Hands-On on DNA damage quantification

geant4-dna.org

The « moleculardna » extended example

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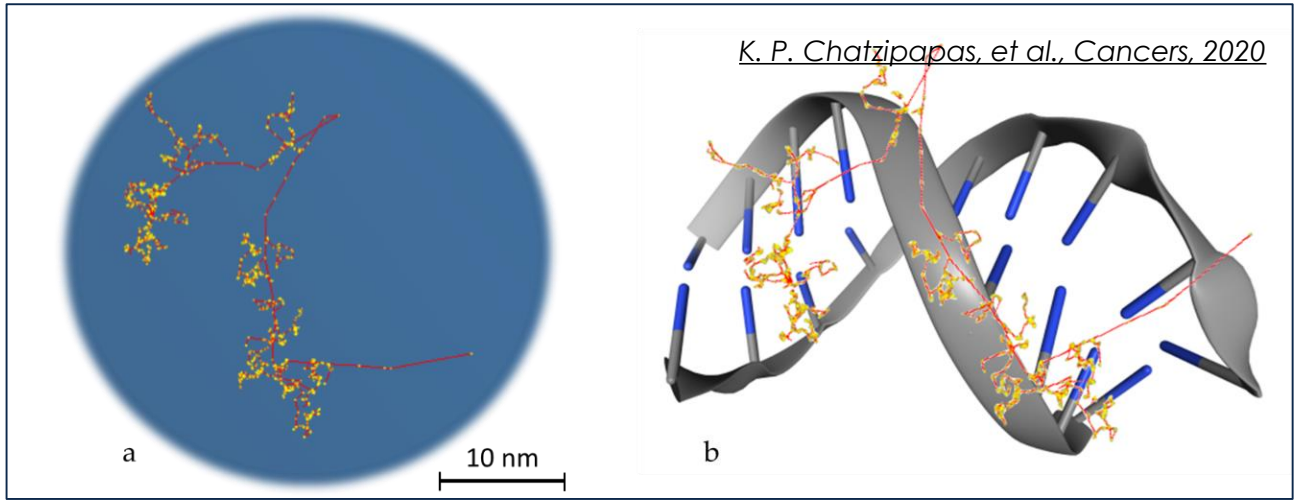
The « moleculardna » extended example

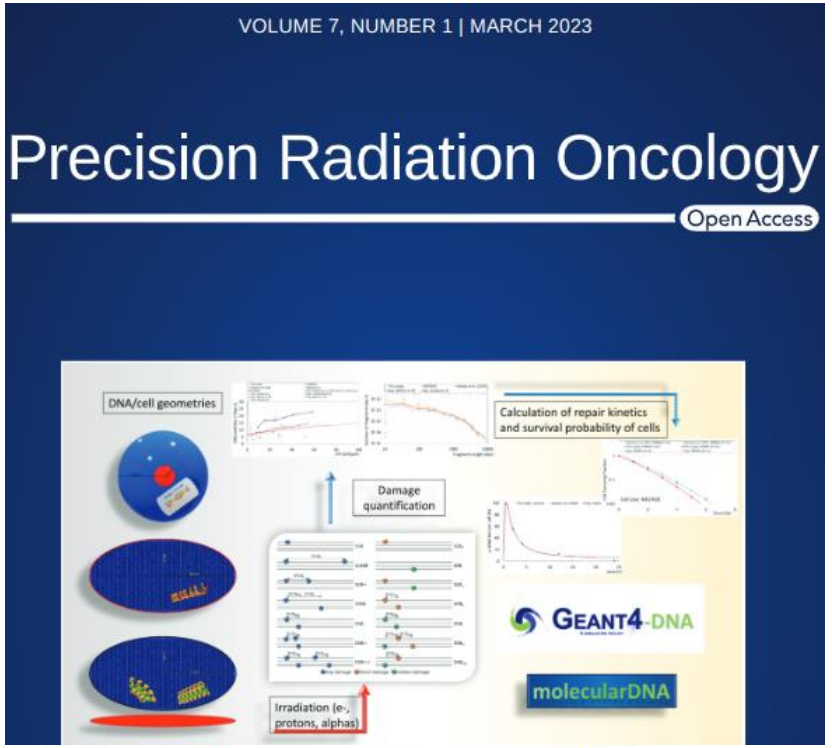
- Try the « **moleculardna** » example, which **combines**
  - Geant4-DNA Physics models
  - Geant4-DNA Chemistry models &
  - Geant4-DNA DNA-scale geometrical models of biological targets

- This is a Geant4 **extended** example and it is located in
  `$G4INSTALL/examples/extended/medical/dna`



The « moleculardna » extended example

- This example aims to demonstrate how the Geant4-DNA toolkit can be used to **quantify DNA damage** induced by ionising irradiation
- Full macro file control of simulation (No C++ knowledge needed)
- Physical interaction and Chemistry models
- Several types of **DNA geometries** are included
  - Cylinders
  - Bacterial DNA
  - Human cells
- Both **direct and indirect** damage can be calculated, taking into account
  - Physics
  - Chemistry (physico-chemical and chemical stages)
- The **complexity of damage** can be investigated

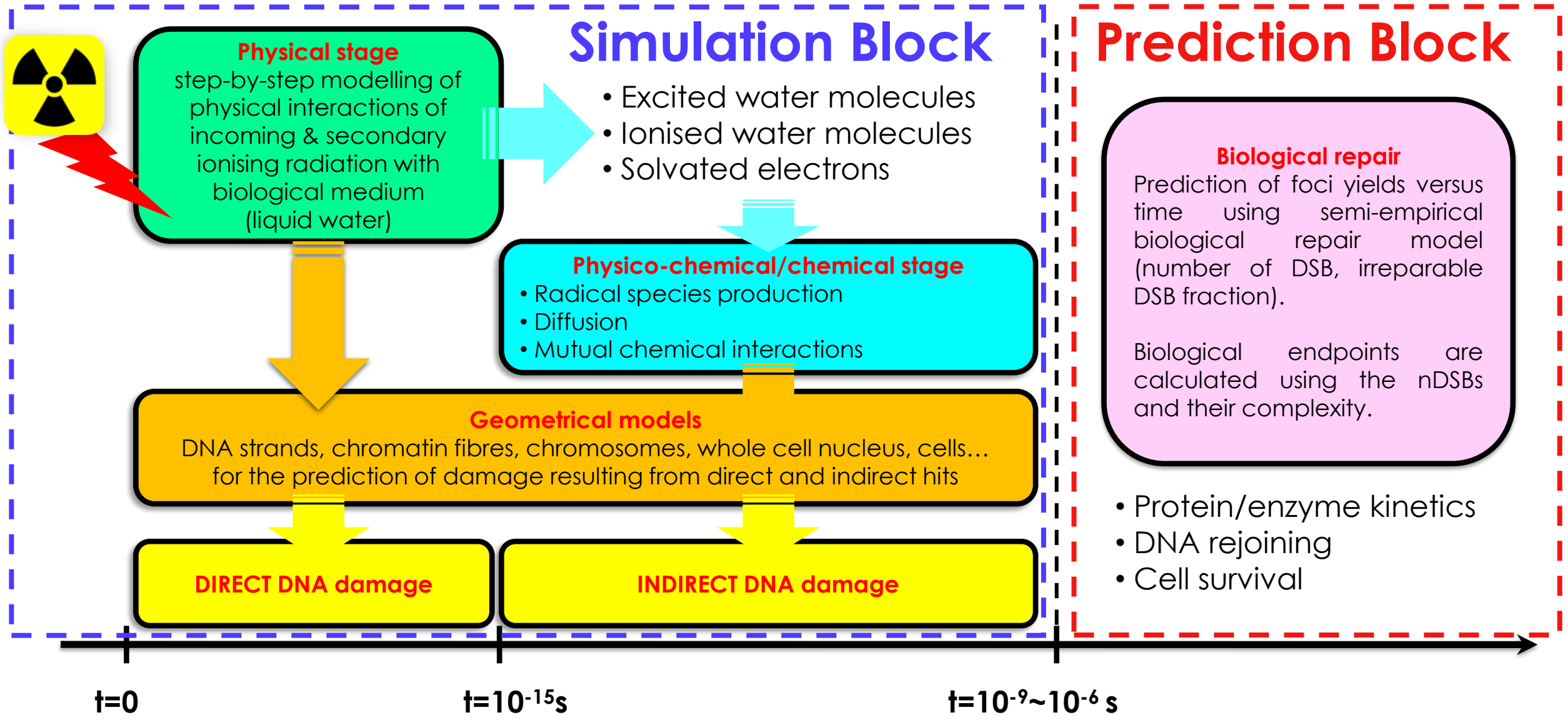


Simulation of DNA damage using Geant4-DNA: an overview of the "molecularDNA" example application

Konstantinos P. Chatzipapas, Ngoc Hoang Tran, Milos Dordevic, Sara Zivkovic, Sara Zein, Wook-Geun Shin, Dousatsu Sakata, Nathanael Lampe, Jeremy M. C. Brown, Aleksandra Ristic-Fira, Ivan Petrovic, Ioanna Kyriakou, Dimitris Emfietzoglou, Susanna Guatelli, Sébastien Incerti
... See fewer authors

https://doi.org/10.1002/pro6.1186
and references therein

moleculardna approach



Physical stage

Recommendation: use Geant4-DNA physics constructors

- G4EmDNAPhysics_**option2**
- G4EmDNAPhysics_**option4**
- G4EmDNAPhysics_**option6**

```
/chem/activate true/false
```

