



Analysis of the effect of electro-optical parameters on neurotransmitter uncaging experiments on cerebellar cells in vitro.

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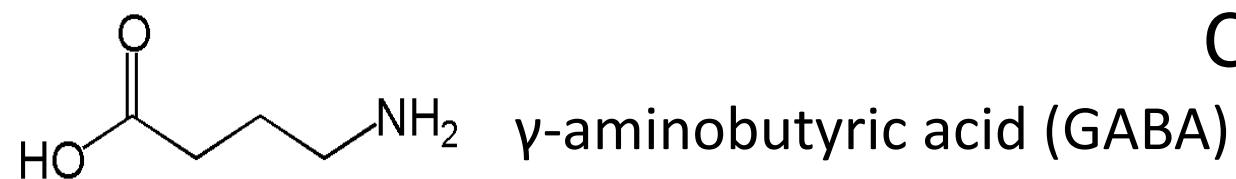
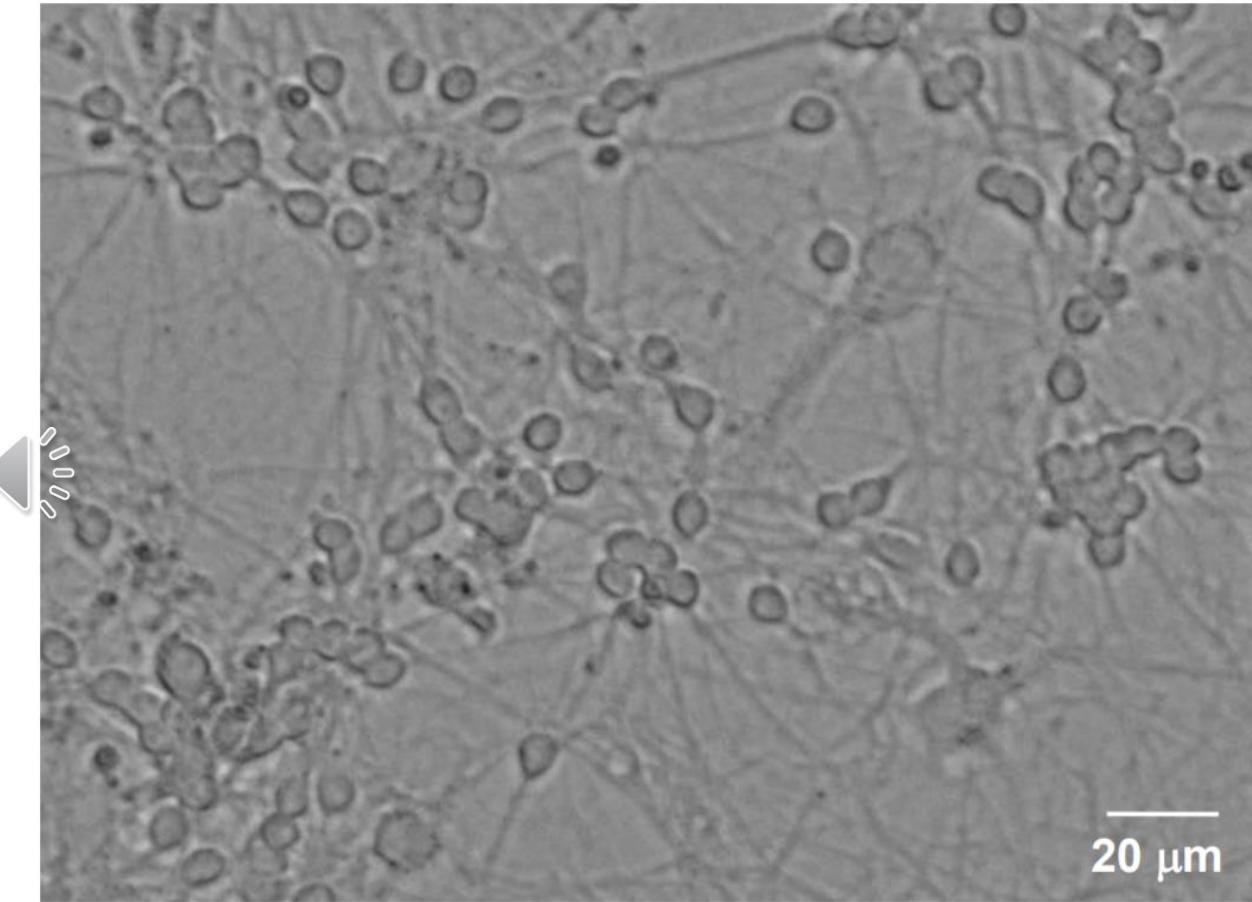
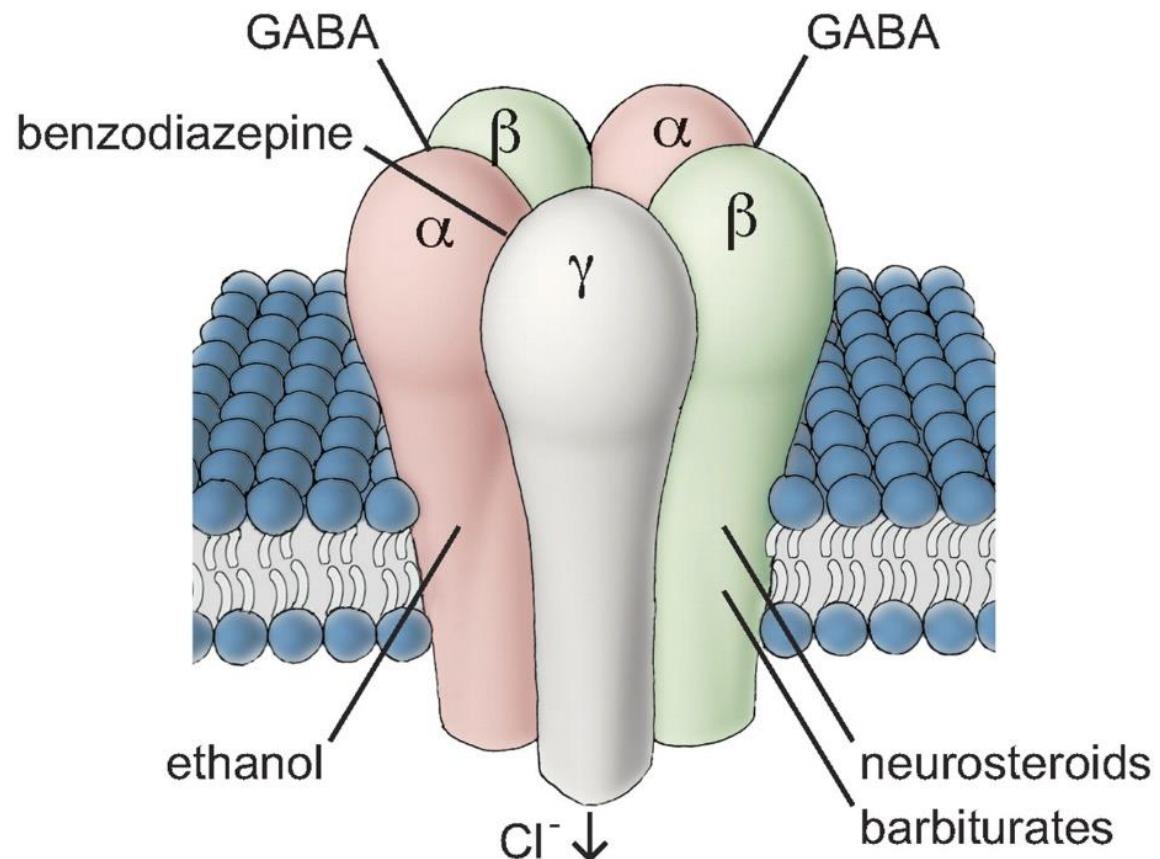
