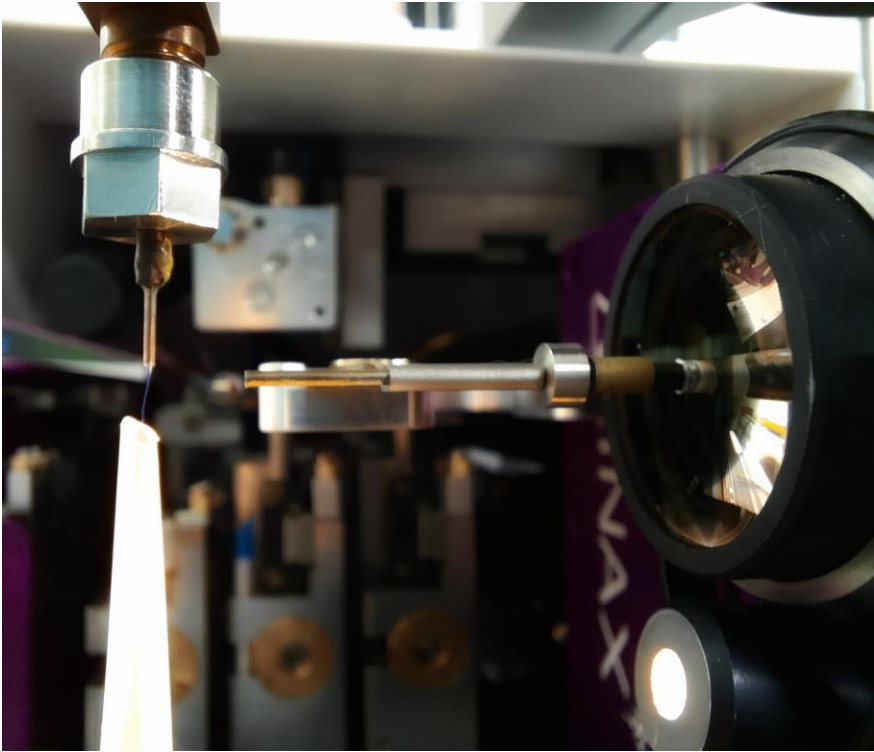


# LEAPS MEETS LIFE SCIENCES CONFERENCE



**New opportunities for time-resolved serial crystallography at the new ID29 from the ESRF extremely brilliant source**

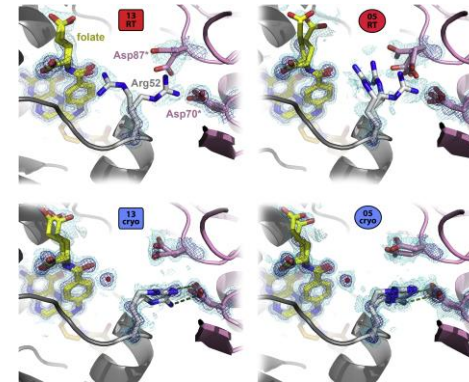
**Julien ORLANS (ESRF ID29)**

# ROOM-TEMPERATURE DATA COLLECTION

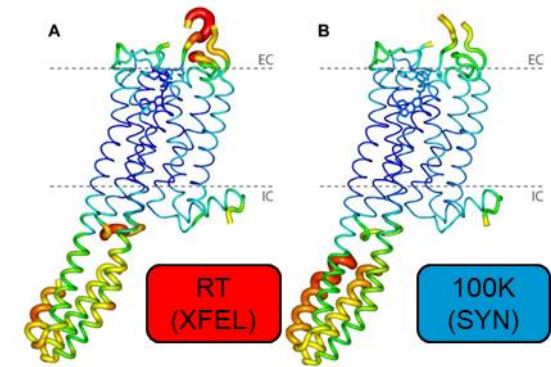
Cryo-structures do not display the same range of conformations as the room-temperature structures:

- Might hide **functional conformations**
- Might prevent **binding of substrates or inhibitors**
- Inhibit **thermal motion** closer to “native” conditions

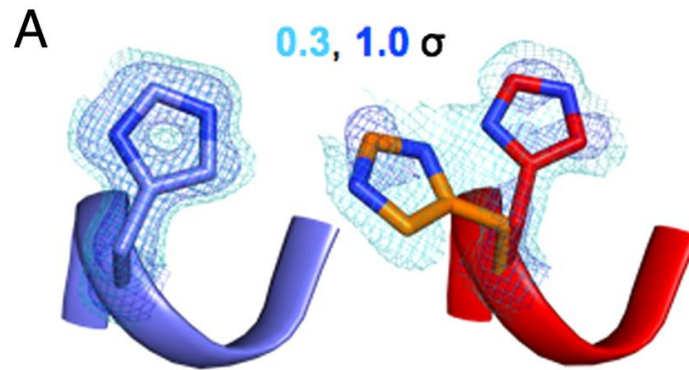
Room-temperature temperature crystal structures reveal physiologically relevant conformations “hidden” at 100 K.



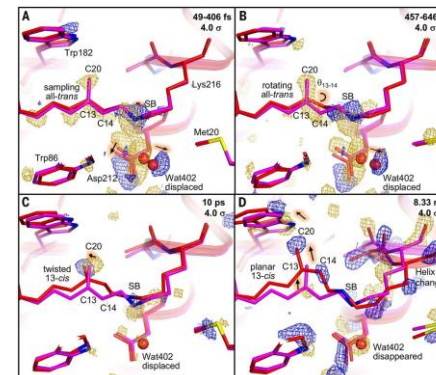
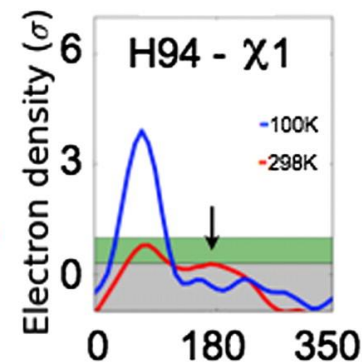
Keedy *et al.*, Structure (2014)



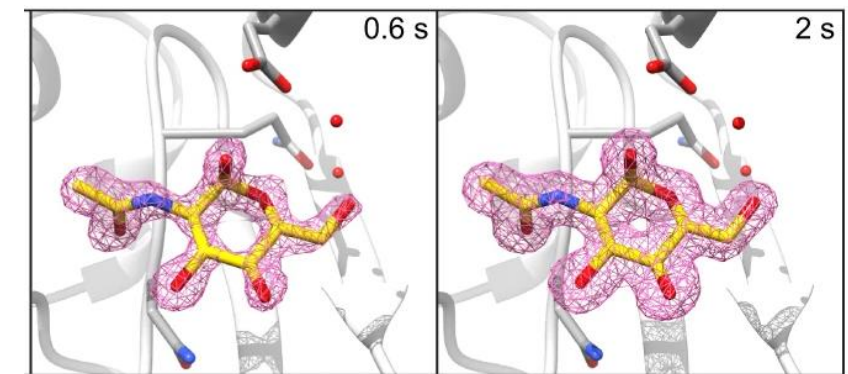
Liu *et al.*, Science (2013)



Fraser *et al.*, PNAS (2011)



Nogly *et al.*, Science (2018)

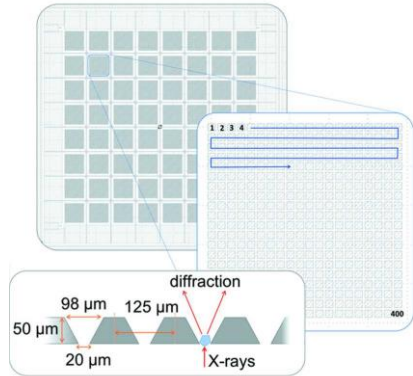


Butryn *et al.*, Nat. Comm. (2021)

Because of radiation damage, **serial crystallography** is the most valuable route to obtaining room-temperature structures.  
(1 crystal = 1 diffraction pattern)

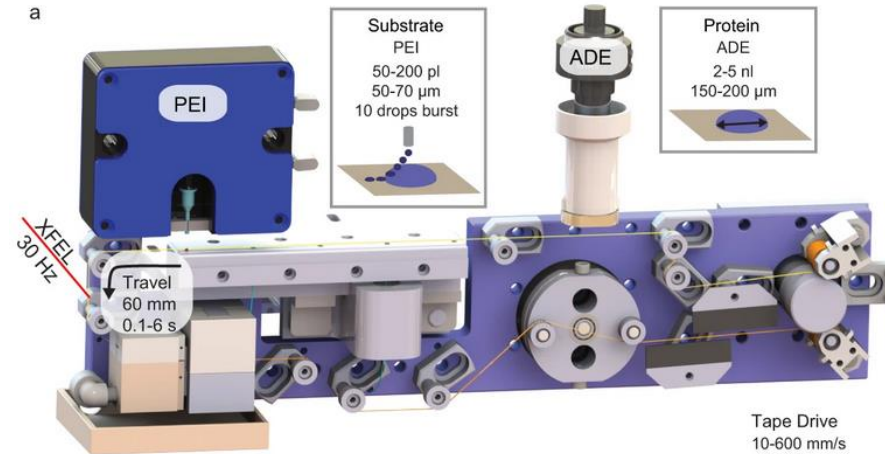
## There are 4 family of sample delivery methods:

### Fixed-Target



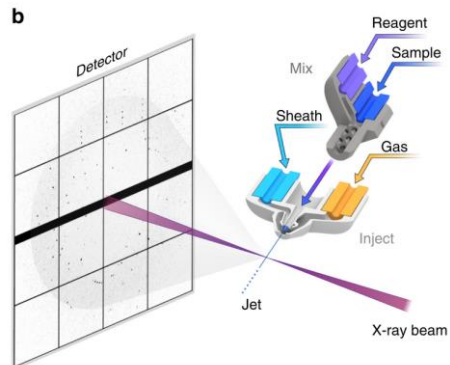
Ebrahim *et al.*, IUCrJ (2019)

### Hybrid (Tape-Drive)

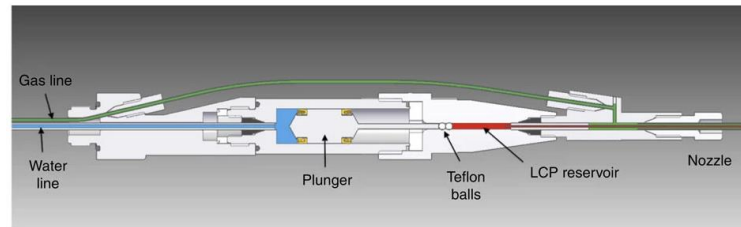


Butryn *et al.*, Nat. Comm. (2021)

### Liquid and Viscous Injectors

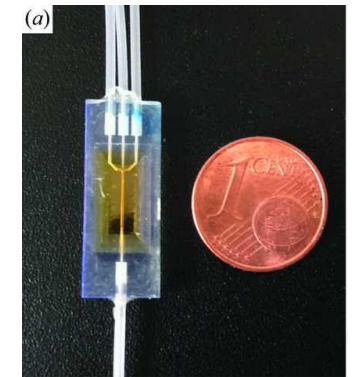


Knoška *et al.*, Nat. Comm. (2020)