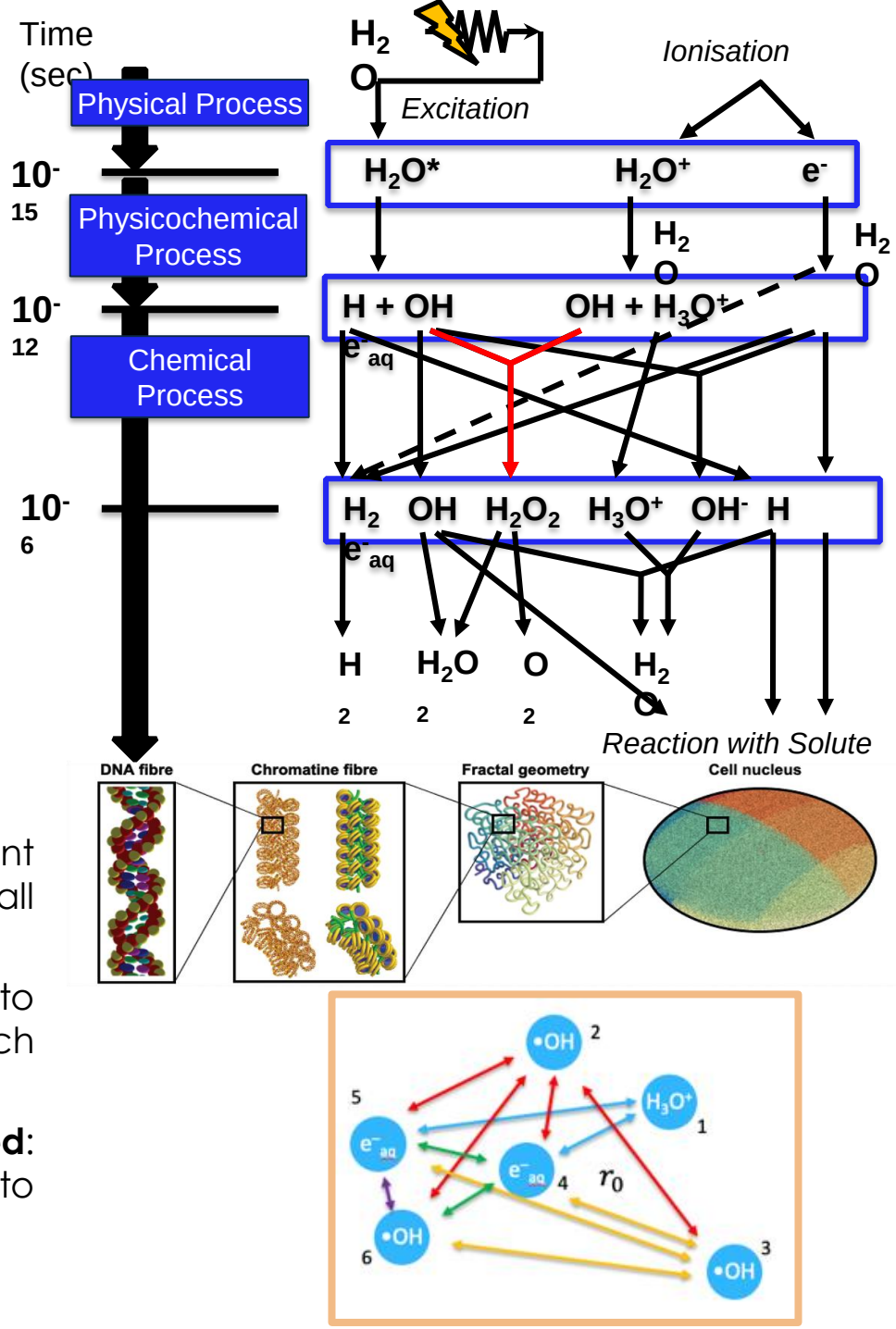
Physicochemical stage

During this stage, free radicals are produced.

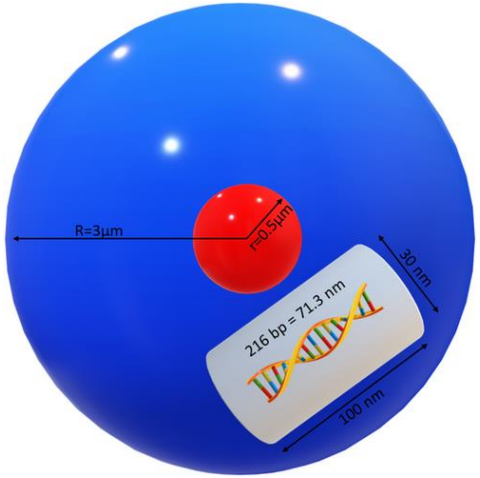
Chemical stage

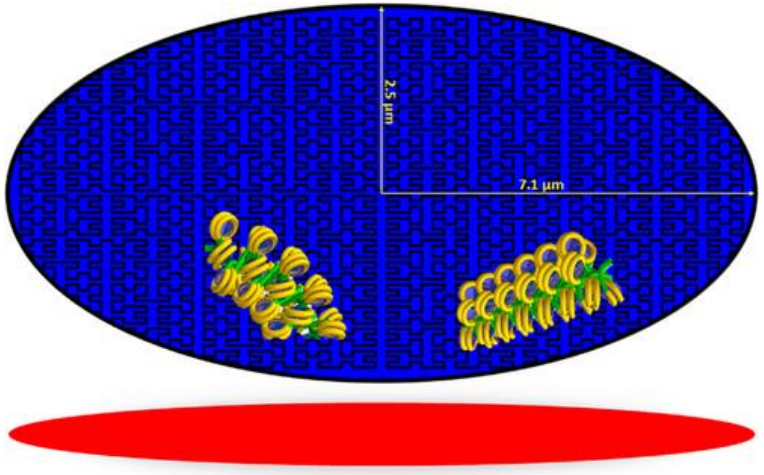
Independent Reaction Times (« IRT ») approach

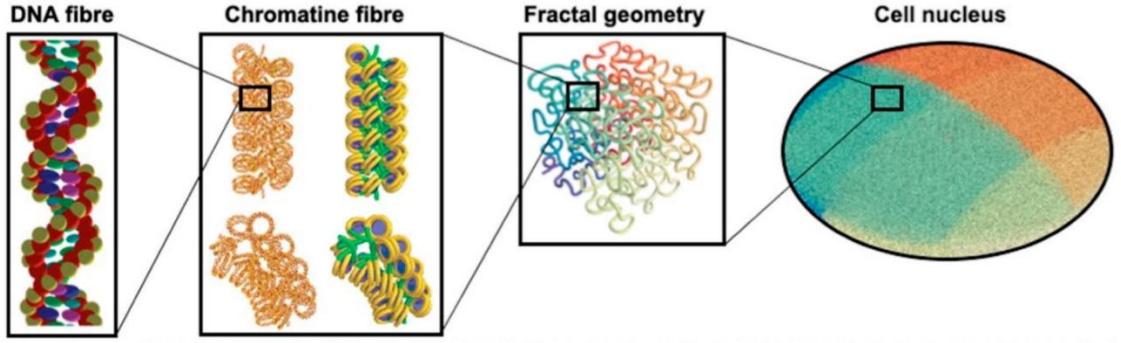
- From the 1980's by Clifford, Green et al., widely used today.
- Iterative process where the approximation of « independent pairs » is assumed: calculates the reaction times between all possible pairs of reactive species, as if they were isolated.
- No longer necessary to diffuse the molecular species and to calculate the possible reactions between the species at each time step.
- A « synchronous » alternative **hybrid version** (« IRT-sync ») **is used**: it gives all spatio-temporal info on radicals, as it is required to combine with the DNA geometries.

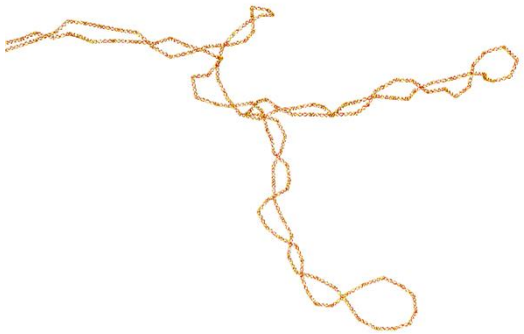


Examples of geometrical models created from the « fractalDNA » tool

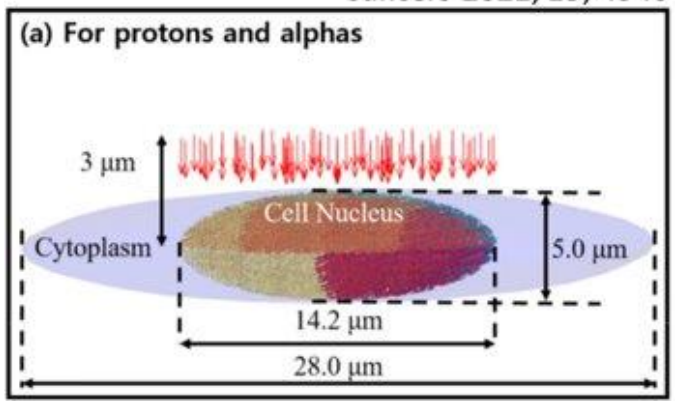


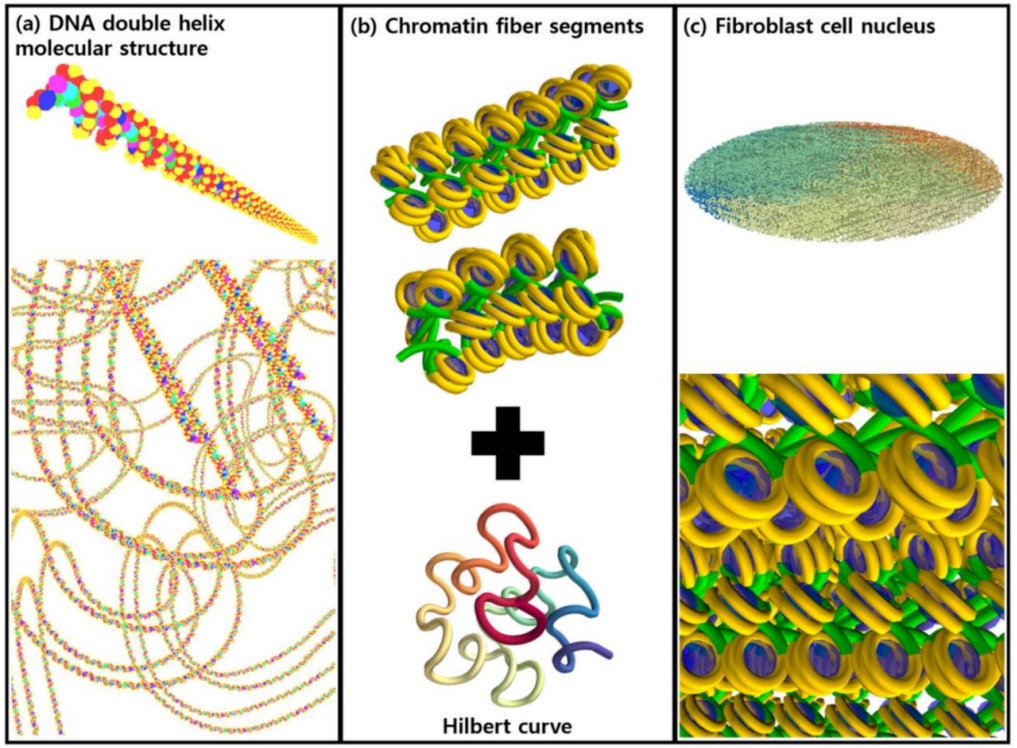




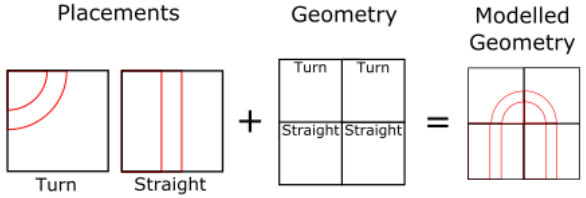


Cancers **2021**, *13*, 4940





The «fractalDNA» tool (by N. Lampe): open-source Python package to build DNA geometries that can be joined together like jigsaw puzzles. Users can build their own geometries based on provided examples.