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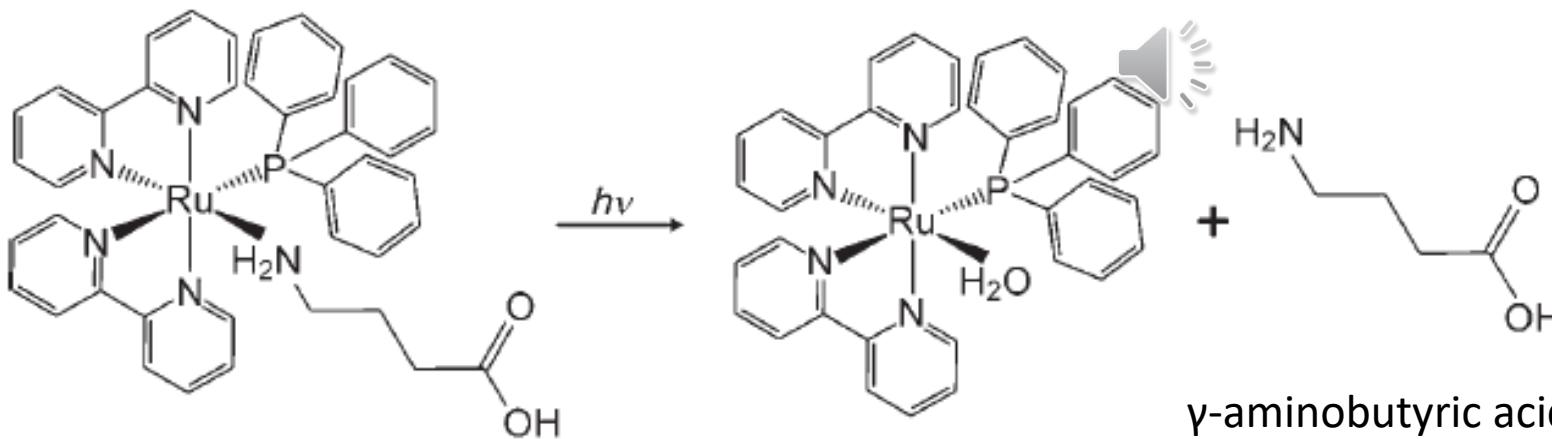
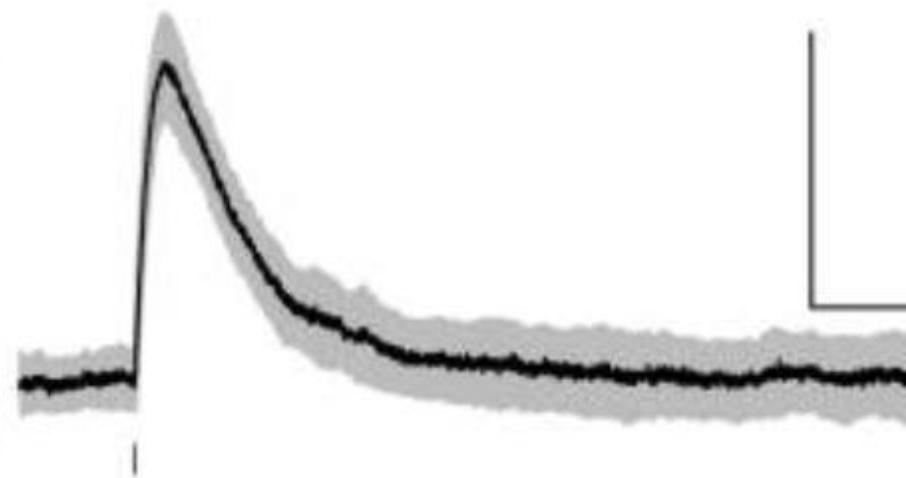
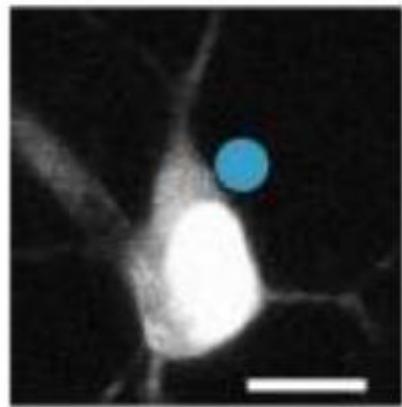
SIF – 106° Congresso Nazionale
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GABA_A receptor