Weierstall *et al.*, Nat. Comm. (2014)

### Microfluidics



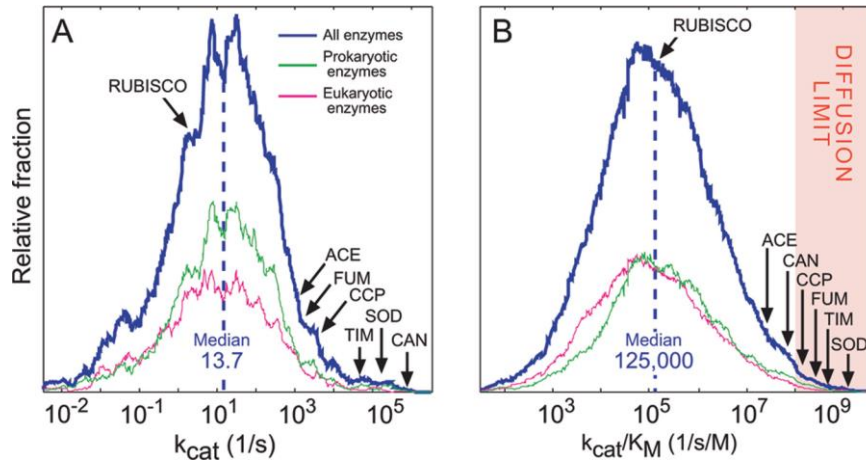
Monteiro *et al.*, IUCrJ (2019)

# ENZYME TIME-SCALE

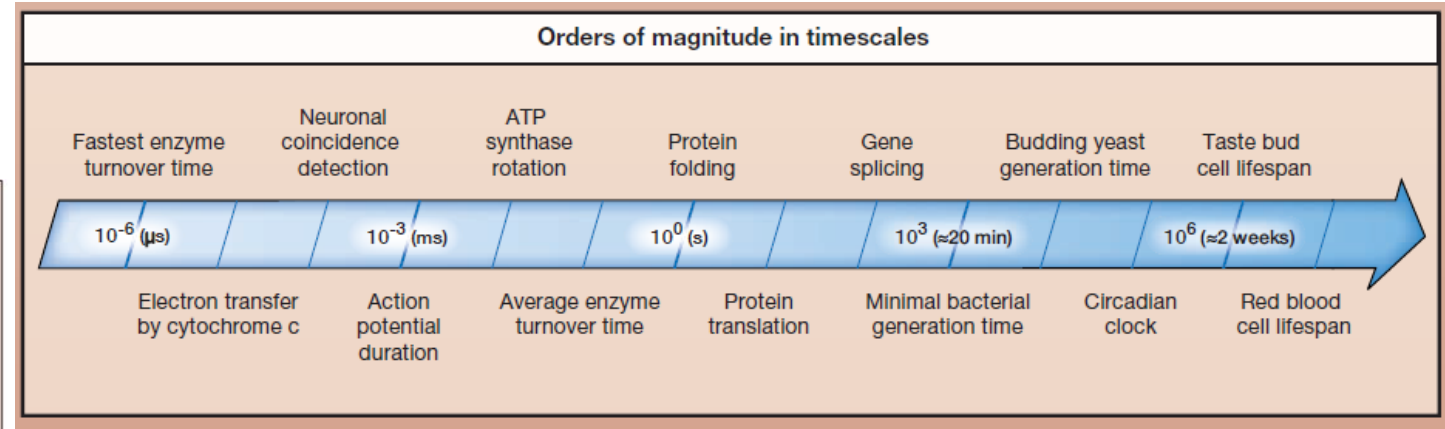
The median turnover time for catalysis in solution is about **70 ms**

More than **60%** of enzymatic reactions exhibit a  $k_{cat}$  value between **1 and 100 s<sup>-1</sup>**

Enzymes catalyzing reactions related to secondary metabolism are typically **30-fold** slower than those of central metabolism



Bar-Even *et al.*, *Biochemistry* (2011)



Shamir *et al.*, *Cell* (2016)

To observe molecular mechanism *in-crystallo*, the reaction can be activated by:

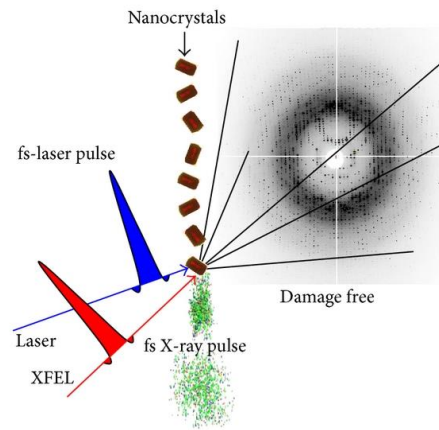
- Photoactivation
- Mixing



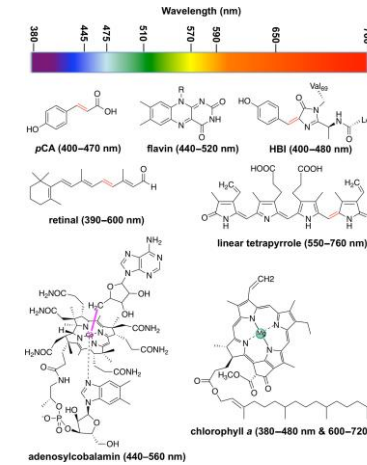
# PROTEIN DYNAMICS IN CRYSTALS

## Pump & probe

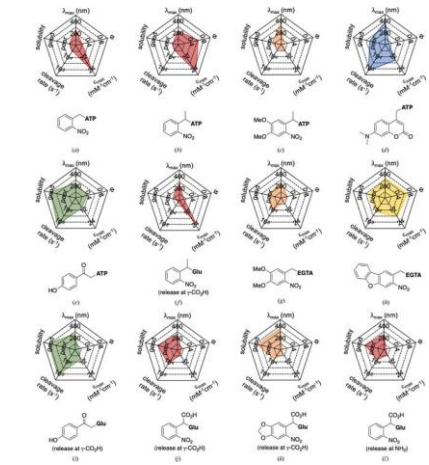
- Photoactivatable proteins (UV-vis)
  - Cleavable cage compounds
  - Activatable cofactors
- Temperature jumps (IR)



Schmidt, Advances in Condensed Matter Physics (2013)



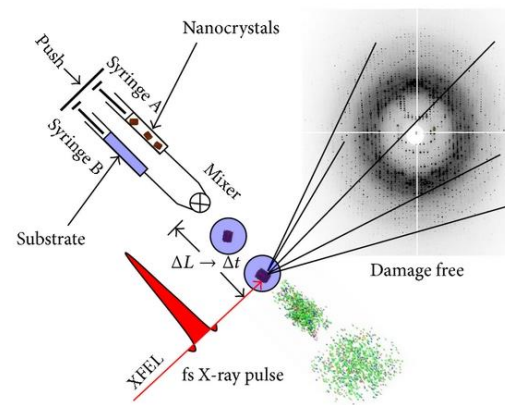
Poddar *et al.*, FEBS (2021)



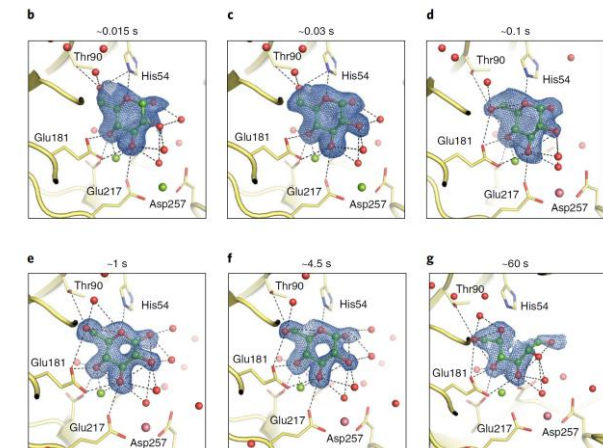
Monteiro *et al.*, Acta D. (2021)

## Ligand and buffer mixing

- Drug binding
- Enzymatic reaction
- pH jump

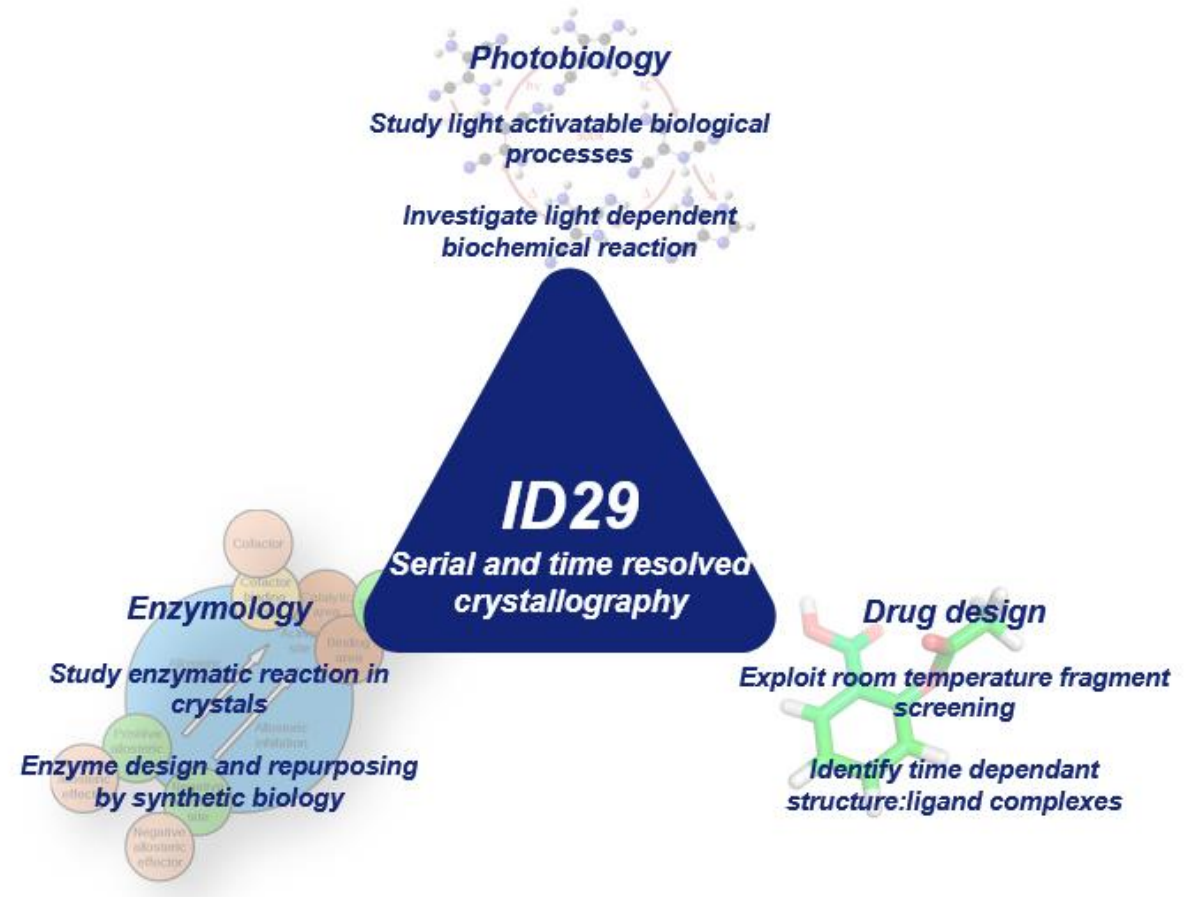


Schmidt, Advances in Condensed Matter Physics (2013)