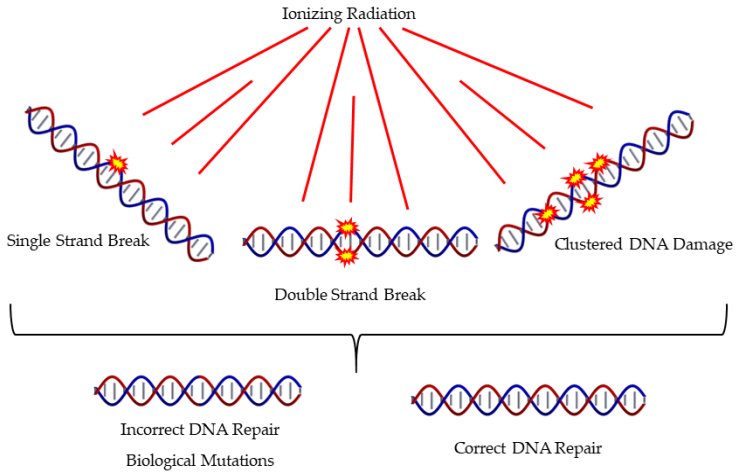
https://pypi.org/project/fractaldna/
http://natl.github.io/fractaldna/

DNA damage classification

- **Direct damage**
  occurs when energy from ionizing particles is deposited near a DNA.

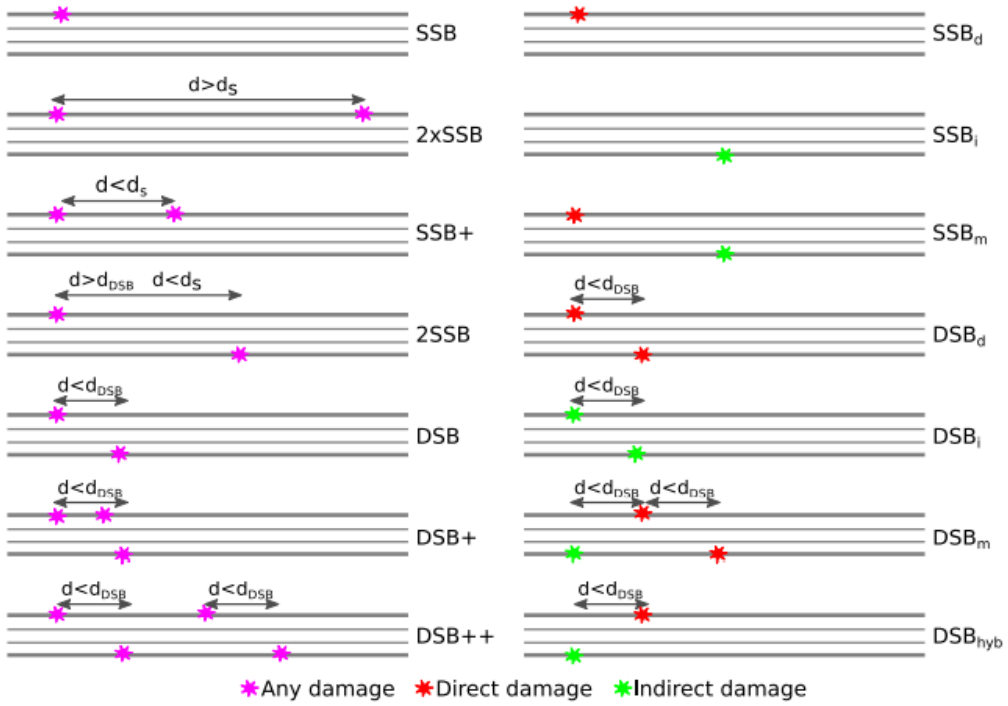
  In molecularDNA, we associate damage either with a **strand** molecule (sugar or phosphate) or a **base** molecule.

- **Indirect damage**
  is scored when a chemical reaction leads to a strand break.



K. P. Chatzipapas et al. Cancers, 2020

Classification scheme

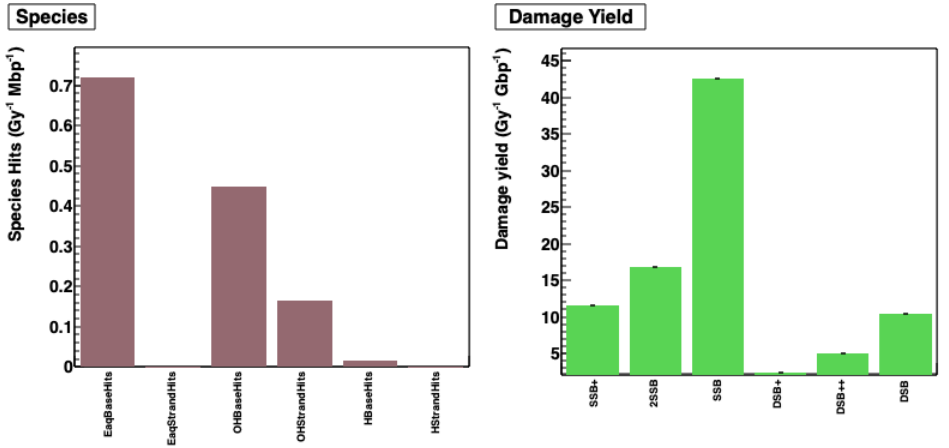


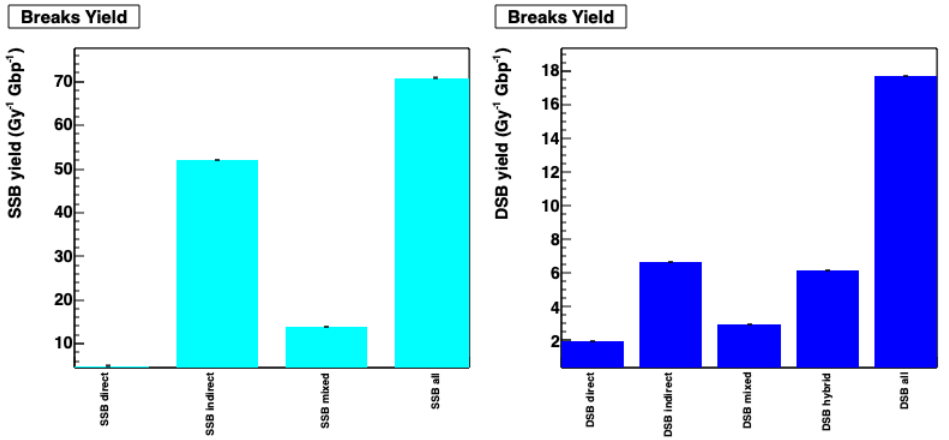
N. Lampe, PhD thesis, 2017

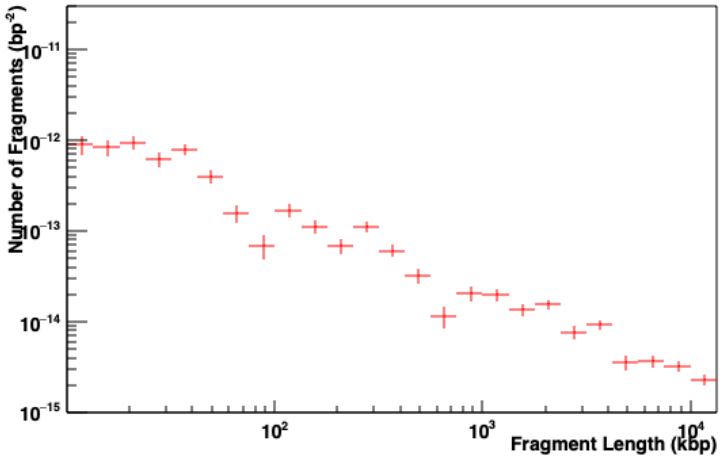
Results: ROOT output file

https://root.cern.ch

- **Species hits** (/Gy/Mbp)
  defined as the name of radical species together with the DNA reaction
  e.g. EaqStrandHits is e^-_{aq} + DNA backbone
- **Damage yield** (/Gy/Gbp)
  defined by DNA damage complexity
  (see classification scheme – previous slide)
- **Break yield** (/Gy/Gbp)
  shown for each break type
  (direct SSB, indirect SSB, DSB,…)
- **Fragment distribution of DNA**
  A fragment is part of DNA between two DSBs, with length equal to their separation distance.
- See more explanations at
  https://geant4-dna.github.io/molecular-docs/docs/overview/results-and-analysis







List of macro files & scripts (1/4)

- **Geant4 macro files** to **run the moleculardna** simulation (*.mac)
  - cylinders.mac
    - Simulates a parameter study based on the publication of *H. Nikjoo et al.*
      Can be used for regression testing of the different parameters, as well as for simulating a population of plasmids.
  - ecoli.mac
    - The geometry of an **E. coli bacterium** has been modeled and can be used to simulate early damage induction by irradiation.
      The length of the DNA contained in the bacterial cell is 4.63 Mbp.
  - human_cell.mac
    - A **human fibroblast cell** has been modeled and is included in this mac file.
      The length of the DNA included in this cell is ~6.4 Gbp.
    - One default model (keratinocyte) and **two alternative models** for HTB177 (lung) and MCF7 (breast) cancer cell lines (see K. Chatzipapas et al.)
- **ROOT macro files** to **analyze damage** results (*.C)
  - cylinders.C
    - Analysis of simulation results of the cylinders.mac file. Prints ***Break Source*** and ***Break Complexity*** frequency.
  - ecoli.C
    - Analysis of simulation results of the ecoli.mac file. Prints ***Species hits***, ***Damage yield***, ***Breaks yield*** and ***Fragments distribution***.
  - human_cell.C
    - Analysis of simulation results of the human_cell.mac file. Prints ***Species hits***, ***Damage yield***, ***Breaks yield*** and ***Fragments distribution***.
    - A specific version for the irradiation with alpha particles, of the geometries HTB177 and MCF7: human_cell_alphas.C
- Extras
  - createSDD.py
    - After the simulation, ROOT data can be converted to ***Standard DNA Damage*** (SDD) data format (see SDD format).
  - **repair_survival_models** folder
  - **phase-space** directory including information of incident particle sources

List of macro files & scripts (2/4)

- **Late damage** estimation with dedicated Python scripts (*.py) (1/3) : see **[1]**[2][3]
  - The **repair model** by **O. Belov**, considers **4 repair pathways** as presented in the function:
    - non-homologous end-joining (NHEJ),
    - homologous recombination (HR),
    - single-strand annealing (SSA), and
    - alternative end-joining mechanism (Alt-NHEJ)

$$\frac{dN_0}{dt} = a(L)\frac{dD}{dt}N_{cDSB} - V_{NHEJ} - V_{HR} - V_{SSA} - V_{microSSA}$$