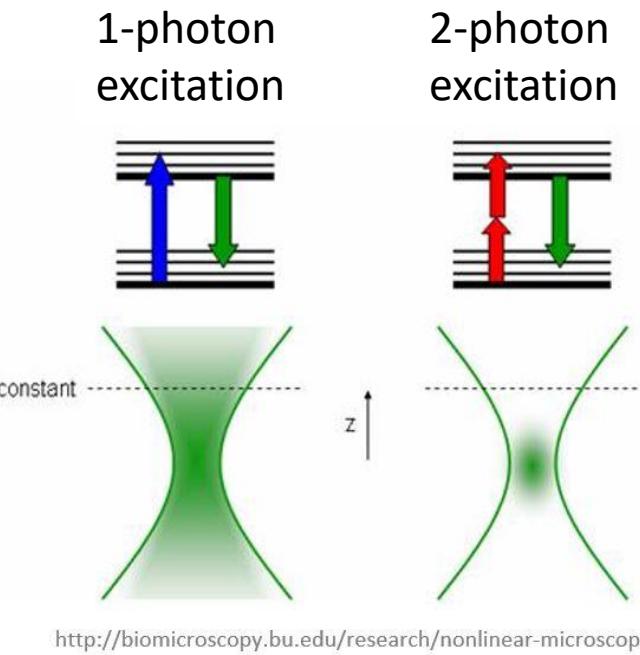


Cerebellar granule cells *in vitro*

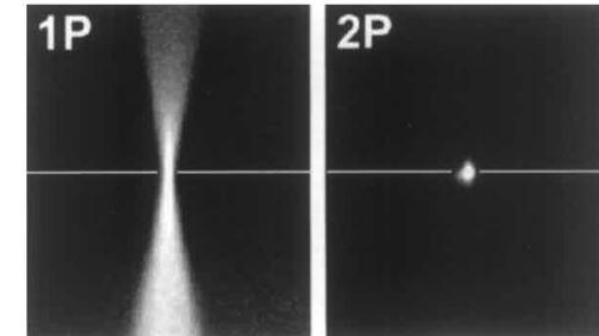
Neurotransmitter uncaging experiments



Caged neurotransmitter
RuBiGABA



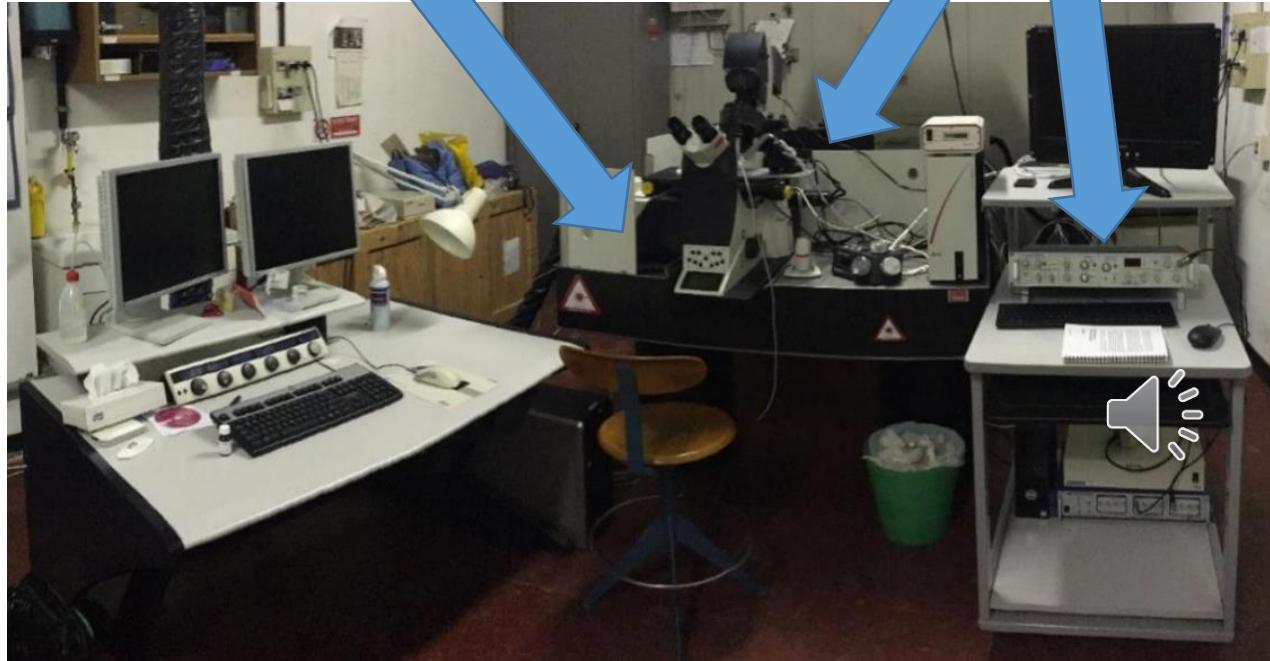
<http://biomicroscopy.bu.edu/research/nonlinear-microscopy>



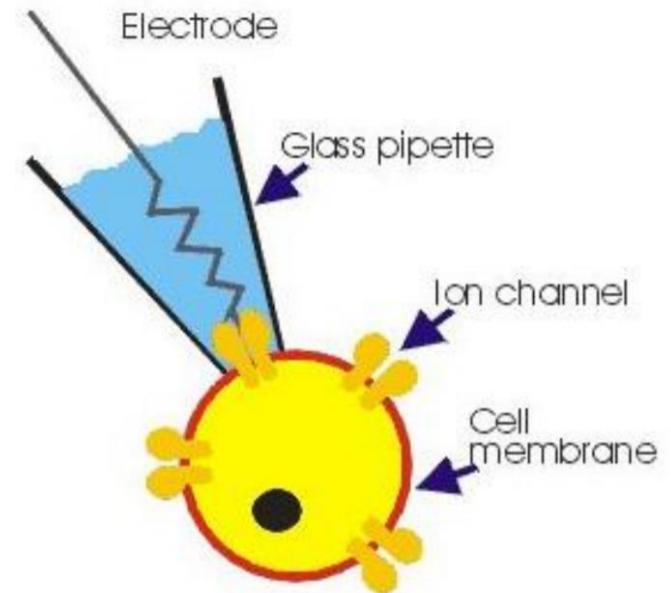
M. Rubart (2004) Two-Photon Microscopy of Cells and Tissue. Circulation Research 95:1154-1166