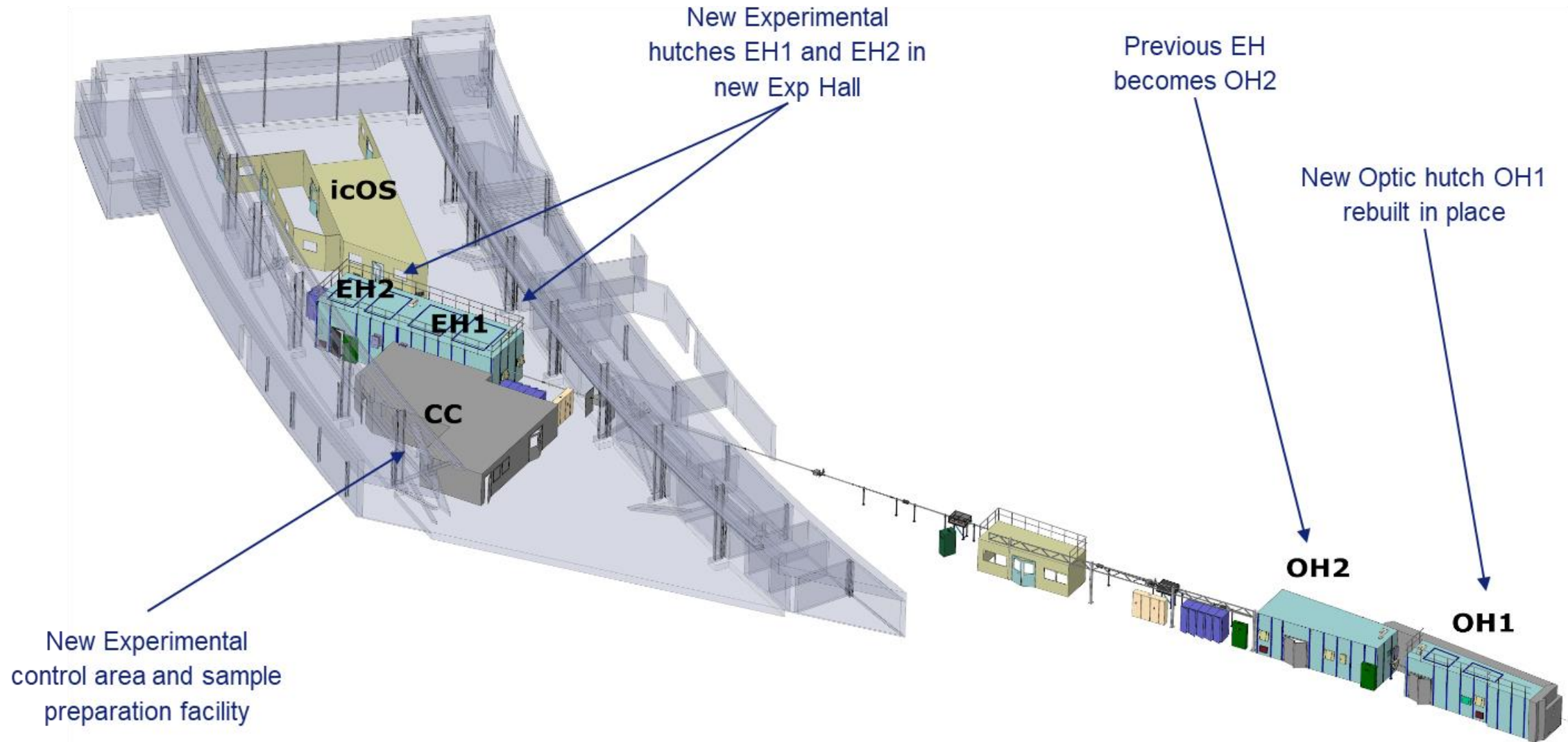


Mehrabi *et al.*, Nat. Meth. (2019)

- Room temperature serial crystallography experiment
- Extremely high flux with exposure time in  $\mu\text{s}$  range and high repetition rate
- Tunable over a large energy range
- Accurate control timing system to trigger events
- Adapt different sample environments and crystal delivery systems
- Optimize sample consumption
- Equipped with sample preparation laboratory and data analysis area

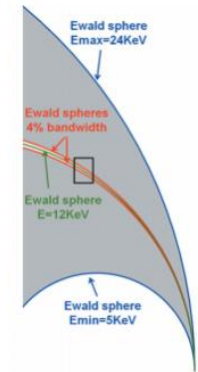
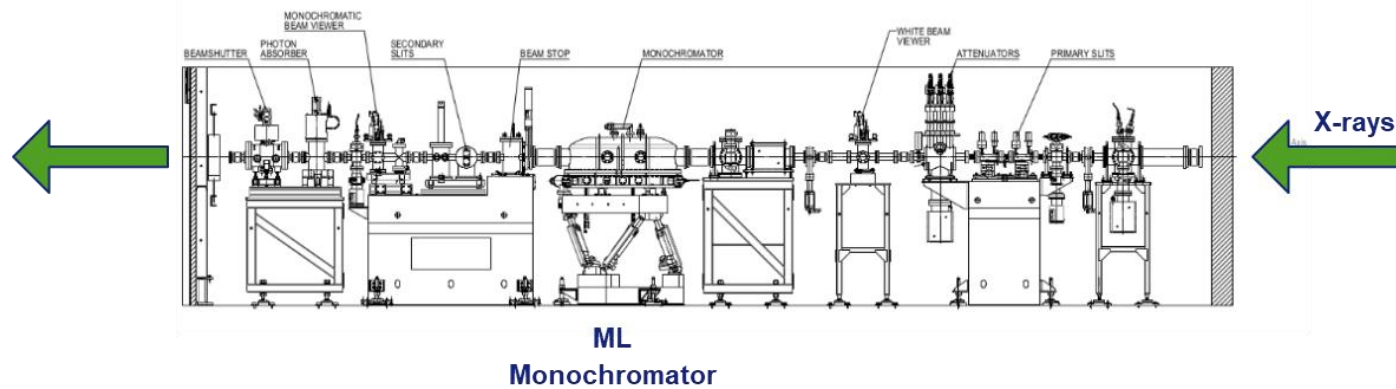


# BEAMLINE LAYOUT

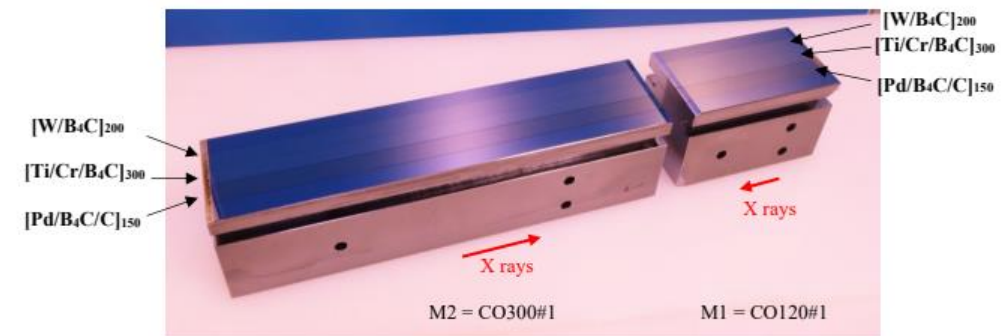




# OPTIC HUTCH 1 (OH1)

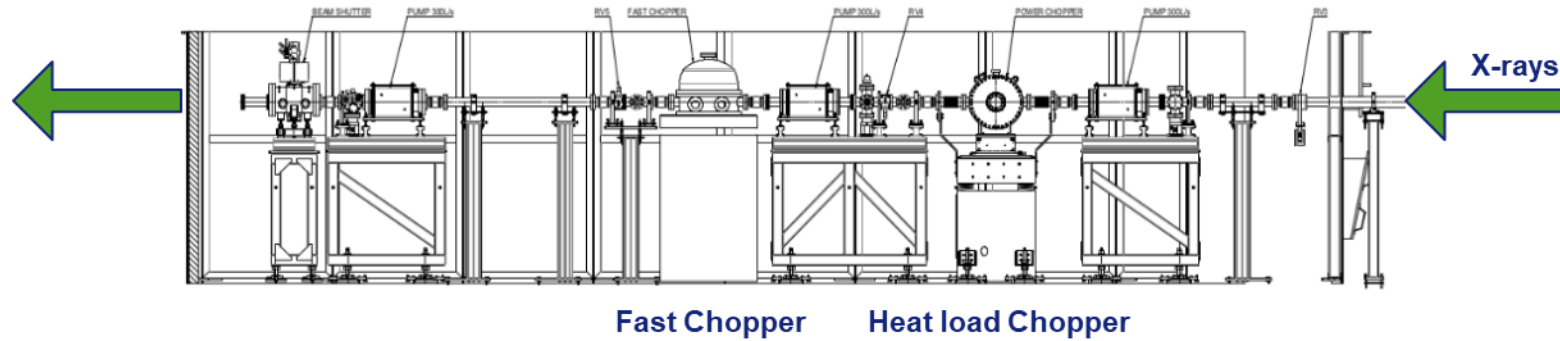


- A multilayer monochromator system was designed to increase bandwidth
- Adjust bandwidth by changing stripe
- The multilayer monochromator permits to deliver higher flux in larger bandwidth
- Three stripes are present to cover whole energy range 10-20 keV (+35 keV) with 0.4% and 1% bandwidth





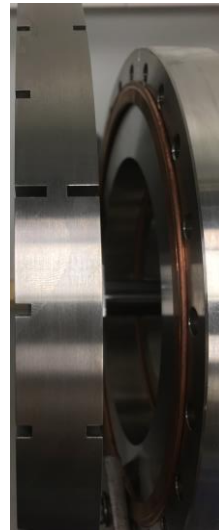
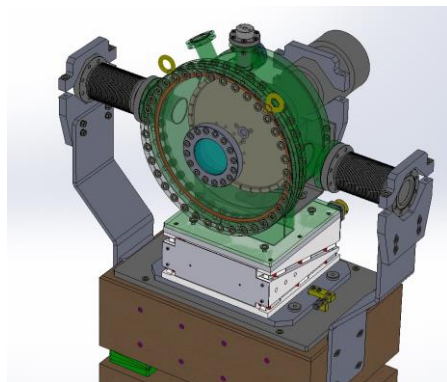
# OPTIC HUTCH 2 (OH2)