  - To use the script, the procedure is typical for Python scripts. In a terminal the user can type:

```
python3 molecularDNArepair.py
```

  - **Several parameters** need to be defined:

```
iRootFile  = "/path/to/molecular-dna.root"
outputFile = "/path/to/molecularDNArepair.txt"
r3 = 7100*1e-09 * 2500*1e-09 * 7100*1e-09    # volume calculation, if ellipsoid cell
mass = 997 * 4 * 3.141592 * r3 / 3           # mass calculation, if ellipsoid cell
NBP = 6405886128      # length of the cellular DNA in base pairs
```


References:
[1] A quantitative model of the major pathways for radiation-induced DNA double-strand break repair, Belov OV, et al. J Theor Biol., Feb 7;366:115-30, 2015: link
[2] Performance Evaluation for Repair of HSGc-C5 Carcinoma Cell Using Geant4-DNA, D. Sakata et al., Cancers, 13, p. 6046, 2021: link
[3] Simulation of DNA damage using Geant4-DNA: an overview of the "molecularDNA" example application, Chatzipapas et al. Prec Radiat Oncol. 1–11. 2023: link

List of macro files & scripts (3/4)

- **Late damage** estimation with dedicated Python scripts (*.py) (2/3) : see **[1]**[2]
  - The **cell survival** model is based on the **two-lesion kinetics (TLK) model**. It includes kinetic processes of fast- and slow-DNA repair, and, based on lethal DNA damage, it can calculate the SF of a cell population.
  - Mathematically, the **Survival Fraction** (SF) of cells is calculated using:

$$SF(t) = ln(-L_f(t)) = ln\left(-\int_0^t (\beta_1\lambda_1 L_1(t) + \beta_2\lambda_2 L_2(t) + \gamma\eta[L_1(t) + L_2(t)]^2)dt\right)$$

    - $L_1(t)$ is the number of lesions per cell in the fast- repair process at a given time t after the beginning of the irradiation.
    - $L_2(t)$ is the number of lesions per cell in the slow- repair process at a given time t.
    - $L_f(t)$ is the number of lethal lesions that may lead to cell death at time t.
  - **Repair probability** coefficients, represent the rate of rejoined lesions (λ and η):
    - λ_1, λ_2, and η correspond to fast-, slow-, and binary- rejoining processes, respectively ($hour^{-1}$).
  - **Lethality probability** coefficients, represent the probability that a residual lesion may lead to cell death (β and γ):
    - β_1, β_2, and γ correspond to fast-, slow-, and binary- rejoining processes, respectively ($hour^{-1}$).

[1] Two-lesion kinetic model of double-strand break rejoining and cell killing, Stewart RD. Radiat Res. 2001: link
[2] Simulation of DNA damage using Geant4-DNA: an overview of the "molecularDNA" example application, Chatzipapas et al. Prec Radiat Oncol. 1–11. 2023: link

List of macro files & scripts (4/4)

$$SF(t) = ln(-L_f(t)) = ln\left(-\int_0^t (\beta_1\lambda_1 L_1(t) + \beta_2\lambda_2 L_2(t) + \gamma\eta[L_1(t) + L_2(t)]^2)dt\right)$$

- **Late damage** estimation with dedicated Python scripts (*.py) (3/3)
  - To use the tool, the procedure is typical for python scripts. In a terminal the user can type :

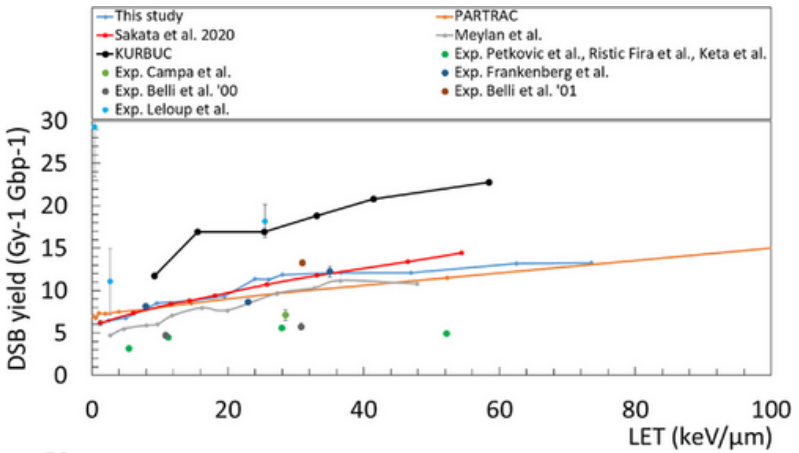
    ```
    python3 molecularDNAsurvival.py
    ```

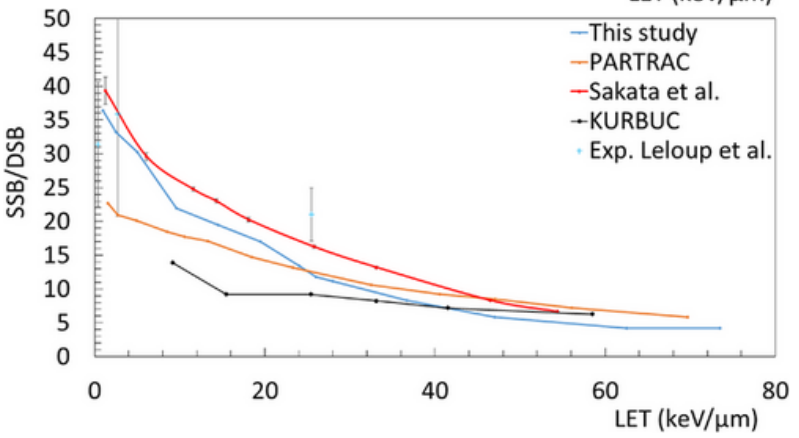
  - **Several parameters** need to be defined (indicative values):

    ```
    iRootFile  = "/path/to/molecular-dna.root"
    outputFile = "/path/to/molecularDNAsurvival.txt"
    r3 = 7100*1e-09 * 2500*1e-09 * 7100*1e-09  # volume calculation, if ellipsoid cell
    mass = 997 * 4 * 3.141592 * r3 / 3         # mass calculation, if ellipsoid cell
    NBP = 6405886128      # length of the cellular DNA in base pairs
    cell  = "test"        # name of the cell
    Lamb1 = 0.0125959     # indicative values of parameters  λ1, λ2, η, β1, β2, γ
    Lamb2 = 1
    Eta   = 7.50595e-06
    Beta1 = 0.0193207
    Beta2 = 0
    gamma = 0.189328
    ```

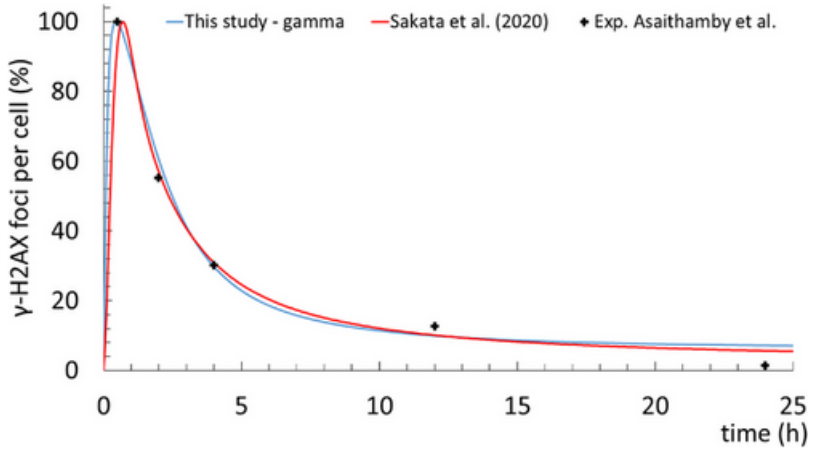
Examples of verification & validation

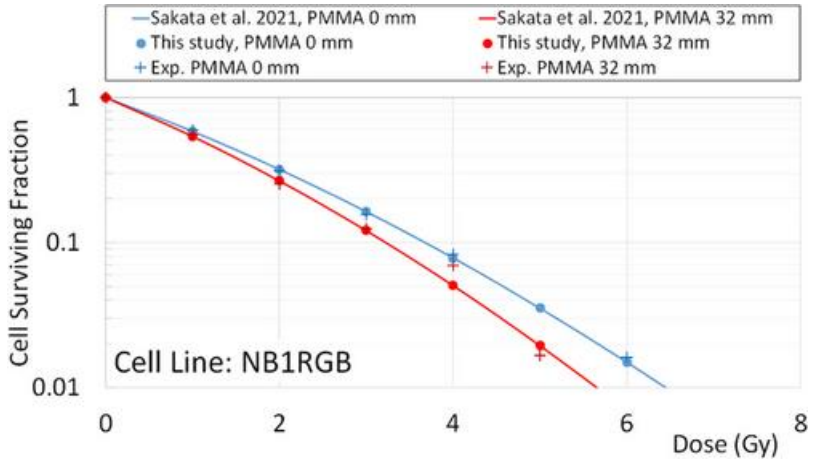
Proton irradiation: MC tools & measurements



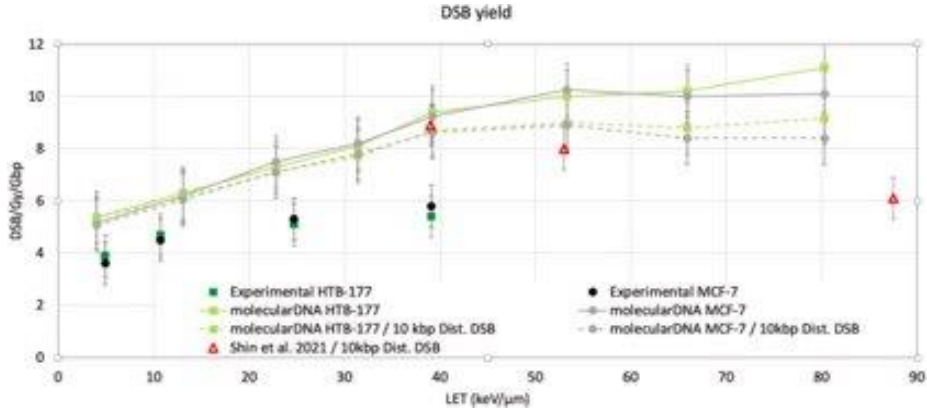


Damage repair and cell survival: moleculardna & measurements

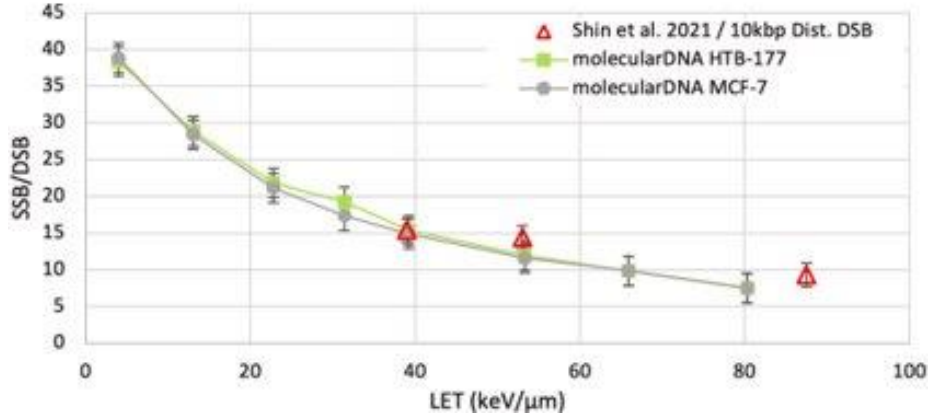




Alpha irradiation: moleculardna & measurements



SSB/DSB



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Geant4-DNA simulation of human cancer cells irradiation with helium ion beams