Experimental set-up

Confocal microscope



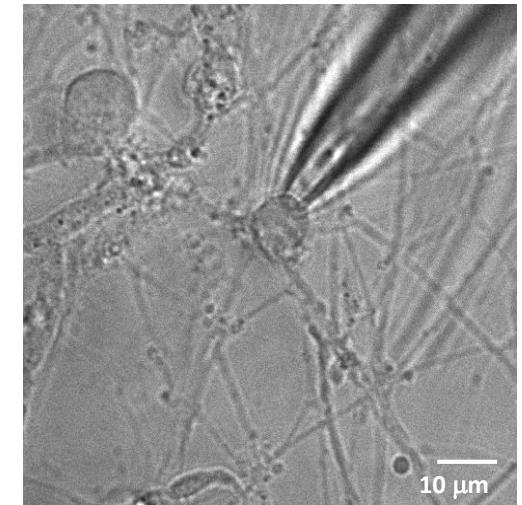
Set-up for patch-clamp



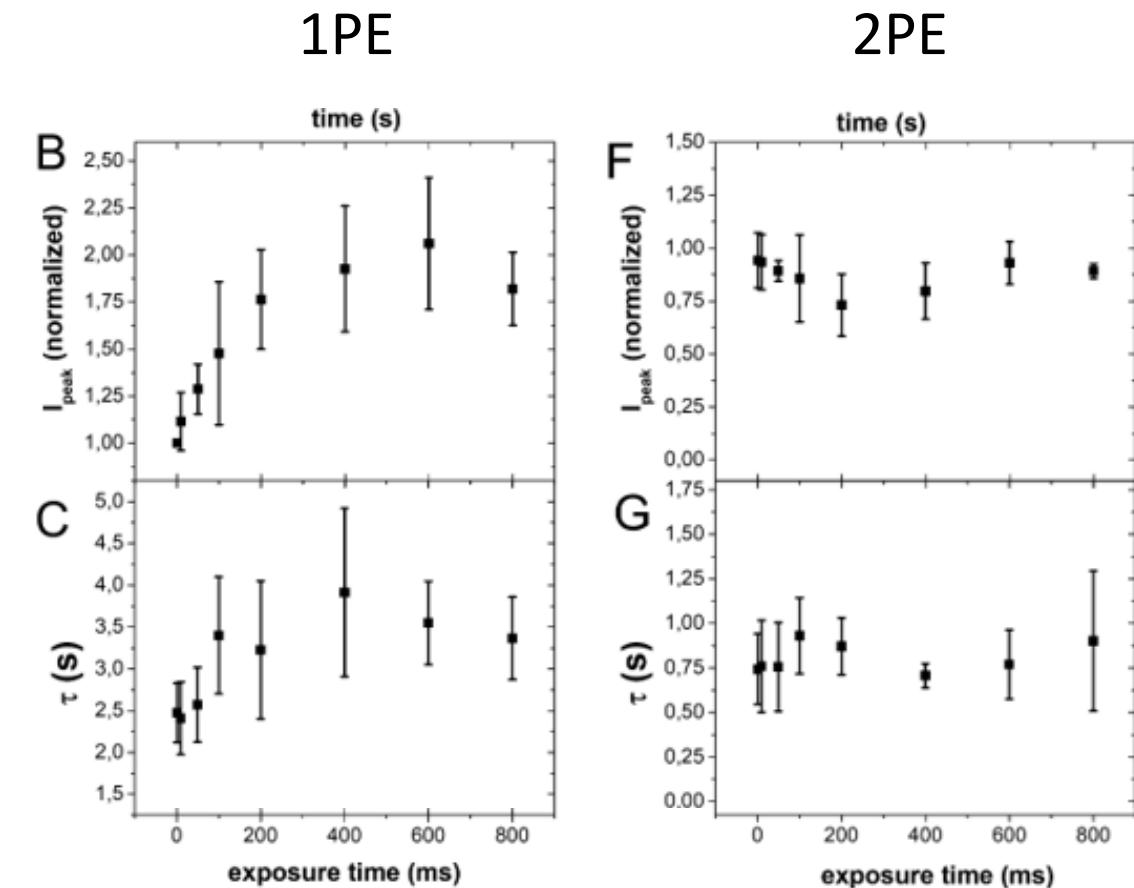
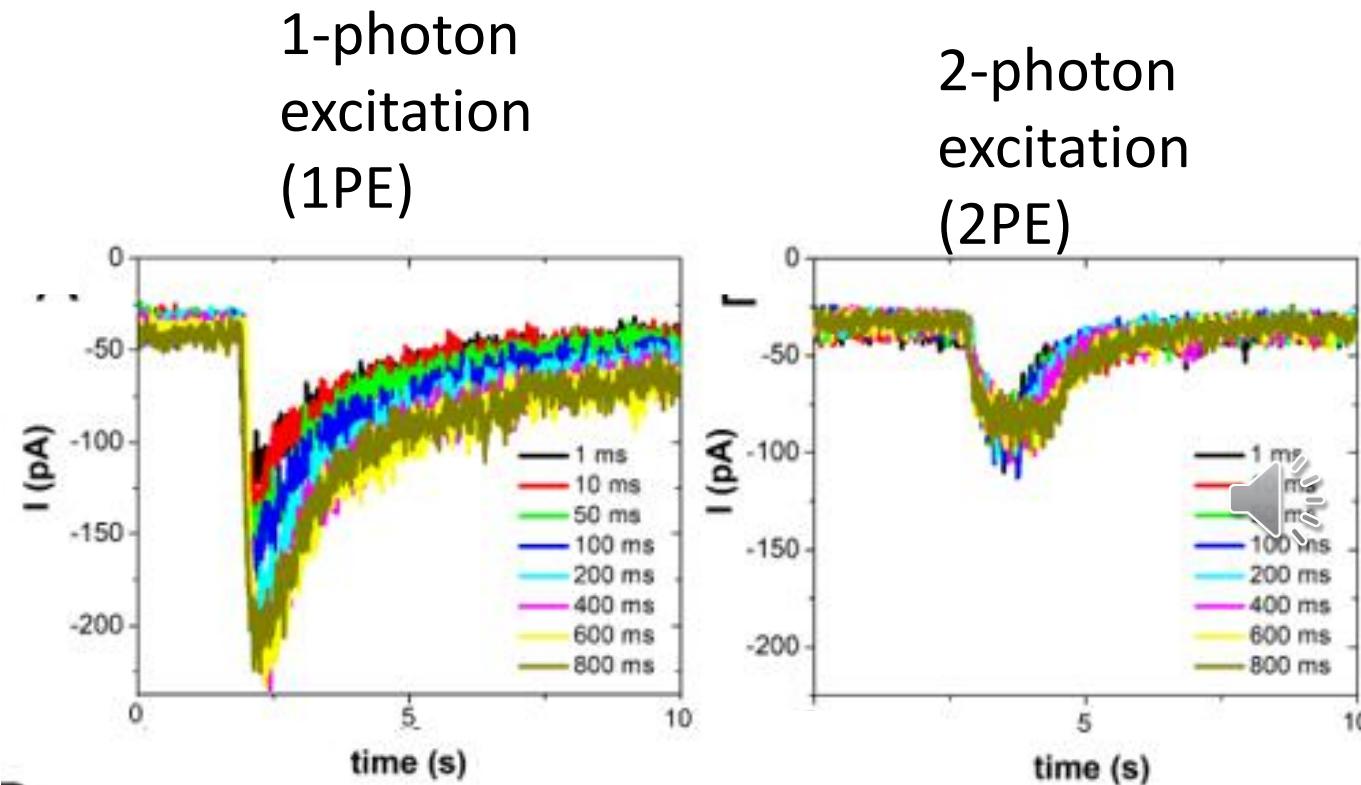
Patch-clamp technique in whole-cell configuration