Fast chopper



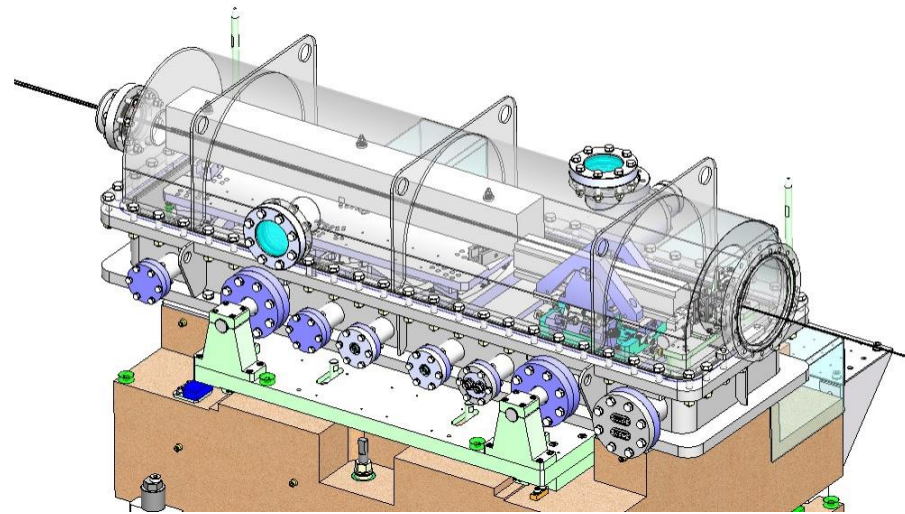
Heat load chopper



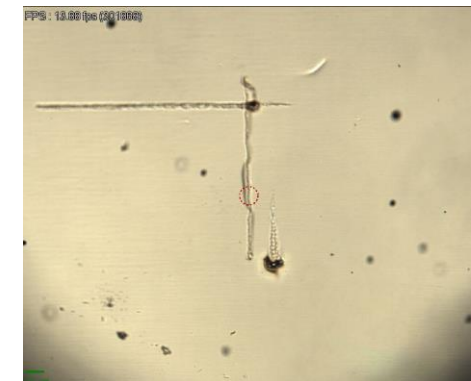
- A double chopper system removes heat load and defines short exposure times
- Heat load chopper generates a 90  $\mu\text{s}$  pulse at different frequencies: 925, 462.5, 231.25 & 115.625 Hz
- Fast chopper generates pulses of 10, 20 & 30  $\mu\text{s}$



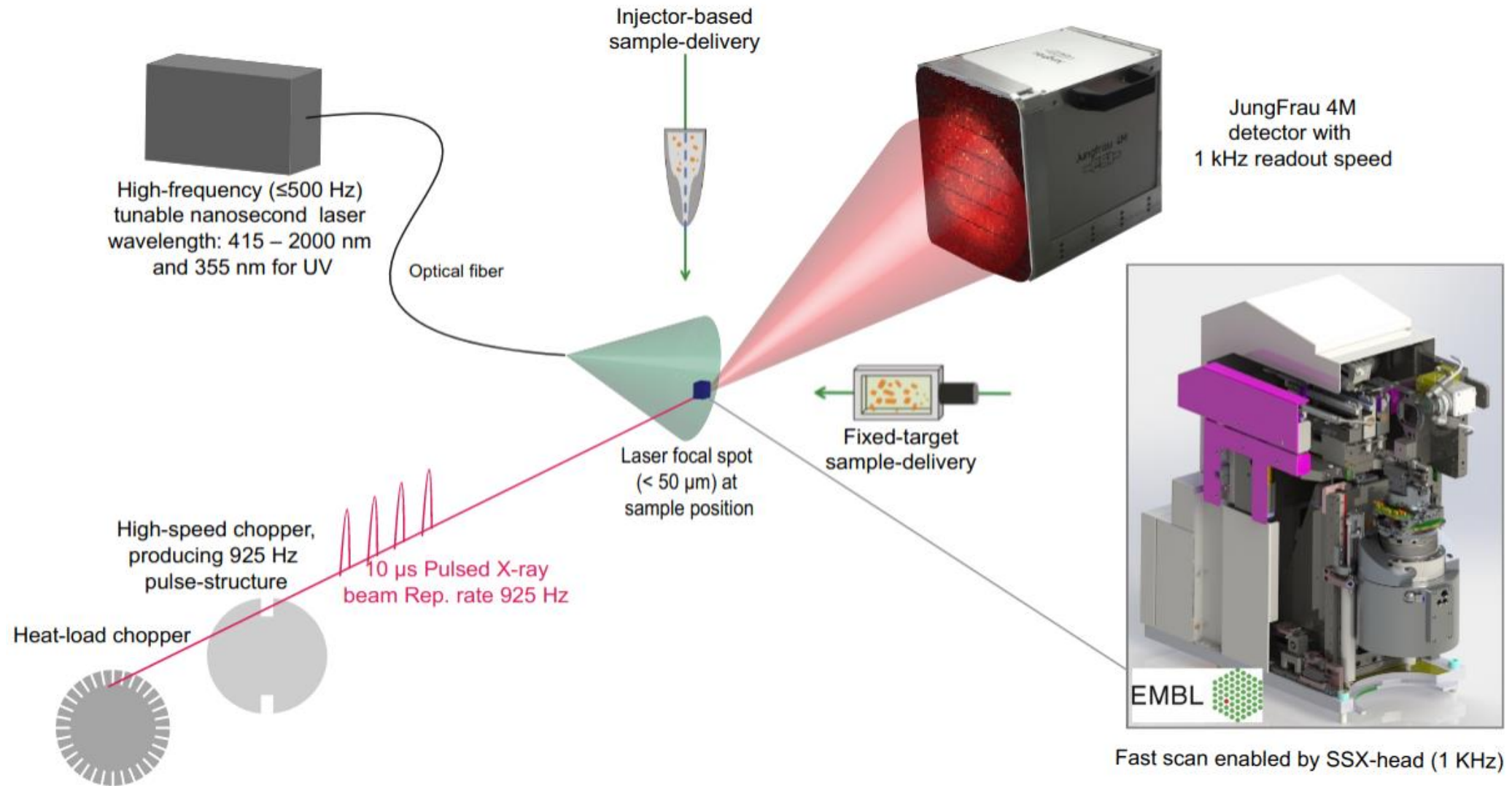
# EXPERIMENTAL HUTCH



- Sample at **107 m** from source
- Elliptical KB mirrors to focus the beam
- Measured beamsize  $\sim 2 \times 4 \mu\text{m}$  (VxH)
- Beam resizing by tuning incident angle
- Flux at the sample position of  $\sim 10^{15}$  ph/sec