Konstantinos Chatzipapas [a], Milos Dordevic [b], Sara Zivkovic [b], Ngoc Hoang Tran [a], Nathanael Lampe [c], Dousatsu Sakata [d], Ivan Petrovic [b], Aleksandra Ristic-Fira [b], Wook-Geun Shin [e], Sara Zein [a], Jeremy M.C. Brown [f], Ioanna Kyriakou [g], Dimitris Emfietzoglou [g], Susanna Guatelli [h], Sebastien Incerti [a]

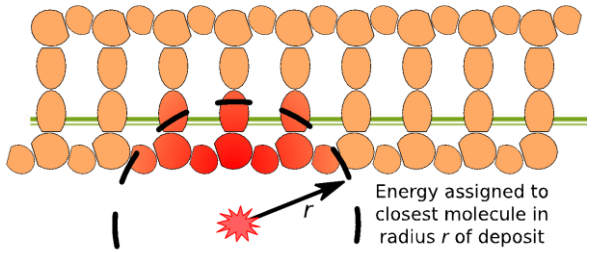
Important: parameter choice

Physics

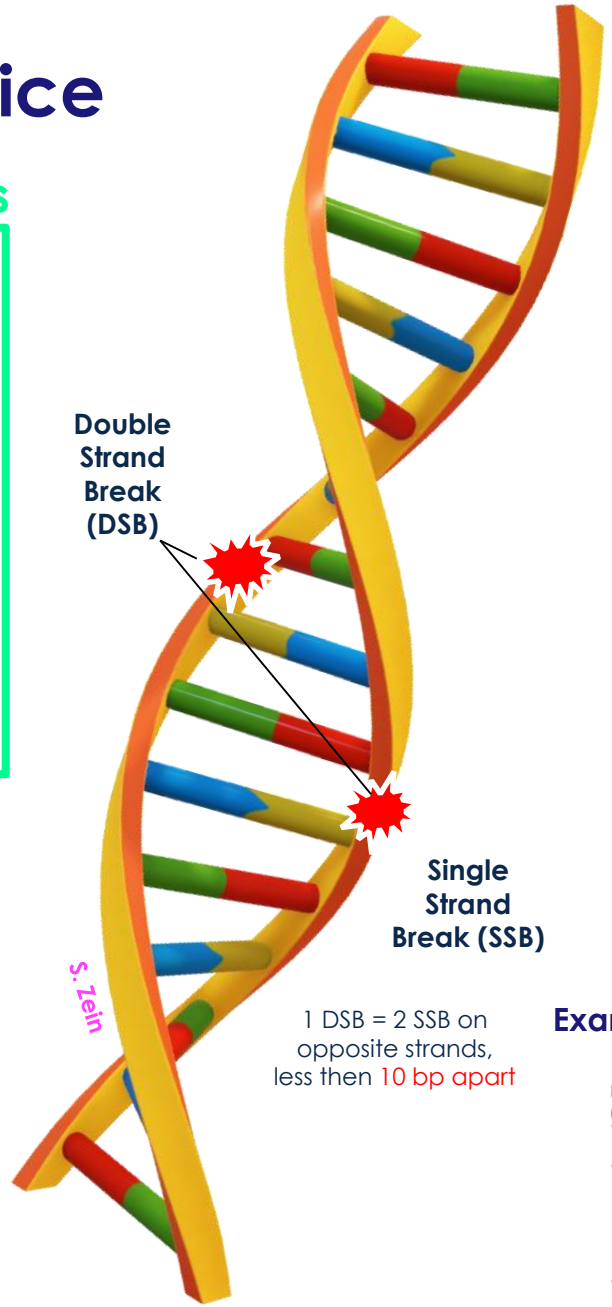
DIRECT damage induction

1. **Choice of G4DNA physics constructor**
2. **Volume for energy deposition scoring in DNA backbone (D or P molecule)**
3. **Probability of Single Strand Break induction in DNA backbone**
   - Threshold, linear…

Example, for item 2. :



E.g. 6 A° for ~3 hydration sh.



1 DSB = 2 SSB on opposite strands, less then 10 bp apart

Chemistry

NON-DIRECT damage induction

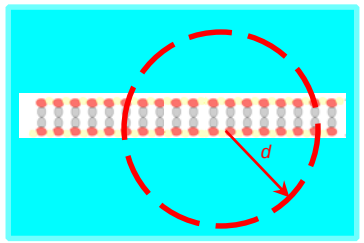
1. **Choice of G4DNA chemistry constructor**
   - Including reactions with DNA components
2. **Probability of non-direct SSB induction**
   - •OH on DNA backbone : e.g. 40.5 %
3. **Distance from DNA to kill radicals (mimic scavenging in cells)**
4. **Histones considered as full scavengers (in cells)**
5. **Radiolysis maximum time steps**
6. **Chemical stage end time**

Example, for item 1. :

Reaction rates used between radicals and DNA components ($\times 10^9$ L mol^{-1} s^{-1}), from Buxton et al. [65].

	$^{\cdot}OH$	$H^{\cdot}$	e^-_{aq}
$C_6H_5O_6P$	1.8	0.029	0.01
Adenine	6.1	0.10	9.0
Thymine	6.4	0.57	18.0
Guanine	9.2	–	14.0
Cytosine	6.1	0.092	13.0

Example, for item 3. :



Content of macro file: e.g. human_cell.mac

```
/process/dna/e-SolvationSubType Meesungnoen2002
#/process/dna/e-SolvationSubType Ritchie1994
#/process/dna/e-SolvationSubType Terrisol1990
```

Physics

```
/run/verbose 1
/control/verbose 1

/world/worldSize 50 um
/cell/radiusSize 14 2.5 14 um

/scheduler/endTime 5.0 ns
/scheduler/maxNullTimeSteps 10000000
```

Chemistry

endTime aims to simulate the radical scavenging that is considered as the radical time life. Beyond this time point radicals are scavenged by the biological medium

```
/dnageom/radicalKillDistance 9 nm
/dnageom/interactionDirectRange 3.5 angstrom
```

Geometry (1)

```
/dnageom/placementSize 75 75 75 nm
/dnageom/fractalScaling 75 75 75 nm
/dnageom/definitionFile geometries/cube-centred-X-8.txt
/dnageom/placementVolume turn geometries/turned_solenoid_750_withHistone.txt
/dnageom/placementVolume turntwist geometries/turned_twisted_solenoid_750_withHistone.txt true
/dnageom/placementVolume straight geometries/straight_solenoid_750_withHistone.txt
```

Geometry (2)

```
/dnadamage/directDamageLower 5 eV
/dnadamage/directDamageUpper 37.5 eV

/dnadamage/indirectOHBaseChance 1.0
/dnadamage/indirectOHStrandChance 0.405
/dnadamage/inductionOHChance 0.00

/dnadamage/indirectHBaseChance 1.0
/dnadamage/indirectHStrandChance 0.0
/dnadamage/inductionHChance 0.00

/dnadamage/indirectEaqBaseChance 1.0
/dnadamage/indirectEaqStrandChance 0.0
/dnadamage/inductionEaqChance 0.00
```

Damage

Geometry (3)

```
/chromosome/add cell ellipse 7100 2500 7100 0 0 0 nm 0 0 0
#/chromosome/add cell sphere 3000 0 0 0 nm

/run/initialize
/run/printProgress 10

# Source Geometry
/gps/pos/type Plane
/gps/pos/shape Circle
/gps/pos/centre 0 3000 0 nm
/gps/pos/rot1 0 0 1
/gps/pos/rot2 1 0 0
/gps/pos/radius 7100 nm
/gps/direction 0 -1 0

# Source Energy
/gps/particle  e-
/gps/energy 0.662 MeV

#/analysisDNA/fileName 50MeV

/run/beamOn 2
```

Output file name

UI commands of moleculardna (1/4)

Geometry related commands (1/2)

- **`/world/worldSize <s> <unit>`**
  Side length for the world.
- **`/dnageom/setVerbose <int>`**
  Print verbose debugging information related to the DNA geometry.
- **`/dnageom/definitionFile <filepath>`**
  Path to file that defines placement locations.
- **`/dnageom/placementVolume <name> <filepath> [<twist>]`**
  Set a placement volume, twist is an optional boolean parameter (written as true or false.
- **`/dnageom/fractalScaling <x> <y> <z> <unit>`**
  Scaling and units for the fractal along each axis.
- **`/dnageom/placementSize <x> <y> <z> <unit>`**
  Side length for each placement.
- **`/dnageom/checkOverlaps <bool>`**
  Check overlaps of molecules and fractal placements being placed for debugging.

UI commands of moleculardna (2/4)

Geometry related commands (2/2)

- **`/dnageom/setSmartVoxels <int>`**
  Change the amount of voxelisation in the Geant4 geometry optimisation for a faster simulation initialisation, but slower overall simulation (1 refers to maximal optimisation in initialisation).

- Chromosomes can be added to define regions of interest. For all chromosome types, a name is required. The x, y and z variables refer to the translation of the chromosome, and the optional rotations in x, y and z are Euler rotations.
  - **`/chromosome/add sphere <name> <rad> <x> <y> <z> <unit> [<rx> <ry> <rz>]`**
    Add a spherical chromosome with a specified radius.
  - **`/chromosome/add cyl <name> <rad> <height> <x> <y> <z> <unit> [<rx> <ry> <rz>]`**
    Add a cylindrical chromosome with a specified height and radius.
  - **`/chromosome/add rod <name> <rad> <height> <x> <y> <z> <unit> [<rx> <ry> <rz>]`**
    Add a rod-shaped chromosome. This is a cylinder of a specified height, with two hemispherical end caps. The radius of the cylinder and end caps is specified.
  - **`/chromosome/add ellipse <name> <sx> <sy> <sz> <x> <y> <z> <unit> [<rx> <ry> <rz>]`**
    Add an ellipsoidal chromosome, with semi-major axes <sx> <sy> and <sz>.