Investigation of these electro-optical parameters:

laser power
exposure time
distance from cell in X-Y plane
Z-axis distance from the cell
photoactivation method.

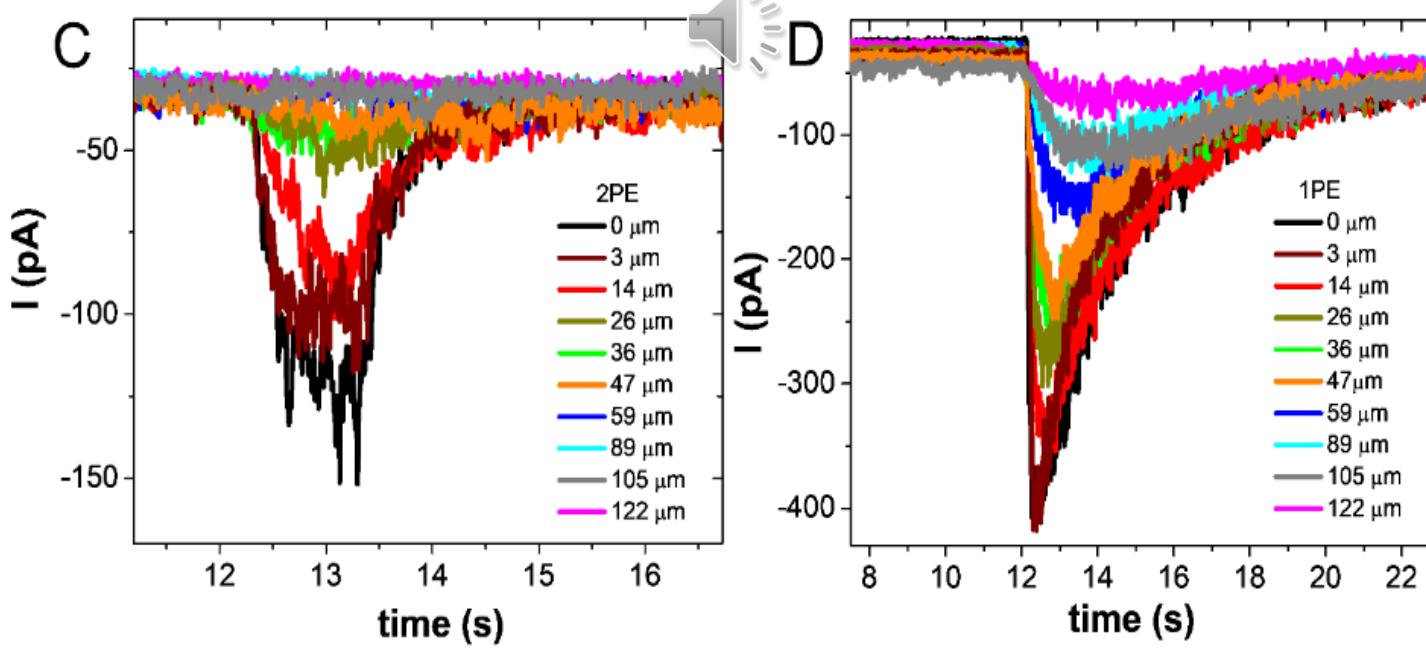
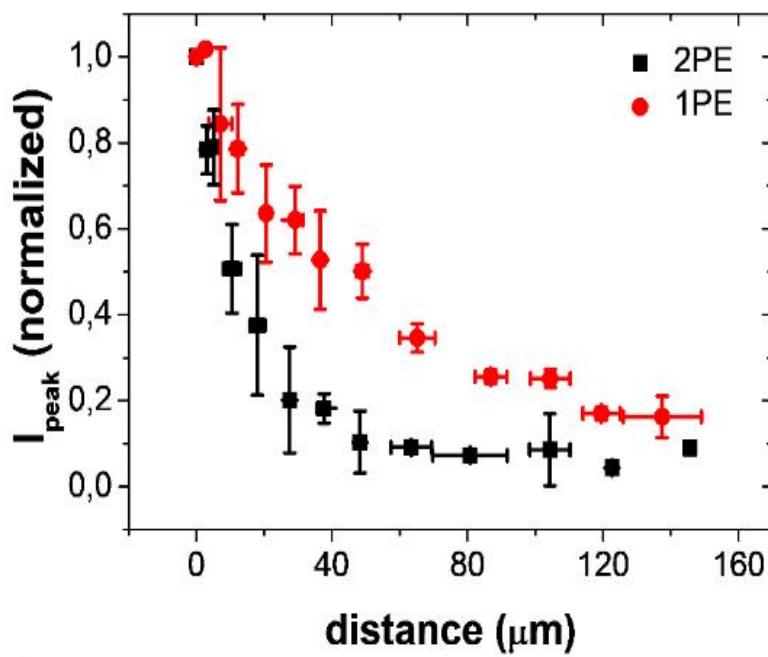
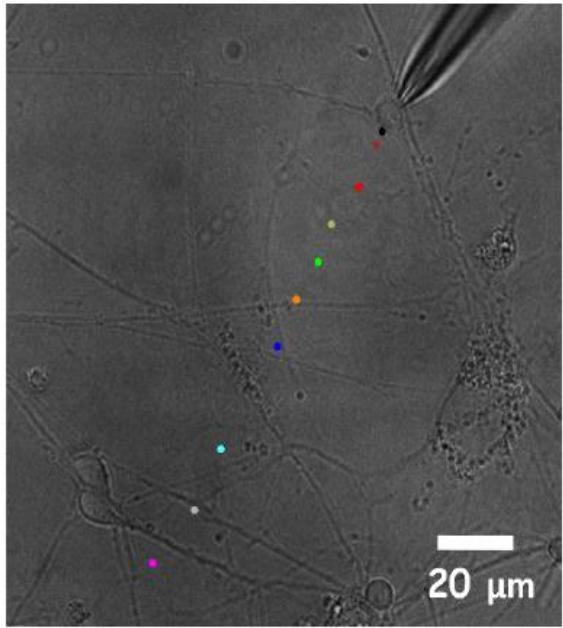