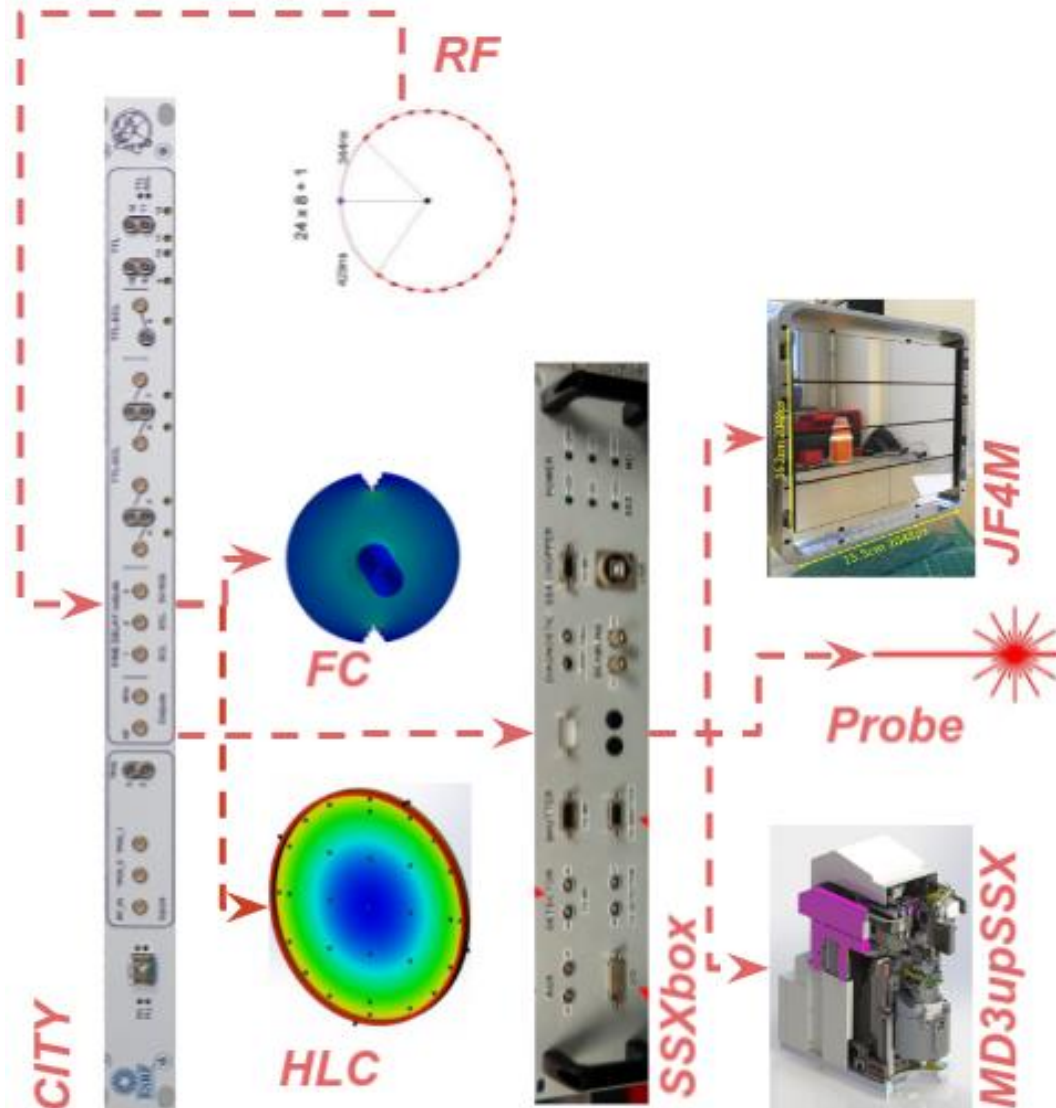


# EXPERIMENTAL SET-UP





# EXPERIMENTAL SET-UP

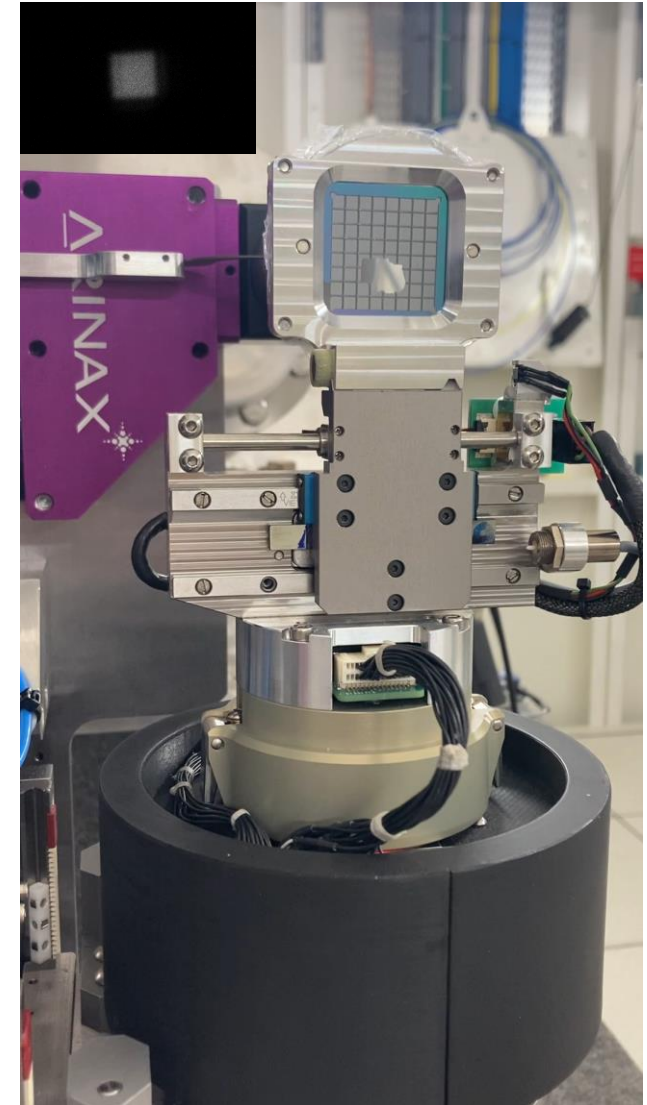
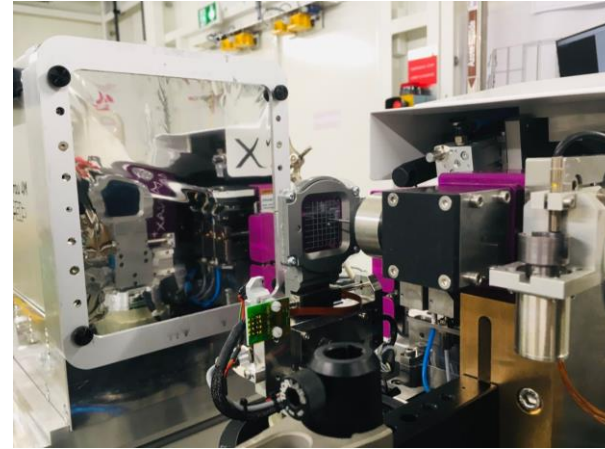
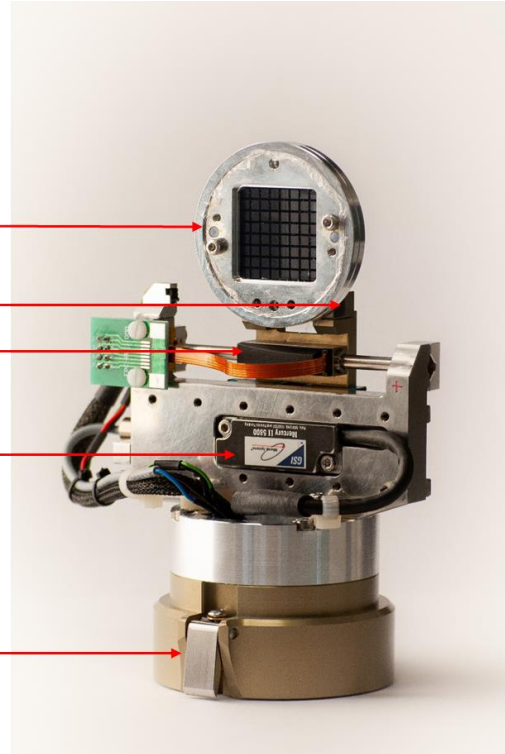


- A new developed timing system synchronises every step of the acquisition with the radiofrequency of the storage ring
- CITY and SSXbox are the two hearts of the system
- Heatload and Fast chopper are synchronised
- SSXbox propagate the clock to the data acquisition devices, MD3upSSX, X-ray detector and additional triggerings
- X-ray pulses of tunable length from 10 to 90 microseconds
- Synchronous triggering of external probes



# FIXED-TARGET

- “Standard” chip support
- Magnetic mount
- Linear brushless motor
- Linear encoder with 5nm resolution High precision scale
- Standard MD head mount (Quicklock)



- In Fixed-Target, crystals are captured in apertures of a silicon chip
- Scan speed: up to **125 mm/s**
- Desired precision at meeting point  $< 1\mu\text{m}$
- Run synchronously with the chopper system

# FIXED-TARGET

The screenshot displays the MXCuBE3 control interface for the opid291 collecting beamline. The top navigation bar includes 'Samples', 'Data collection', 'Equipment', and 'System log'. The main control area shows the following parameters:

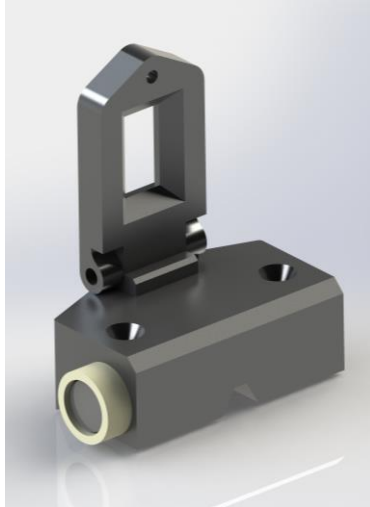
- Energy: 12.4000 KeV
- Resolution: - Å
- Transmission: 10.0 %
- Wavelength: 1.00 Å
- Detector: 10.1 mm
- Flux: 1.06e+10 ph/s

On the right side, there are status buttons for Detector (UNKNOWN), Sample Changer (READY), Capillary (IN), Fast Shutter (CLOSED), Safety shutter (CLOSED), and Ring Current (-1). Below these are 'Run Queue' and 'Unmount' buttons, along with a 'Settings' dropdown.

The central video feed shows a dark field with a bright spot on the right and a green dashed circle centered on the left. The left sidebar contains controls for 'Beam size' (set to 50), 'Chip (Diamond Chip)' (with a 'Navigate' button), 'Omega' (270.00 degrees), 'X' (-0.256 mm), and 'Y' (-3.382 mm). Below these are 'Sample alignment' controls and a 'Show motors' dropdown.

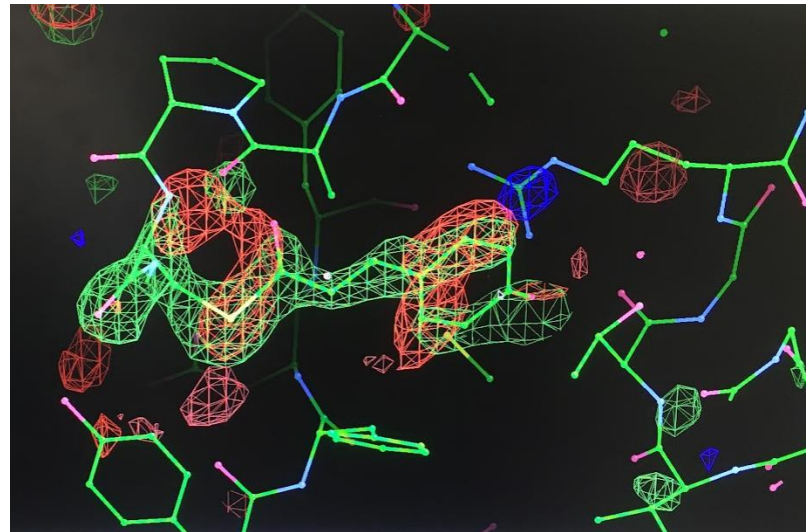
The bottom right panel shows a log of messages, with a recent entry: [17:12:01]: Mounting sample: lyso\_chip. The system tray at the bottom shows various application icons and a digital clock displaying 17:12:19.

# CHIPLESS CHIP

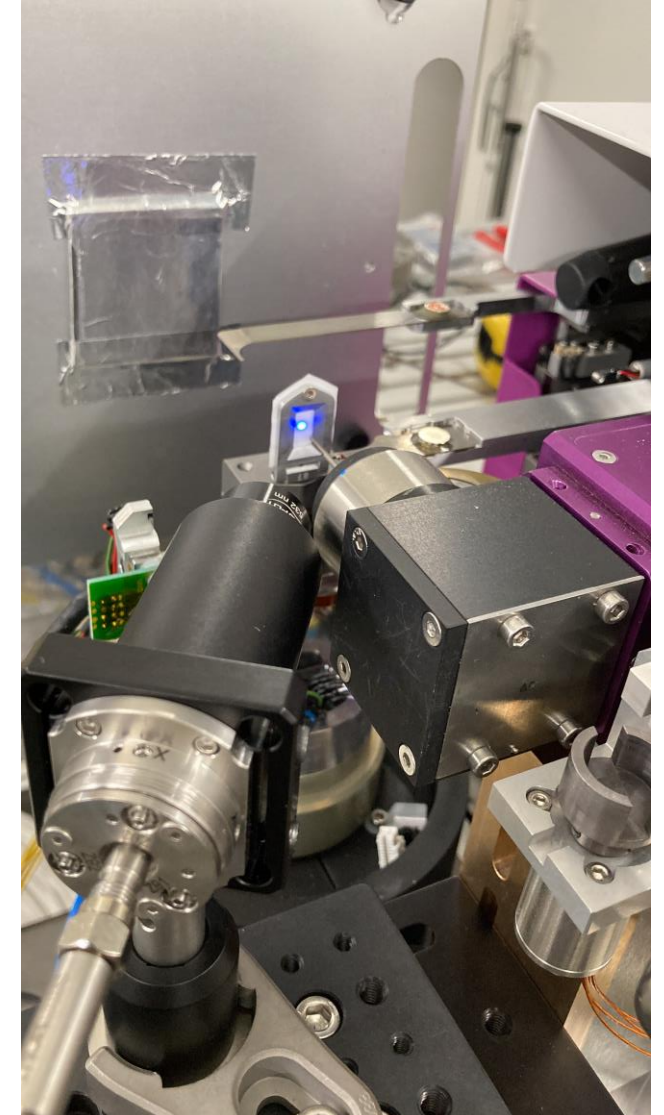


MPI-SOSchip

- Test the sample diffraction with only **3  $\mu\text{L}$**
- Can collect a complete dataset depending on crystal density
- **8 min** to collect **80k images**
- **Steady-state studies**