- **`/chromosome/plotData <filename>`**
  Save a scatter plot (x,y,z data points) of all chromosome positions.

UI commands of moleculardna (3/4)

Damage related commands (1/2)

- **`/dnageom/interactionDirectRange <d> <unit>`**
  Distance from DNA molecule at which energy deposits count towards DNA damage.
- **`/dnageom/radicalKillDistance <d> <unit>`**
  Distance from DNA at which to stop tracking radicals.
- **`/dnadamage/directDamageLower <d> <unit>`**
  Minimum Energy required for an SSB.
- **`/dnadamage/directDamageUpper <d> <unit>`**
  Maximum energy required for an SSB to occur.
- **`/dnadamage/indirectOHBaseChance <d>`**
  Chance ∈ [0,1] of a •OH damaging a base.
- **`/dnadamage/indirectOHStrandChance <d>`**
  Chance ∈ [0,1] of a •OH damaging a sugar-phosphate moiety.
- **`/dnadamage/inductionOHChance <d>`**
  Chance ∈ [0,1] of a reaction between a base and •OH yielding a strand break.

UI commands of moleculardna (4/4)

Damage related commands (2/2)

- **`/dnadamage/indirectHBaseChance <d>`**
  Chance ∈ [0,1] of a H• damaging a base.
- **`/dnadamage/indirectHStrandChance <d>`**
  Chance ∈ [0,1] of a H• damaging sugar-phosphate moiety.
- **`/dnadamage/inductionHChance <d>`**
  Chance ∈ [0,1] of a reaction between a base and H• yielding a strand break.
- **`/dnadamage/indirectEaqBaseChance <d>`**
  Chance ∈ [0,1] of a e^-_{aq} damaging a base.
- **`/dnadamage/indirectEaqStrandChance <d>`**
  Chance ∈ [0,1] of a e^-_{aq} damaging sugar-phosphate moiety.
- **`/dnadamage/inductionEaqChance <d>`**
  Chance ∈ [0,1] of a reaction between a base and e^-_{aq} yielding a strand break.
- **`/scheduler/endTime <d> <unit>`**
  End time of the simulation (related to the chemical part).

Hands-on

Hands-on practice with the « moleculardna » extended example

- **Copy** the moleculardna extended example to your local directory, **create** your build directory and **compile** moleculardna
  - `cd`
  - `cp -R $G4EXAMPLES/examples/extended/medical/dna/moleculardna .`
  - `cd moleculardna`
  - `mkdir build`
  - `cd build`
  - `cmake ..`
  - `make`

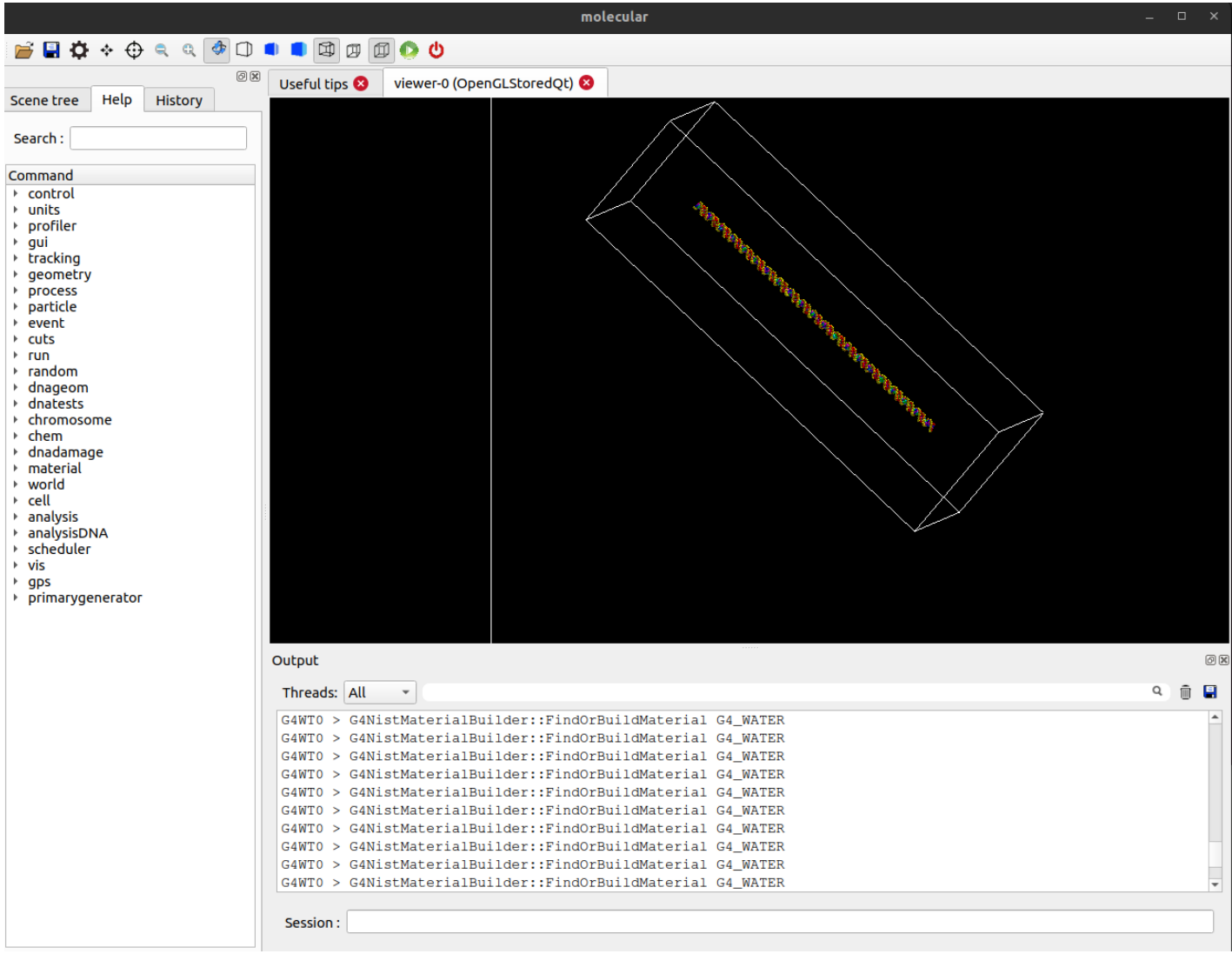
`make -jN` if you have N cores

The example needs internet to download geometry library and be installed

```
[  2%] Building CXX object CMakeFiles/molecular.dir/molecular.cc.o
[  5%] Building CXX object CMakeFiles/molecular.dir/src/ActionInitialization.cc.o
[  8%] Building CXX object CMakeFiles/molecular.dir/src/AnalysisManager.cc.o
[ 11%] Building CXX object CMakeFiles/molecular.dir/src/AnalysisMessenger.cc.o
[ 13%] Building CXX object CMakeFiles/molecular.dir/src/ChemistryList.cc.o
[ 16%] Building CXX object CMakeFiles/molecular.dir/src/ChromosomeFactory.cc.o
[ 19%] Building CXX object CMakeFiles/molecular.dir/src/ChromosomeHit.cc.o
[ 22%] Building CXX object CMakeFiles/molecular.dir/src/ChromosomeMapper.cc.o
[ 25%] Building CXX object CMakeFiles/molecular.dir/src/ChromosomeMessenger.cc.o
[ 27%] Building CXX object CMakeFiles/molecular.dir/src/CylindricalChromosome.cc.o
[ 30%] Building CXX object CMakeFiles/molecular.dir/src/DNAGeometry.cc.o
[ 33%] Building CXX object CMakeFiles/molecular.dir/src/DNAGeometryMessenger.cc.o
[ 36%] Building CXX object CMakeFiles/molecular.dir/src/DNAHashing.cc.o
[ 38%] Building CXX object CMakeFiles/molecular.dir/src/DNAHit.cc.o
[ 41%] Building CXX object CMakeFiles/molecular.dir/src/DNAWorld.cc.o
[ 44%] Building CXX object CMakeFiles/molecular.dir/src/DamageModel.cc.o
[ 47%] Building CXX object CMakeFiles/molecular.dir/src/DamageModelMessenger.cc.o
[ 50%] Building CXX object CMakeFiles/molecular.dir/src/DetectorConstruction.cc.o
[ 52%] Building CXX object CMakeFiles/molecular.dir/src/DetectorMessenger.cc.o
[ 55%] Building CXX object CMakeFiles/molecular.dir/src/EllipticalChromosome.cc.o
[ 58%] Building CXX object CMakeFiles/molecular.dir/src/EventAction.cc.o
[ 61%] Building CXX object CMakeFiles/molecular.dir/src/IRTDamageReactionModel.cc.o
[ 63%] Building CXX object CMakeFiles/molecular.dir/src/OctreeNode.cc.o
[ 66%] Building CXX object CMakeFiles/molecular.dir/src/ParallelWorldPhysics.cc.o
[ 69%] Building CXX object CMakeFiles/molecular.dir/src/PhysicsList.cc.o
[ 72%] Building CXX object CMakeFiles/molecular.dir/src/PlacementVolumeInfo.cc.o
[ 75%] Building CXX object CMakeFiles/molecular.dir/src/PrimaryGeneratorAction.cc.o
[ 77%] Building CXX object CMakeFiles/molecular.dir/src/RodChromosome.cc.o
[ 80%] Building CXX object CMakeFiles/molecular.dir/src/RunAction.cc.o
[ 83%] Building CXX object CMakeFiles/molecular.dir/src/SphericalChromosome.cc.o
[ 86%] Building CXX object CMakeFiles/molecular.dir/src/StackingAction.cc.o
[ 88%] Building CXX object CMakeFiles/molecular.dir/src/SteppingAction.cc.o
[ 91%] Building CXX object CMakeFiles/molecular.dir/src/TimeStepAction.cc.o
[ 94%] Building CXX object CMakeFiles/molecular.dir/src/UtilityFunctions.cc.o
[ 97%] Building CXX object CMakeFiles/molecular.dir/src/VirtualChromosome.cc.o
[100%] Linking CXX executable molecular
[100%] Built target molecular
```

Hands-on practice with the « moleculardna » extended example

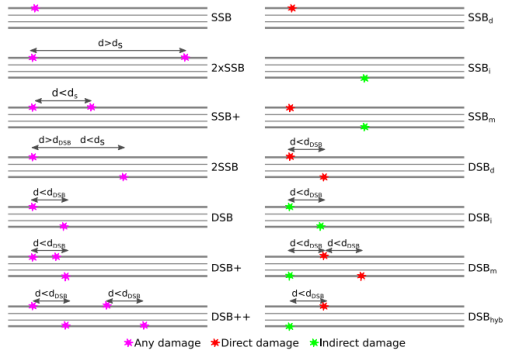
- Run moleculardna in **interactive mode** with GUI commands
  - `./molecular`
- Using `/control/execute` command, you can run any example in interactive mode.
- Be careful on the RAM used.

