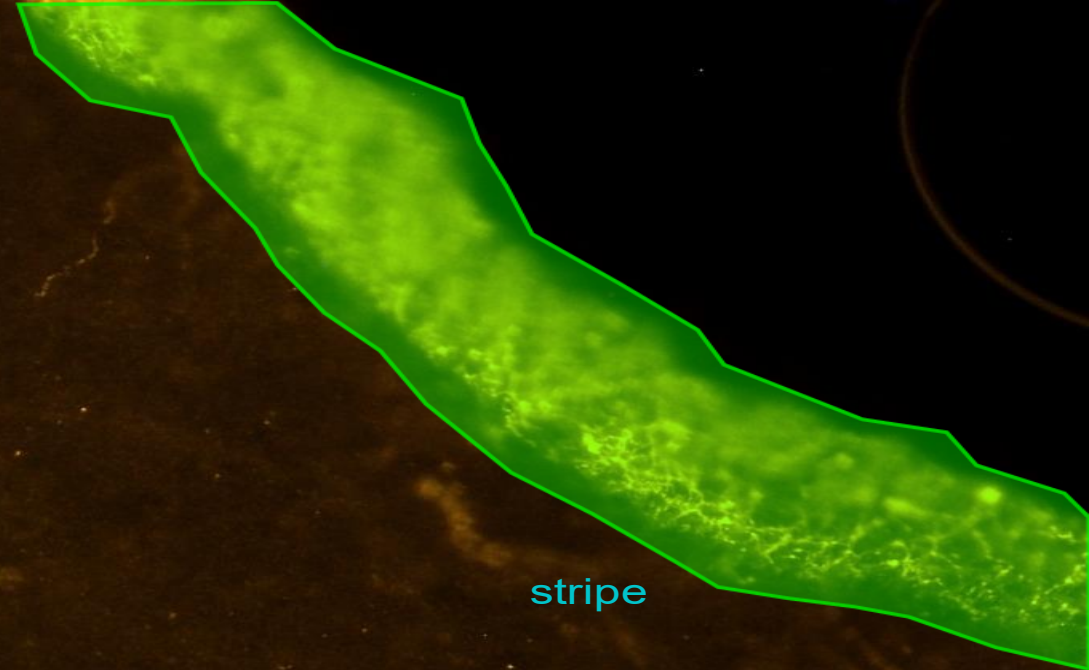
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cells agglomerate

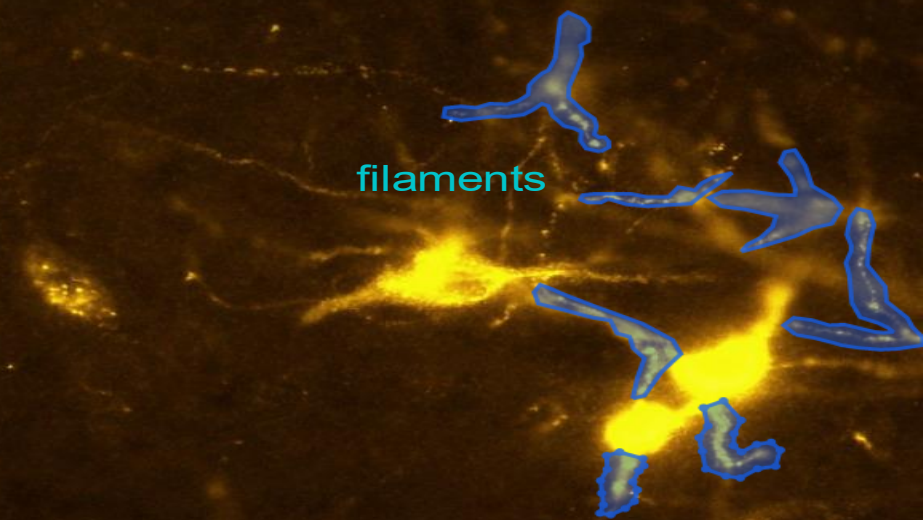
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27/11/2023

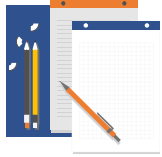
10  $\mu\text{m}$



stripe



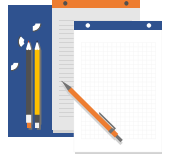
filaments



# Ablation studies configurations

	BCE	Dice	Focal	Focal Tversky	Combined	Combined FT
Hyperparameters	w_cell	smooth	gamma	gamma	$\lambda_1, \lambda_2, \lambda_3$	$\lambda_1, \lambda_2, \lambda_3$
Values	[50, 100, 200]	$1 \times 10^{-6}$	2	2	balanced: [0.3, 0.3, 0.4] overcrowd: [0.2, 0.5, 0.3] CellViT: [0.5, 0.3, 0.5]	balanced: [0.3, 0.3, 0.4] overcrowd: [0.2, 0.5, 0.3] CellViT: [0.5, 0.3, 0.5]

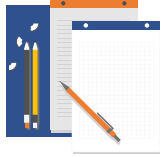




# Training setup

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- Adam optimizer
- Learning rate test for initial LR
- 200 epochs
- Cyclical learning rates
- Best model on validation dice coefficient



# Loss functions

- **Weighted Binary Cross Entropy:** higher weight to underrepresented class

$$L_{W-BCE}(y, \hat{y}) = -(\beta * y \log(\hat{y}) + (1 - y) \log(1 - \hat{y}))$$

- **Dice Loss:** targets segmentation performance directly, low impact of small objects

$$DL(y, \hat{p}) = 1 - \frac{2y\hat{p} + 1}{y + \hat{p} + 1}$$

- **Focal Loss:** oversample hard examples

$$FL(p_t) = -\alpha_t (1 - p_t)^\gamma \log(p_t), \quad p_t = \begin{cases} p, & \text{if } y = 1 \\ 1 - p, & \text{otherwise} \end{cases}$$

- **Focal Tversky Loss:** bring together advantages of Dice and Focal losses

$$FTL = \sum_c (1 - TI_c)^\gamma, \quad TI(p, \hat{p}) = \frac{p\hat{p}}{p\hat{p} + \beta(1 - p)\hat{p} + (1 - \beta)p(1 - \hat{p})}$$