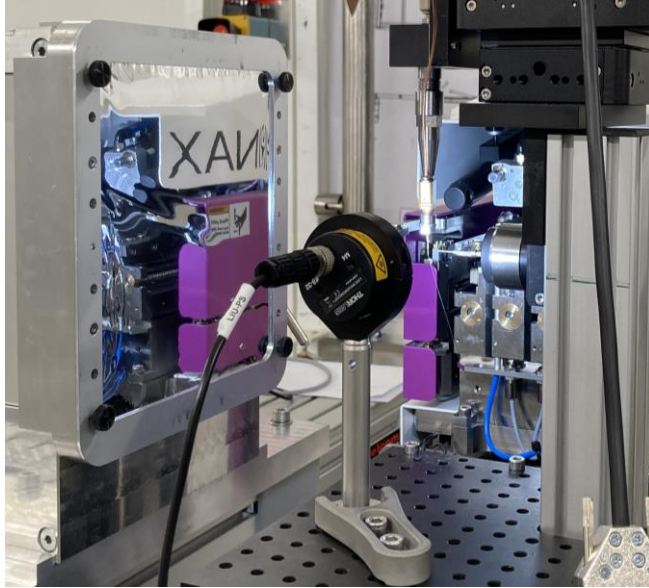


Schmidt group

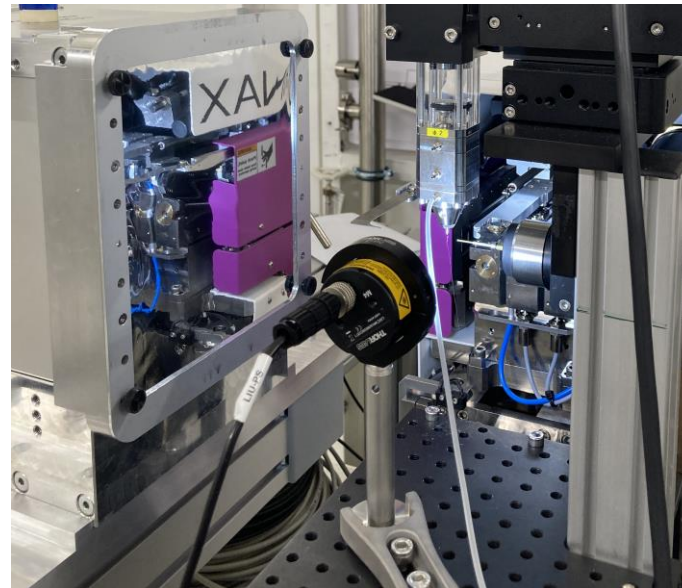




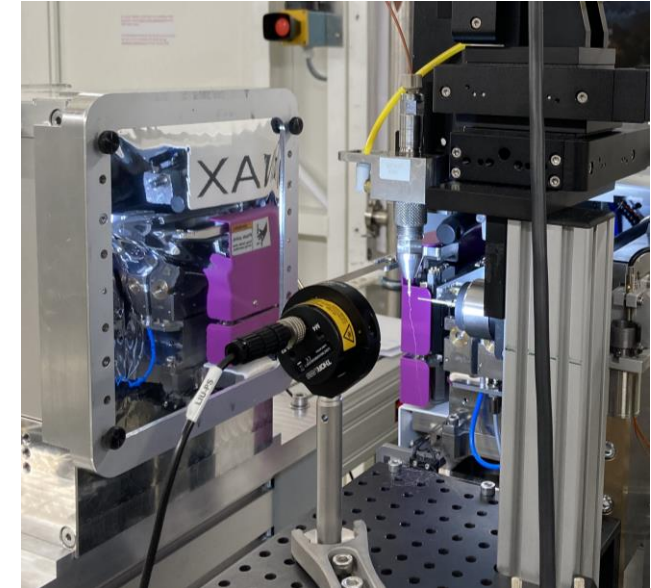
# HIGH-VISCOSITY INJECTOR



ASU injector



SACLA injector



MPI injector

Microcrystals are embedded in viscous media:

- LCP
- HEC (Hydrogel-based)
- Grease (Oil-based)



# TAPE-DRIVE



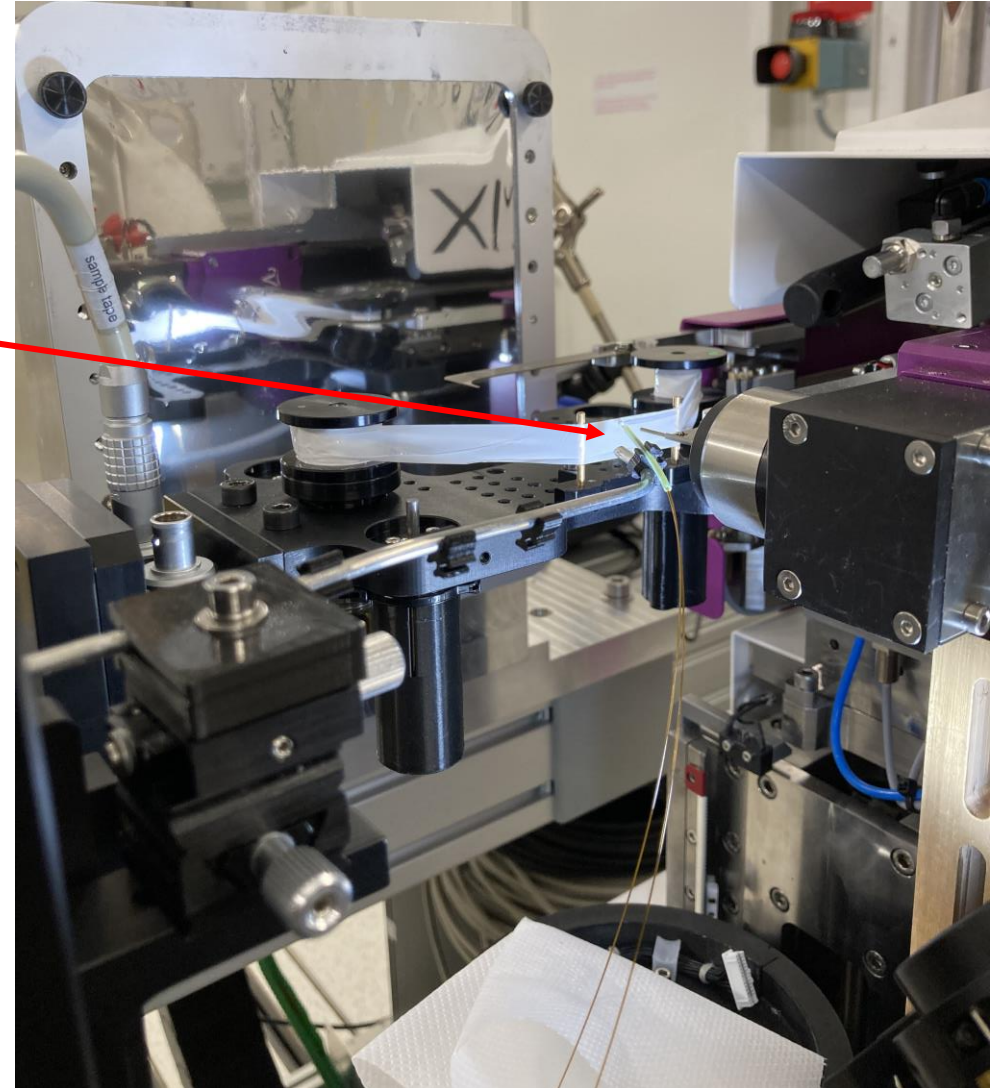
Lübeck University

3D-printed nozzle  
from CFEL

350  $\mu\text{m}$

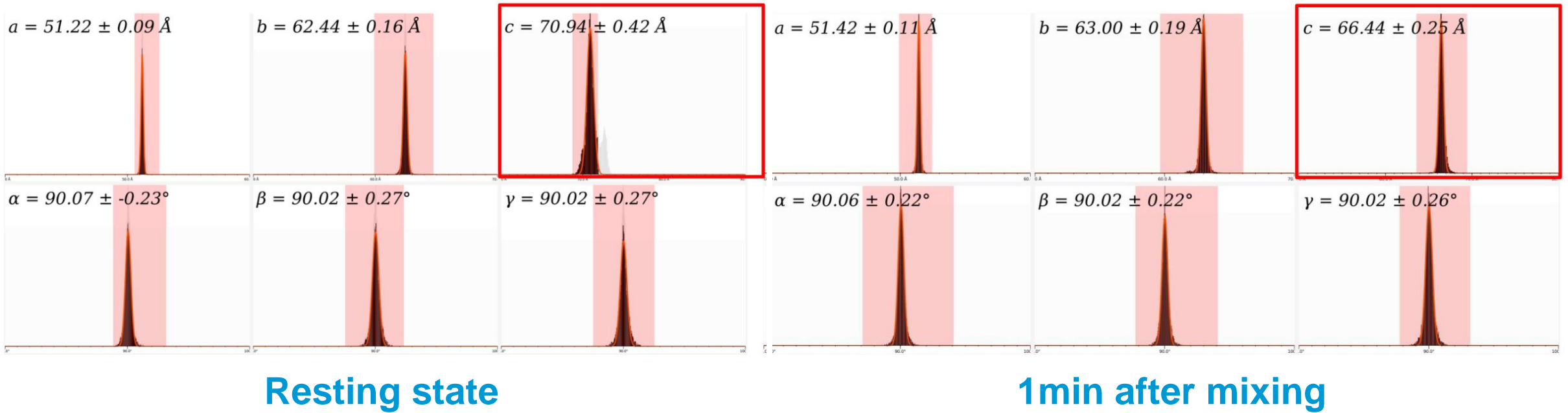


Henkel *et al.*, IUCrJ (2023)

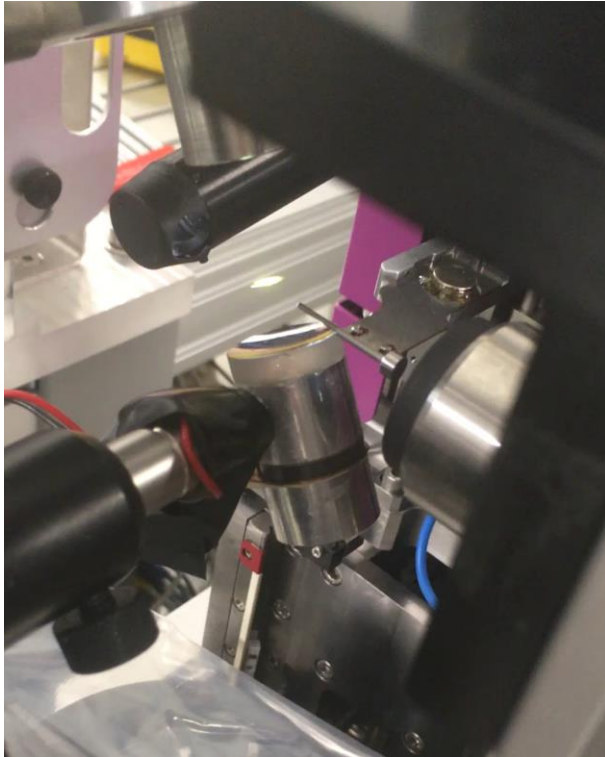


- Time-delay from **0.5 s to several minutes**
- Enzyme catalysis
- Ligand/inhibitor binding
- pH jump