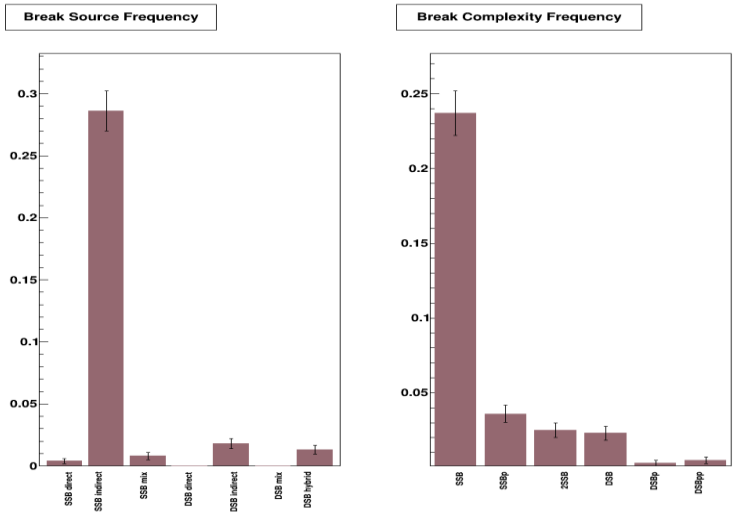


Hands-on practice with the « moleculardna » extended example

- Run moleculardnas in **batch mode** using a macro file
  - `./molecular -m cylinders.mac -p 4 -t 1`
  - You can run for 1000 electrons of 4.5 keV
  - No visualization by default
  - Results are saved in molecular-dna_t0.root file
- Results can be analyzed using ROOT (depending on the version these commands my differ)
  - `root`
  - ROOT is already installed on your Geant4 virtual machine
  - `.X cylinders.C`
  - Histograms will be plotted

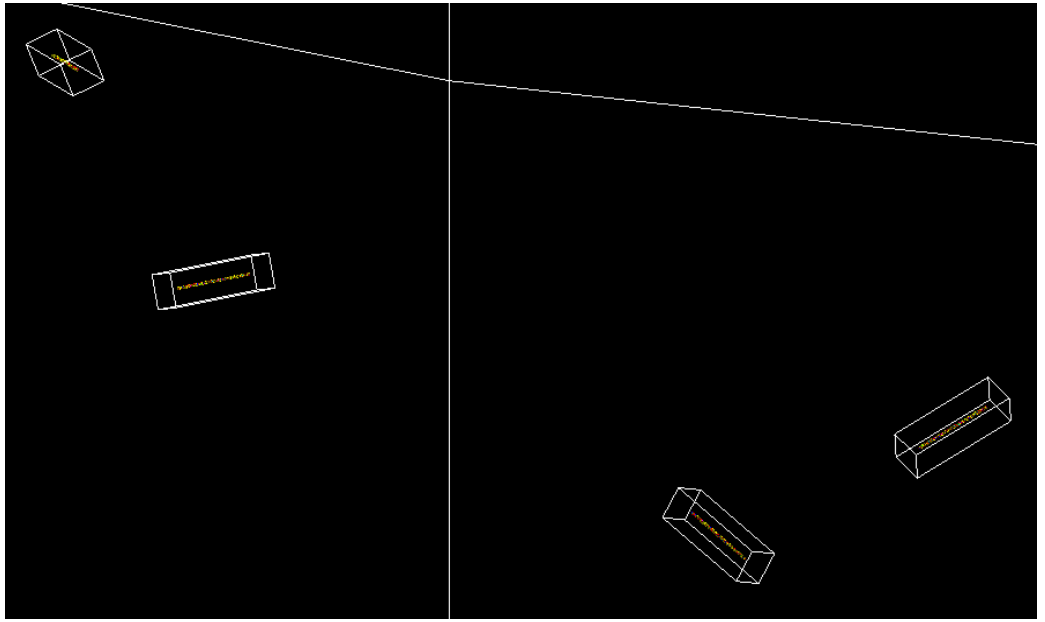
```
konstantinos@kc64:~/software/testfolder/moleculardna/build$ root
   ------------------------------------------------------------------
  | Welcome to ROOT 6.26/10                        https://root.cern |
  | (c) 1995-2021, The ROOT Team; conception: R. Brun, F. Rademakers |
  | Built for linuxx8664gcc on Nov 16 2022, 10:42:54                 |
  | From tags/v6-26-10@v6-26-10                                      |
  | With c++ (Ubuntu 11.3.0-1ubuntu1~22.04) 11.3.0                   |
  | Try '.help', '.demo', '.license', '.credits', '.quit'/'.q'       |
   ------------------------------------------------------------------

root [0] .X cylinders.C
hadd Target file: molecular-dna.root
hadd compression setting for all output: 1
hadd Source file 1: molecular-dna_t0.root
hadd Target path: molecular-dna.root:/
hadd Target path: molecular-dna.root:/hists
hadd Target path: molecular-dna.root:/tuples
Paricle : e     Energy [/MeV] : 0.0045  number : 1000
 Output Damages :
       SSB direct : 0.004       error : 0.00199699
       SSB indirect : 0.286     error : 0.0162008
       SSB mix : 0.008  error : 0.0028185

       DSB direct : 0   error : 0
       DSB indirect : 0.018     error : 0.00420639
       DSB mix : 0      error : 0
       DSB hybrid : 0.013       error : 0.00358383

       SSB  : 0.237     error : 0.0148003
       SSBp : 0.036     error : 0.00589396
       2SSB : 0.025     error : 0.00493957

       DSB : 0.023      error : 0.00474273
       DSBp : 0.003     error : 0.00173032
       DSBpp : 0.005    error : 0.00223159
```

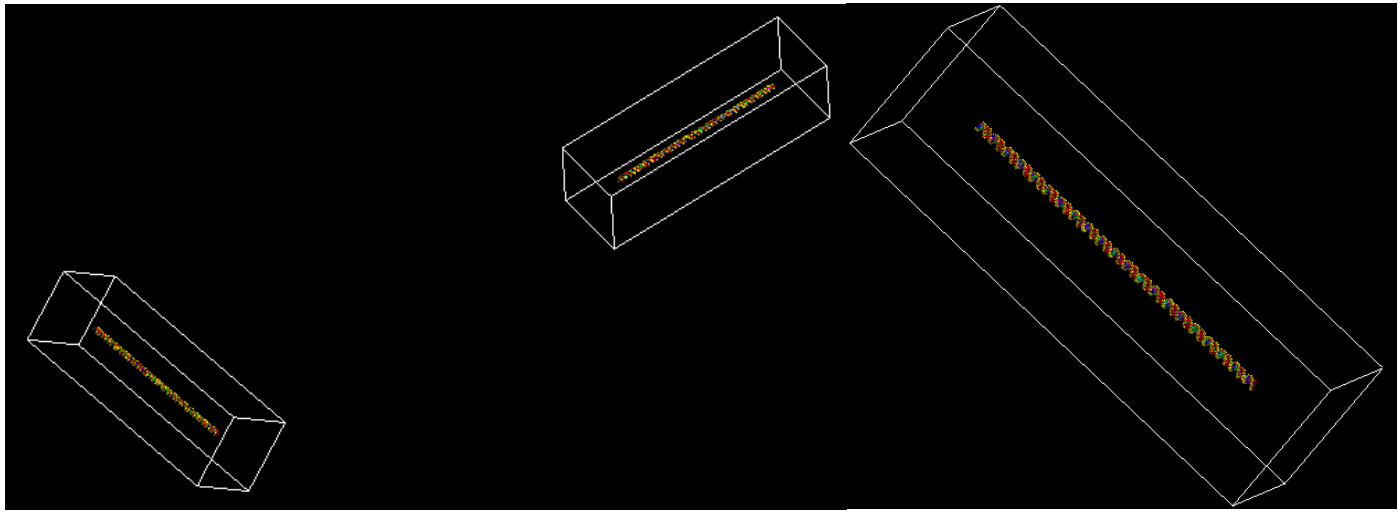




Hands-on practice with the « moleculardna » extended example

1. Open molecular.cc
   - Go to line 115 and comment it
2. Open DetectorConstruction.cc
   - Go to line 51 and enable « DNAWorld »
     - G4bool useParallelPhysicsWorld = true;
3. Open PhysicsList.cc
   - Go to line 78 and enable « DNAWorld »
     - G4bool useParallelPhysicsWorld = true;
4. Recompile moleculardna (Do `make`)
5. Create a copy of prisms200k_r3000.txt
6. Open prisms200k_r3000.txt:
   - Keep only the first 50 prisms (0-49)(up-to line 51).
7. Open cylinders.mac:
   - Comment the last line: `/run/beamOn`
   - Un-comment line /control/execute vis.mac
   - Replace prisms200k_r3000.txt, to the new one and save
8. In the terminal type `./molecular`
9. In the Qt window type: `/control/execute cylinders.mac`
10. Observe DNA geometry !