Current versus exposure time analysis

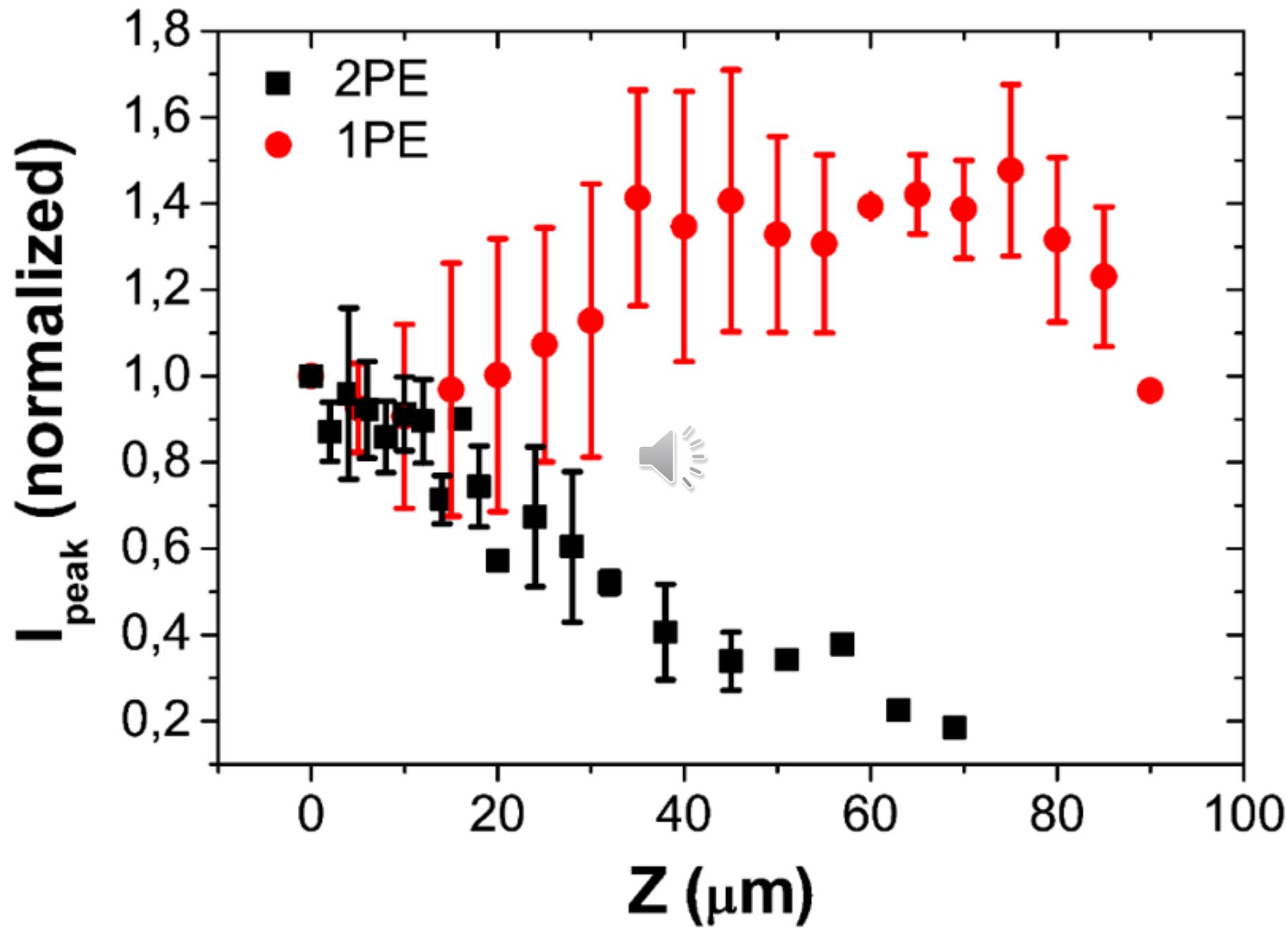


(10 μ M uncaged RuBi-GABA V=-80 mV)