T-Cer pH-jump showed a shrinkage on the c axis

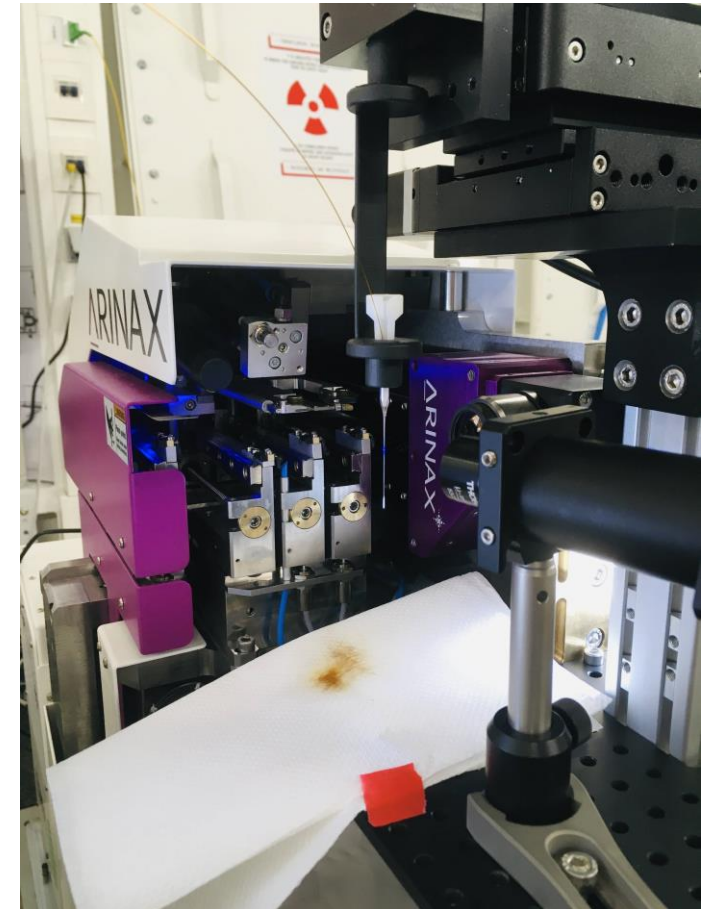
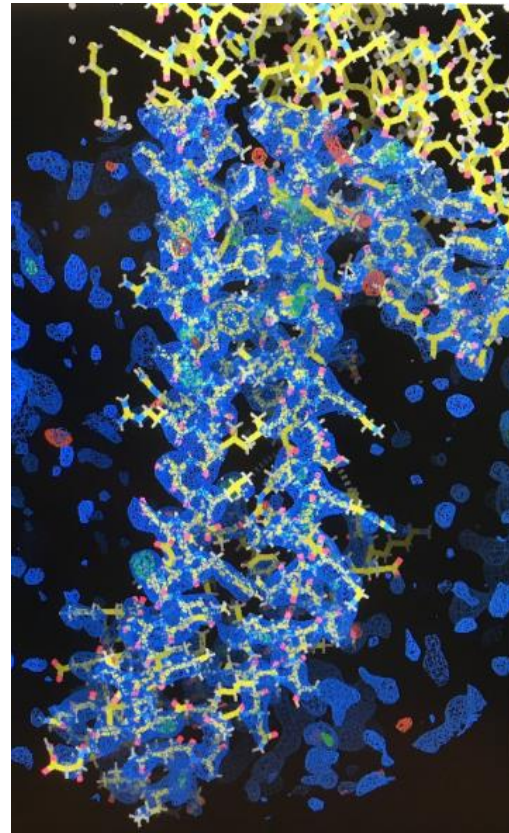


# OTHER SAMPLE DELIVERY METHODS



**PSI Acoustic  
Levigator**

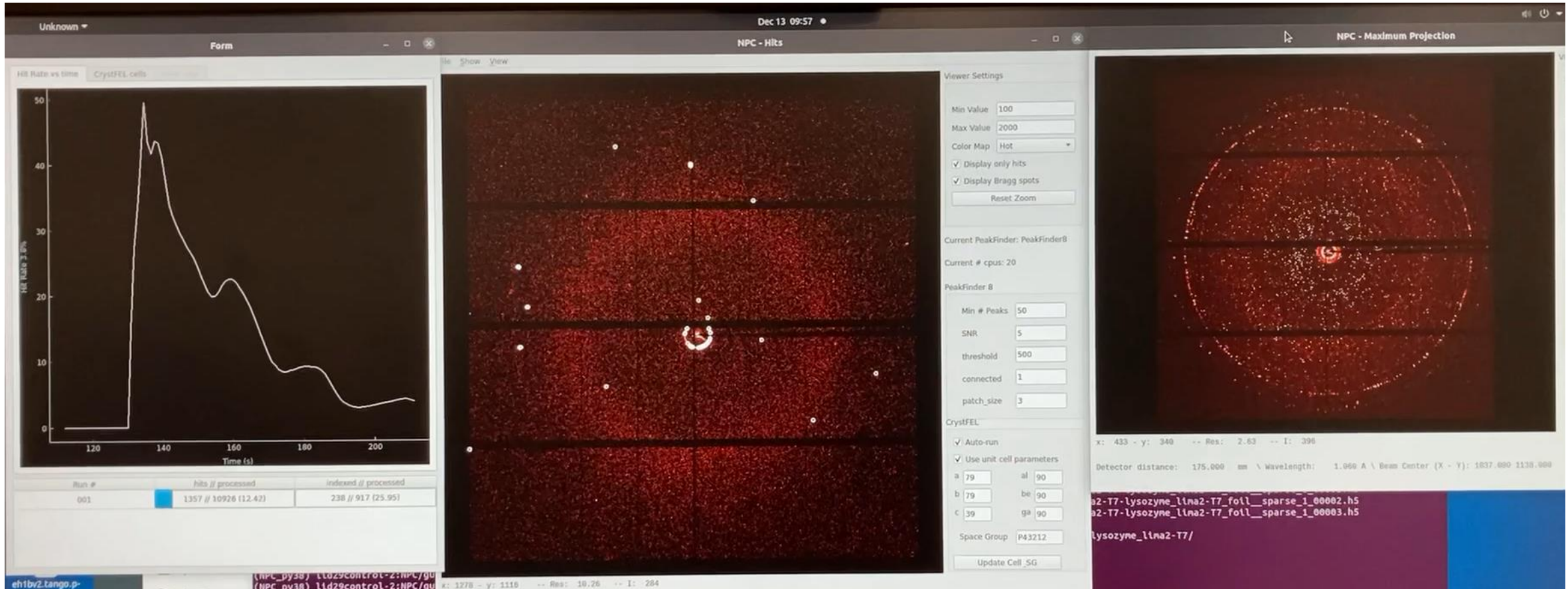
**First membrane protein  
collected on ID29**



**SerialX Gothenburg  
University**

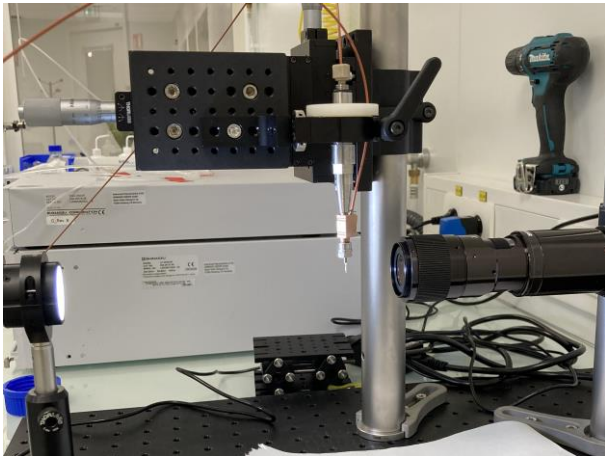
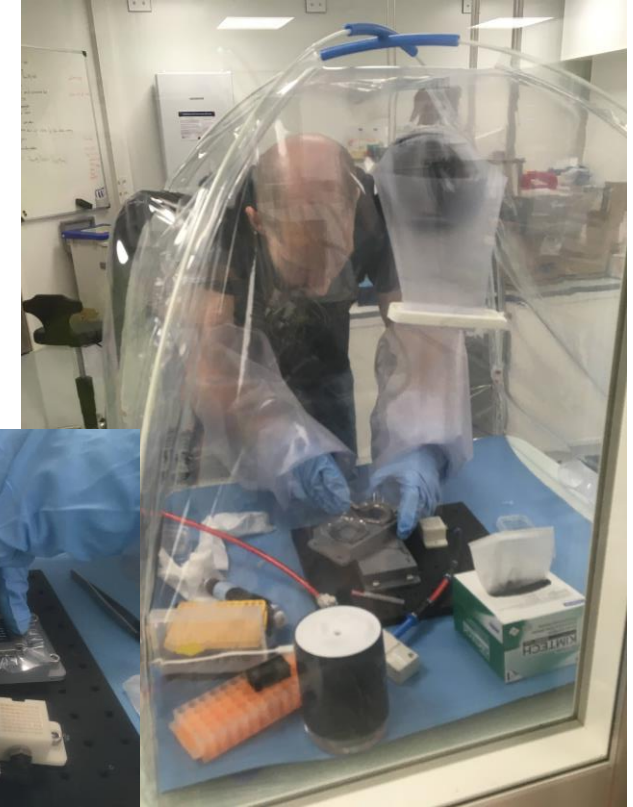


# NPLIVE – NEAR REAL TIME FEEDBACK ON PROCESSING WITH GPU

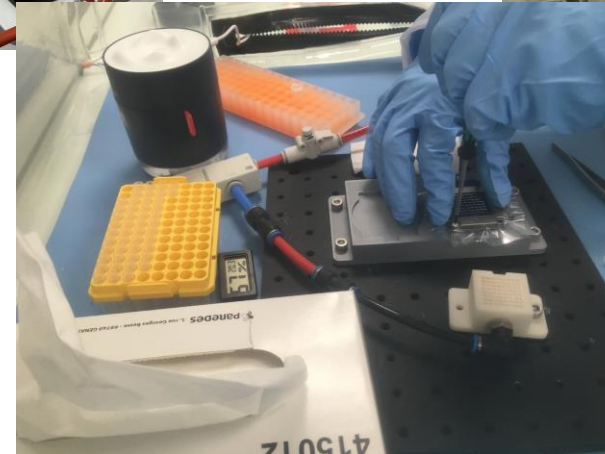
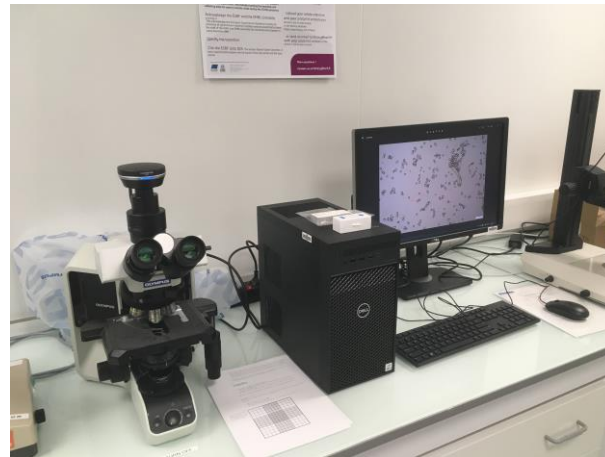