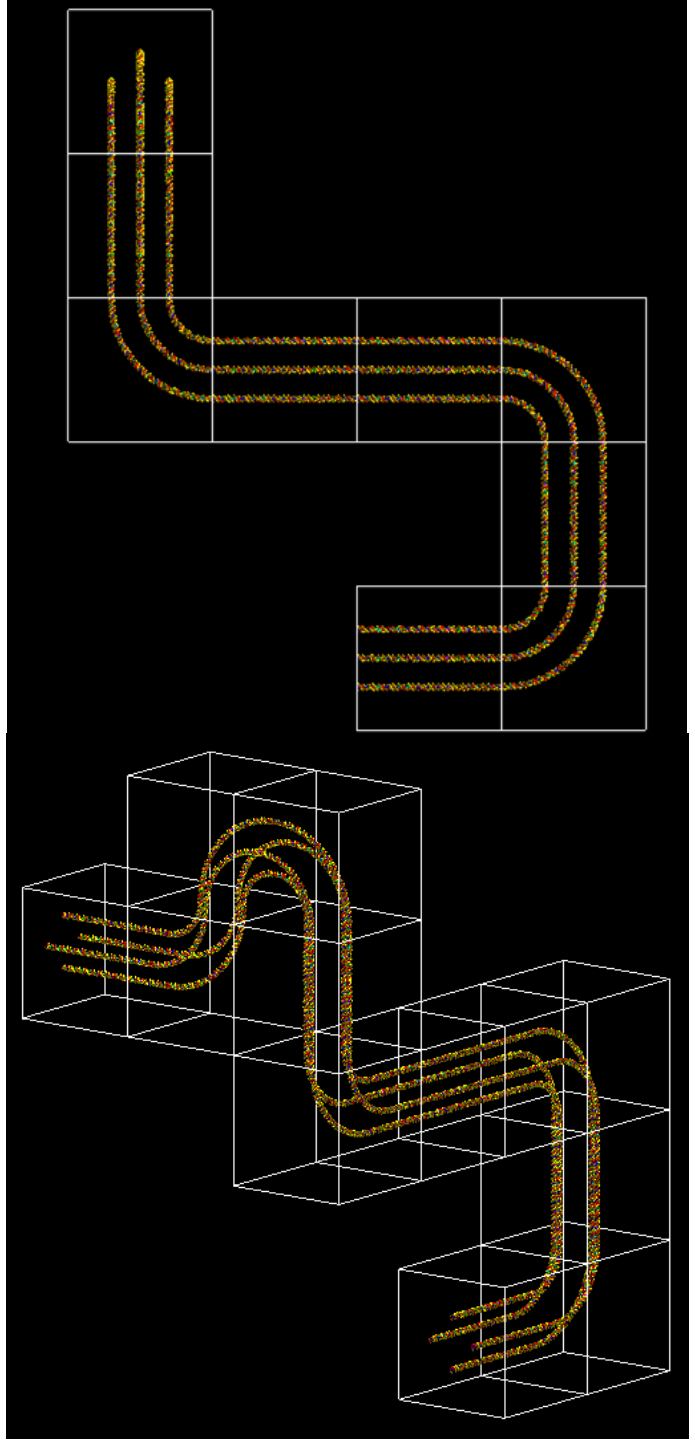




Hands-on practice with the « moleculardna » extended example

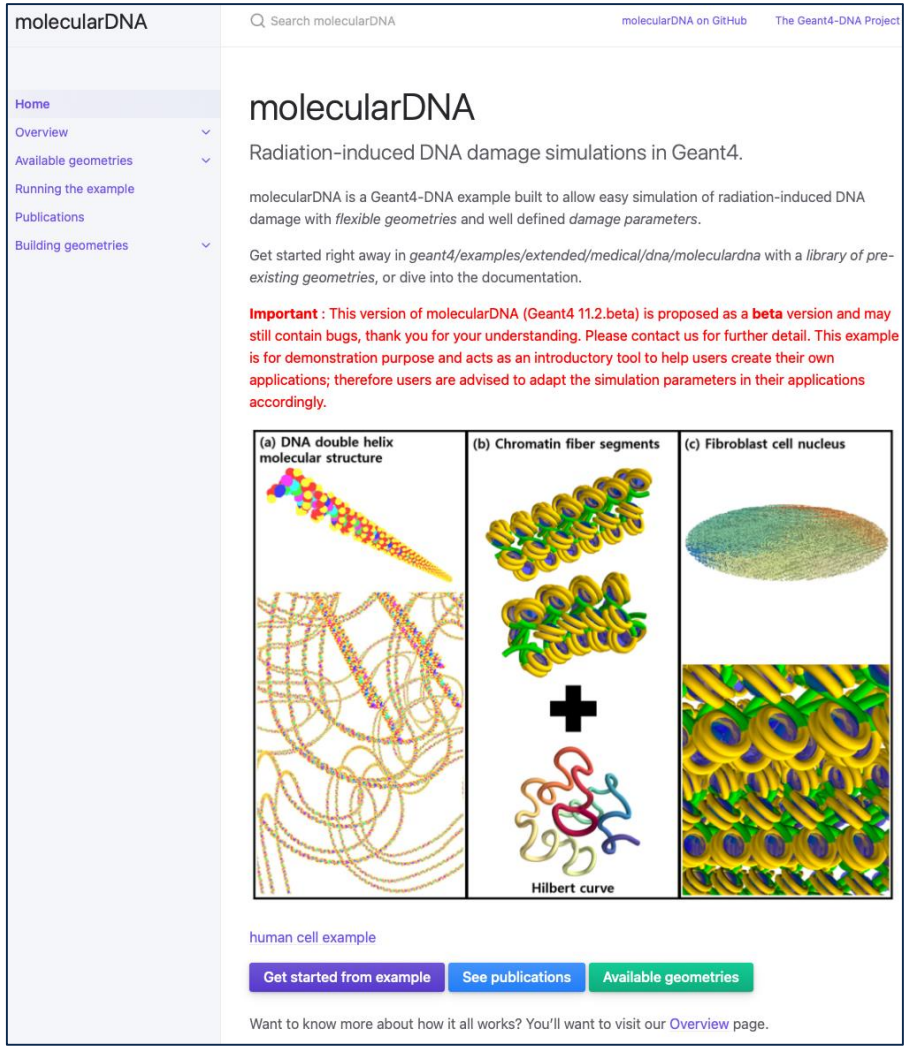
- Using 4strands_50nm_straight.txt and 4strands_50nm_turn.txt, create a new geometry like the one shown in the following images.
  Use also cylinders.mac as a starting point.
- The following lines are given to help you…
  - `/dnageom/placementSize 50 50 50 nm`
  - `/dnageom/fractalScaling 1 1 1 nm`
  - `/dnageom/definitionFile geometries/newGeometry.txt`
  - `/dnageom/placementVolume straight geometries/4strands_50nm_straight.txt`
  - `/dnageom/placementVolume turn geometries/4strands_50nm_turn.txt`

  - `# IDX TYPE POS_X POS_Y POS_Z EUL_PSI EUL_THETA EUL_PHI`
  - `0 straight 0 0 0 0 0 0`
  - `1 turn 0 0 50 0 0 1.5708`
  - `2 straight 0 50 50 1.5708 0 0`
  - `3 turn 0 100 50 0 1.5708 1.5708`

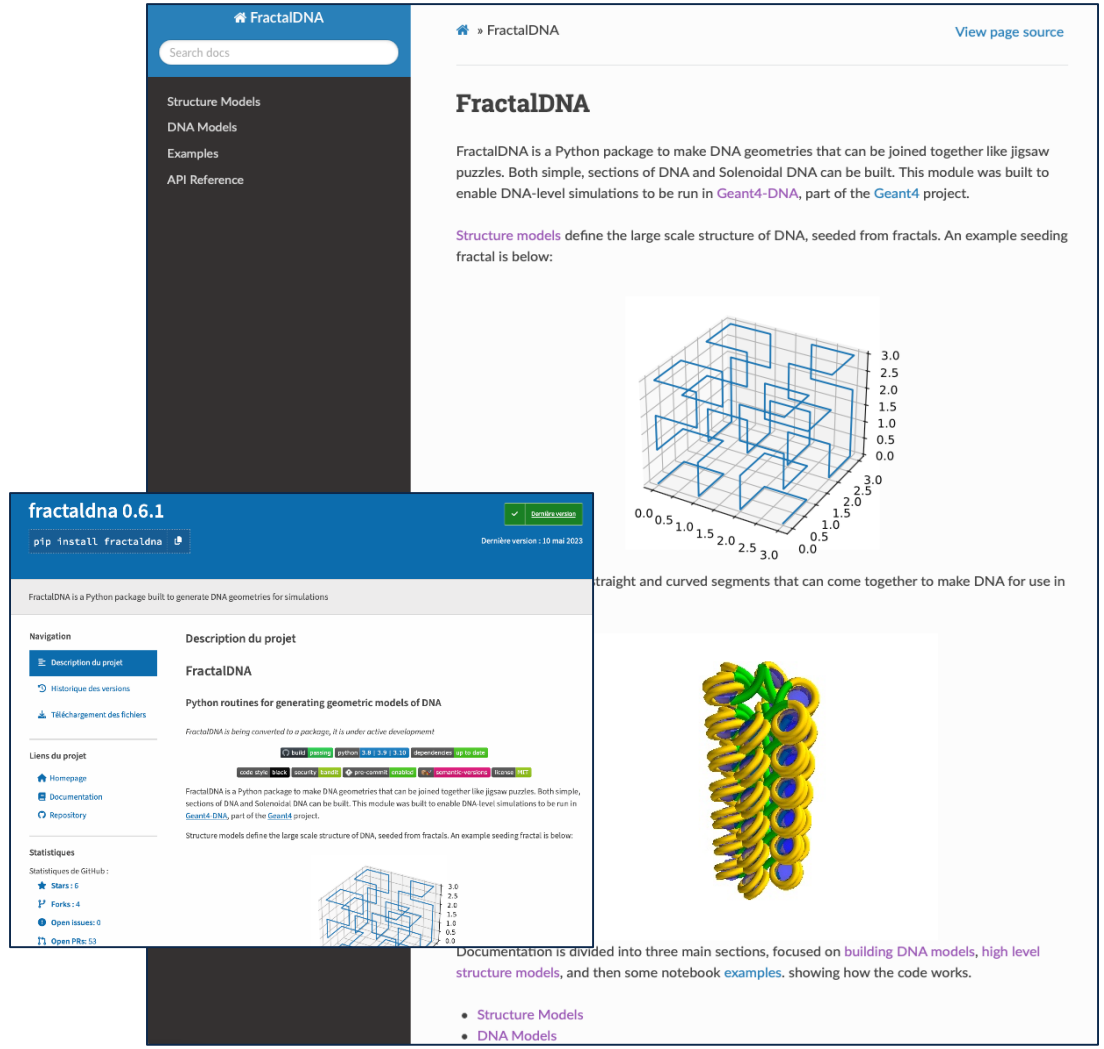


Documentation

moleculardna documentation
https://geant4-dna.github.io/molecular-docs/



FractalDNA documentation
https://pypi.org/project/fractaldna/
http://natl.github.io/fractaldna/



Main publications

- Geant4-DNA simulation of human cancer cells irradiation with helium ion beams, K. Chatzipapas et al., Phys. Med. 112 (2023) 102613 (link)
- Simulation of DNA damage using Geant4-DNA: an overview of the "molecularDNA" example application, K. Chatzipapas et al., Prec. Radiat. Oncol. (2023) 1–11 (link)
- A Geant4-DNA evaluation of radiation-induced DNA damage on a human fibroblast, W.-G. Shin et al., Cancers 13 (2021) 4940 (link)
- Fully integrated Monte Carlo simulation for evaluating radiation induced DNA damage and following repair using Geant4-DNA, D. Sakata et al., Sc. Rep. 10 (2020) 20788 (link)
- Evaluation of early radiation DNA damage in a fractal cell nucleus model using Geant4-DNA, D. Sakata et al., Phys. Med. 62 (2019) 152-157 (link)
- Mechanistic DNA Damage Simulations in Geant4-DNA Part 2: Electron and Proton Damage in a Bacterial Cell, N. Lampe et al., Phys. Med. 48 (2018) 146-155 (link)
- Mechanistic DNA Damage Simulations in Geant4-DNA Part 1: A parameter study in a simplified geometry, N. Lampe et al., Phys. Med. 48 (2018) 135-145 (link)

http://geant4-dna.org
http://geant4.in2p3.fr

More information

http://geant4-dna.org