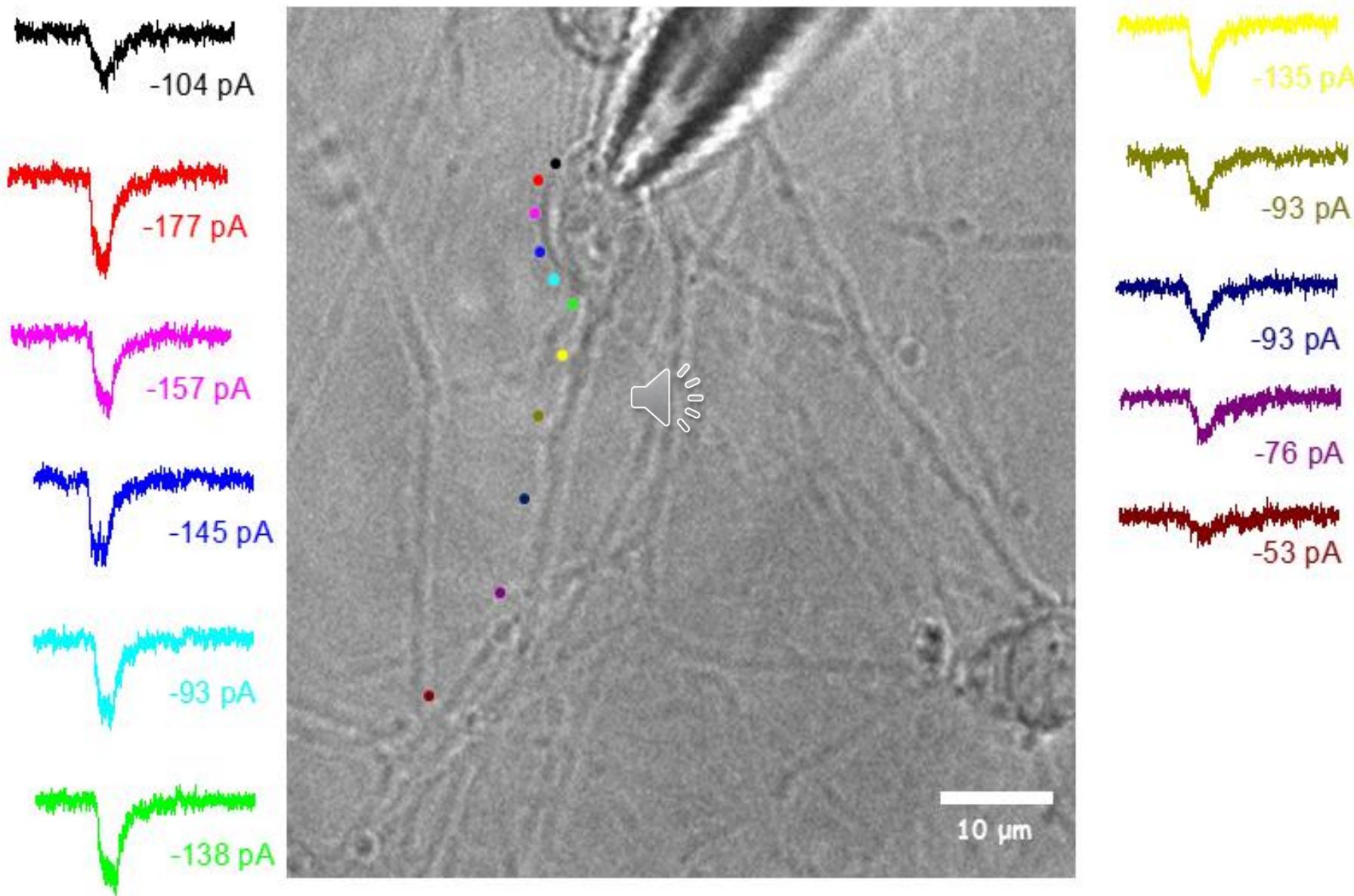
X-Y plane distance from the cell



1PE (458 nm, 100 ms,
9.36 μW)
2PE (750 nm, 100 ms,
77.3 mW)
10 μM RuBi-GABA
at $V=-80$ mV



RuBi-GABA uncaging to map GABAergic receptors distribution



Conclusions

The combination of electro-optical techniques allows electrophysiologically examining a confined area of the cells, enabling researchers to define detailed maps on kinetics, physiology, and pharmacology of specific molecules.



Comparison of 1PE and 2PE based uncaging processes allows refining protocols for experiments related to specific mechanisms such as to neuronal cell communications.