# SAMPLE PREPARATION LABORATORY



**HVE Offline Bench**



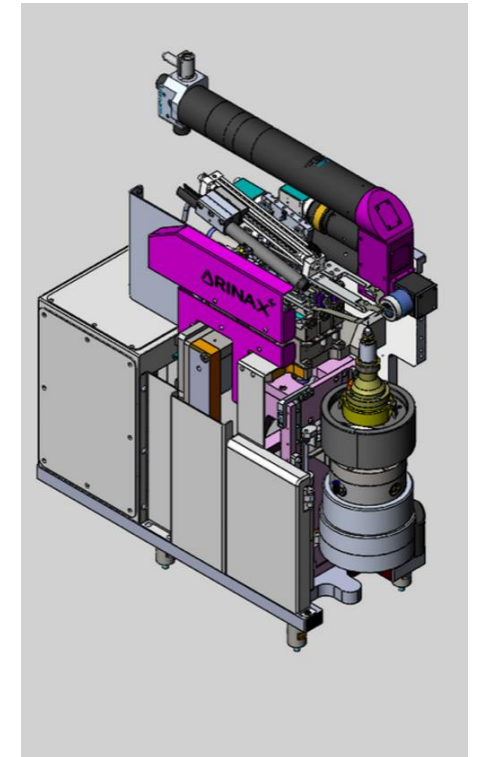
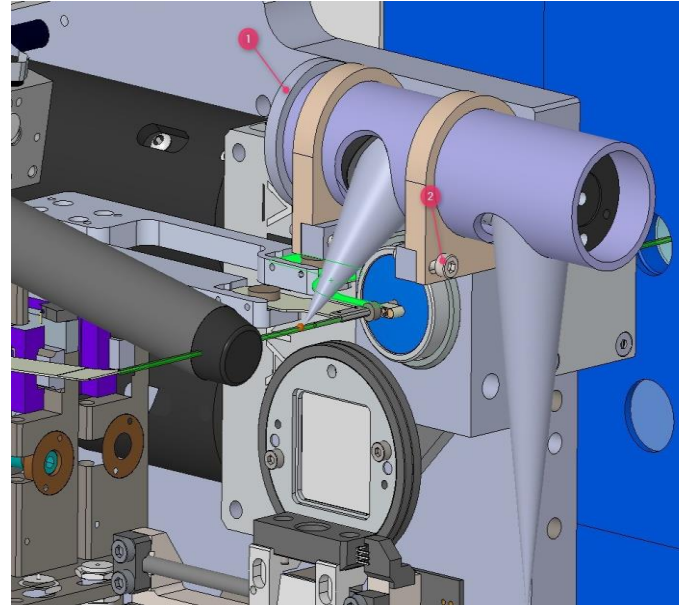
**Humidity Tent**

# NANOSECOND LASER INTEGRATION

- 400 - 2000 nm laser (+355 nm) at 500 hz, 10 mJ max pulse energy
- New development injection through OAV
- Synchronised with experiment clock

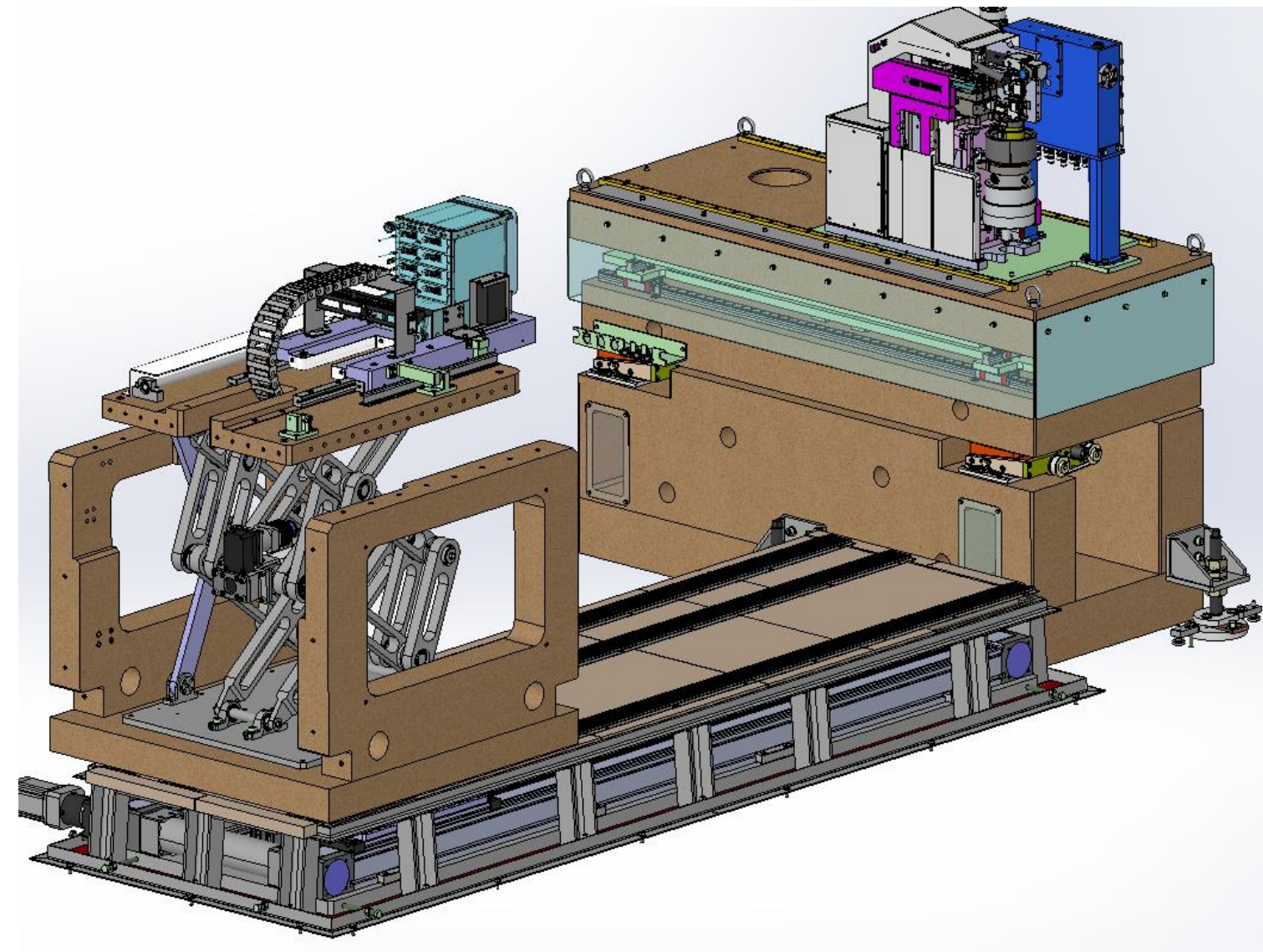
## Laser specification:

- wavelength range (400-2000 nm)
- pulse energy (~10–20 mJ)
- pulse duration (~5 ns)
- externally triggered
- repetition rate ~500 Hz minimum
- and portable with optical fibre



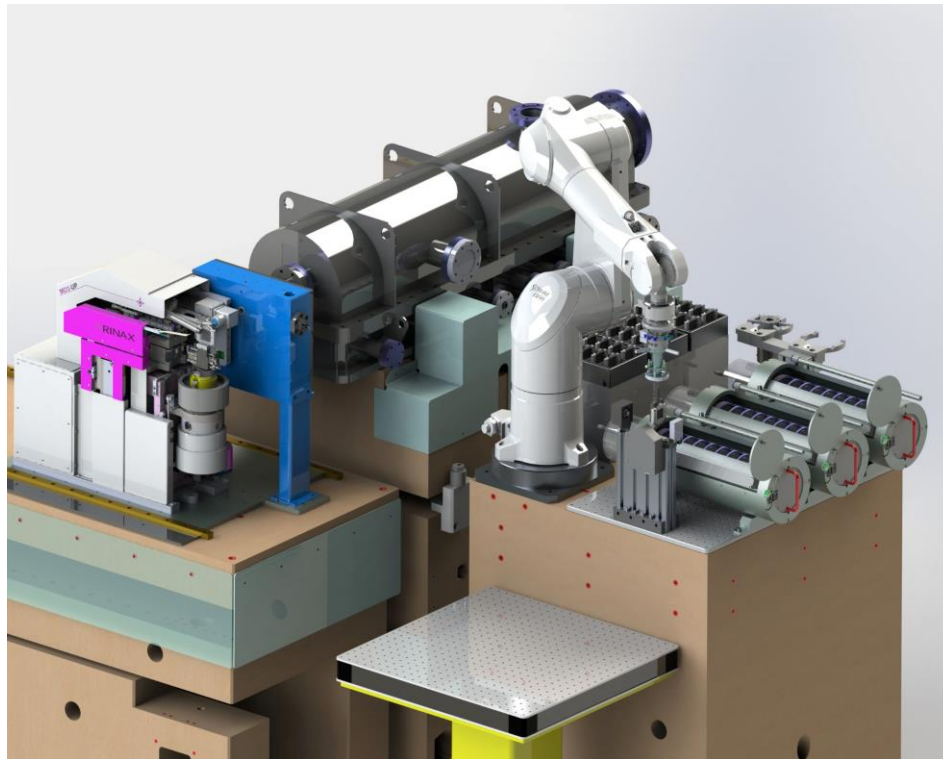


# DETECTOR TABLE



# FLEX SAMPLE-CHANGER FOR FIXED-TARGET

- Brand new FLEX gripper to mount fixed target chips
- Dedicated humidity controlled hotel for storing Si-chips
- Standardization in sample mounting





**Photon flux: ~  $10^{15}$  ph/sec**

**Pulse length: 90, 30, 20 & 10  $\mu$ s**

**Repetition rate: 925, 462.5, 231.25 & 115.625 Hz**

**Energy: 10-20 kEv (+35 kEv)**

**Beam size: ~ 2 x 4  $\mu$ m (VxH)**

**11 BAG proposals world wide (Operation from Sept, 2022)**

**More than 600Tb collected in 6 months**

**Daniele de Sanctis**  
**Shibom Basu**  
**Samuel Rose**

### ESRF Structural Biology Group

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Hugo Caserotto  
Nicolas Coquelle  
Fabien Dobias  
David Flot  
Jonathan Gignes  
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**Users**